

Chemical risk assessment: How well do *in vitro* and *in silico* data predict the *in vivo* situation?



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New Approach Methods (NAMs)

- New approach methods (NAMs) are important for screening thousands of untested chemicals.
- NAMs can be used to predict point of departure (POD) using *in vitro* and *in silico* methods
- How do NAM-based PODs compare with *in vivo* PODs?

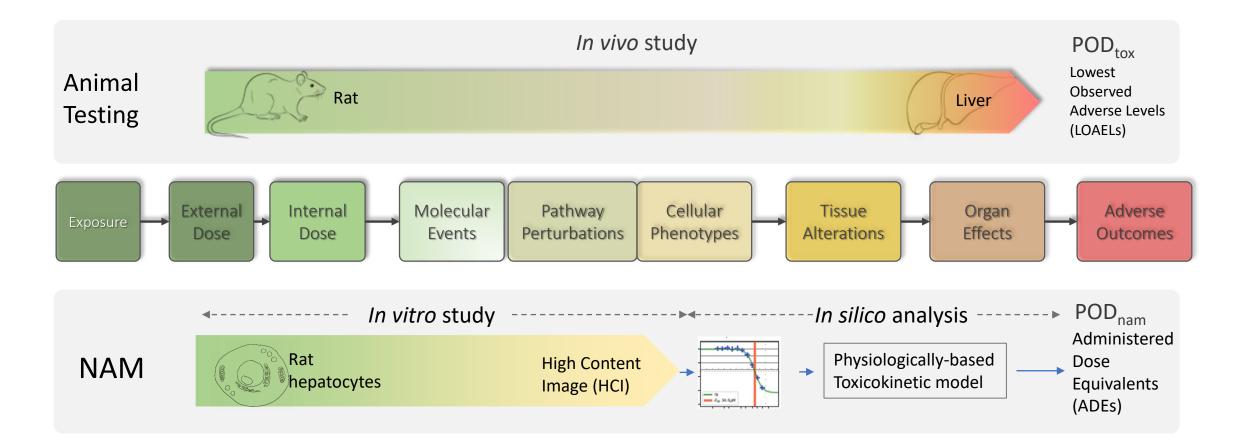
NAM = in vitro | in silico

Any technology, methodology, approach, or combination of methods that can provide information about chemical hazard and risk assessment without using whole animals.



Kavlock, et al 2018

A NAM for Repeat-Dose Rat Liver Toxicity Testing



Approach: Retrospectively compare POD from NAMs (POD_{nam}) with POD from traditional toxicity testing (POD_{tox})

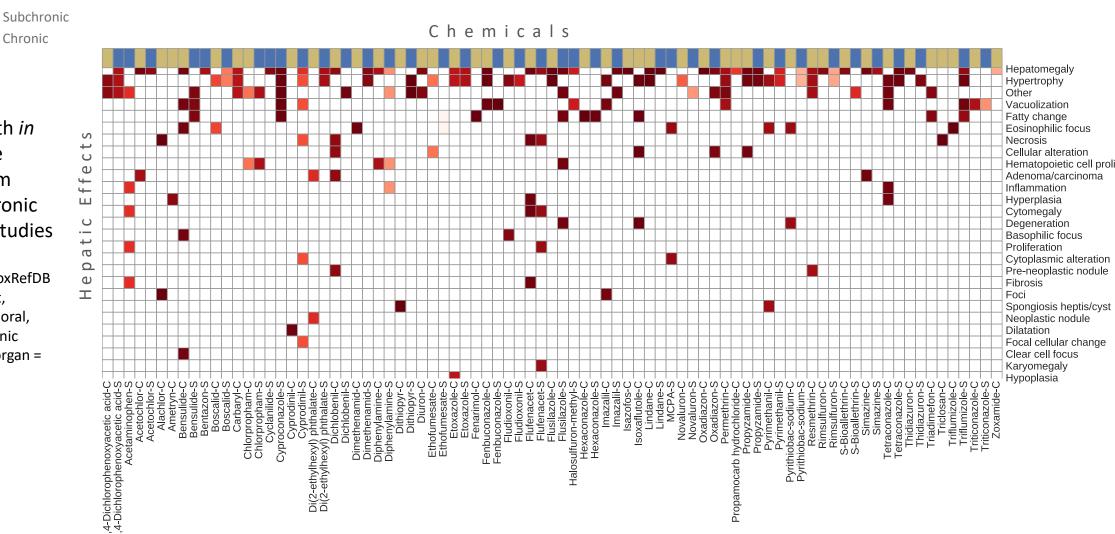
Repeat-Dose Toxicity Data POD_{tox} values for 51 chemicals





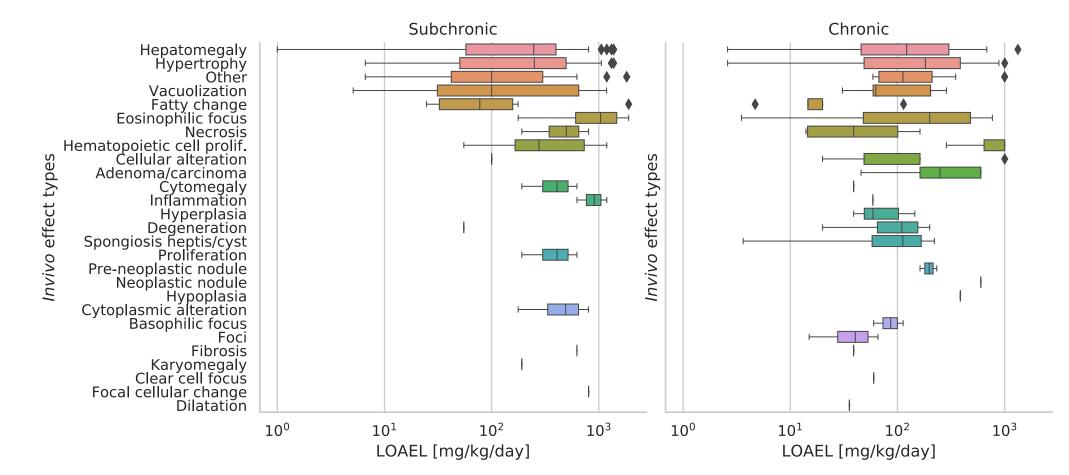
51 chemicals with *in vivo* repeat-dose toxicity data from guideline subchronic and/or chronic studies

Data obtained from ToxRefDB v2.0 using species=rat, administration route=oral, study type = (subchronic |chronic) and target organ = Liver



Distribution of POD_{tox} values

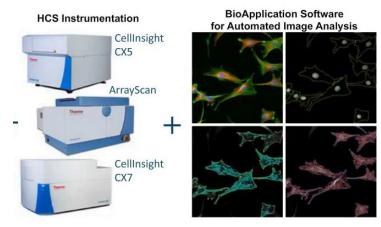




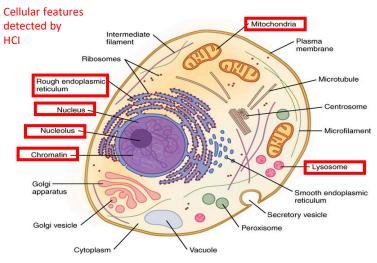
Identify 51 chemicals with *in vivo* repeat-dose toxicity data from guideline Data obtained from ToxRefDB v2.0 using species=rat, administration subchronic and chronic studies route=oral, study type = (subchronic | chronic) and target organ = Liver

In vitro Rat Hepatocyte HCI Assay (HepRn)

- Cell model: Rat primary hepatocytes
- Assay: High-content imaging (HCI)
 - Steatosis: LipidTox[®]
 - ER Stress: GADD153 (CHOP)
 - Mitochondrial function: MitoTracker Red
 - Lysosomal Mass: LysoTracker Red
 - Apoptosis: Cytochrome C
 - DNA texture: Hoechst 33342
 - Nuclear size: Hoechst 33342
 - Cell number: Hoechst 33342
- Chemical treatments
 - Controls: (-) DMSO; (+) CCCP, Bupivacaine, Tamoxifen, Nimesulide
 - Conc: 0.2, 0.39, 0.78, 1.56, 3.12, 6.24, 12.5, 25, 50 and 100 μM
 - Duration: 24, 48 and 72 h.
 - Reps: 2 on plate

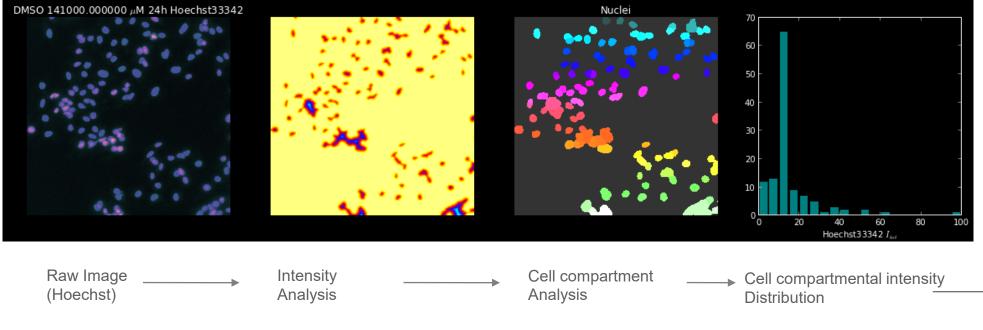


Thermofisher.com



Wikimedia.org

HCI Data Based on Cytomorphological Features



Well level aggregation

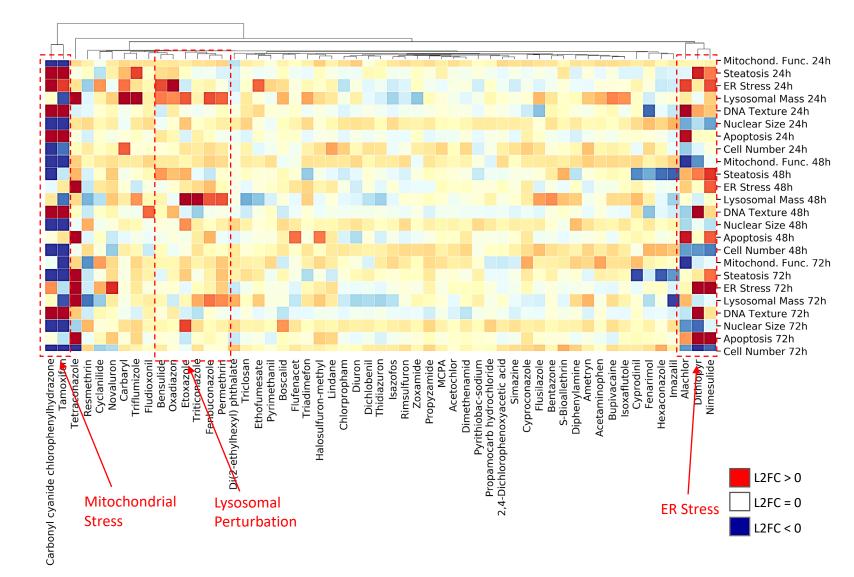
Rat hepatocyte HCI endpoints (HepRn assay)

- Mitochondrial function (MF)
- Lysosomal mass (LM)
- ER stress (ES)
- Steatosis (St)

- DNA texture (DT)
- Nuclear size (NS)
- Apoptosis (Ap)
- Cell number (CN)

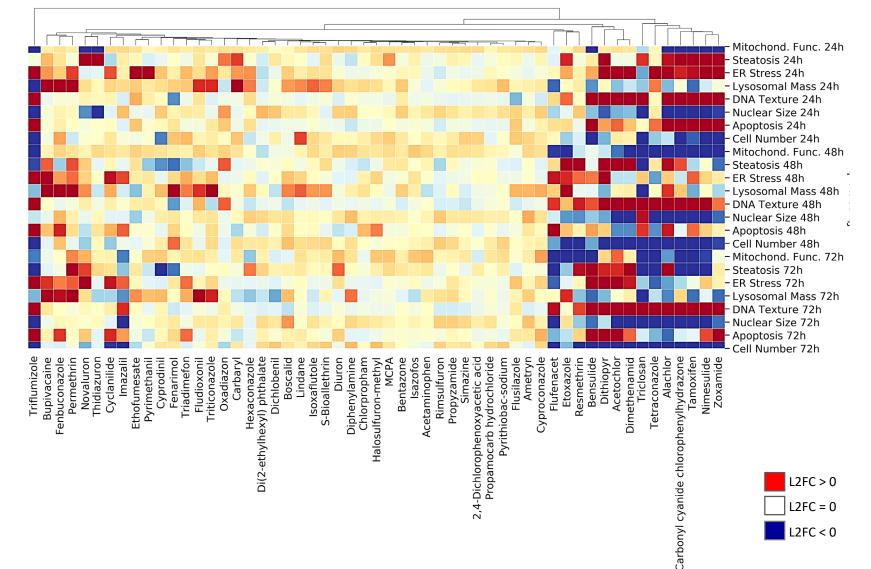
In vitro Effects – Measured by HCI (50µM)

- Log2 Fold Change (L2FC) by comparison with DMSO controls
- Summarize L2FC of all chemicals at $50 \mu M$
- Heatmap shows chemicals (columns) vs HCI features at 24, 48 and 72h and L2FC values (blue=decrease and red=increase)
- Phenotypic response categories
 - Mitochondrial stress ± cell death
 - Lysosomal ± cell death
 - ER Stress ± cell death
 - No effect
- Cross-talk between stressresponses

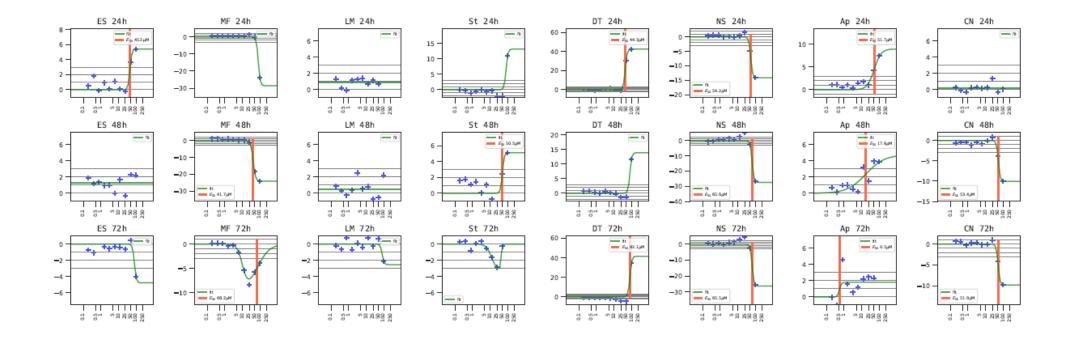


In vitro Effects – Measured by HCI (100 μ M)

- Summarize L2FC of all chemicals at 100µM
- Heatmap shows chemicals (columns) vs HCl features at 24, 48 and 72h and L2FC values (blue=decrease and red=increase)
- Phenotypic response categories
 - Increased cell death
 - Increased perturbations
 - Some chemicals still produce no effects



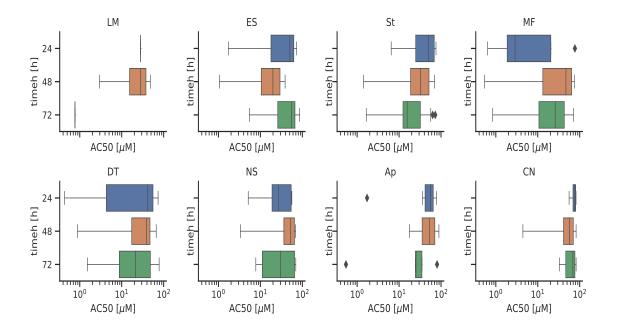
Conc-Response Analysis: E.g. Alachlor



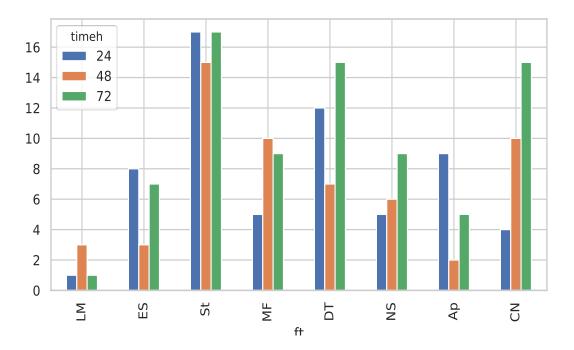
- Conduct concentration-response analysis of all chemicals using the R/tcpl (Filer, 2014) package
- Standardize the data using DMSO controls and identify the best curve-fits (constant, exponential, hill and gainloss)
- For non-constant fits find the AC₅₀ for each chemical, assay, time point

- Filter 51*8*3 curve-fits for quality manually:
 - AC₅₀ within range of test conc.
 - More than one response |Z|>3
 - Maximum response |Z|>3
 - Noisy fits

HepRn HCI Potency values and Hits



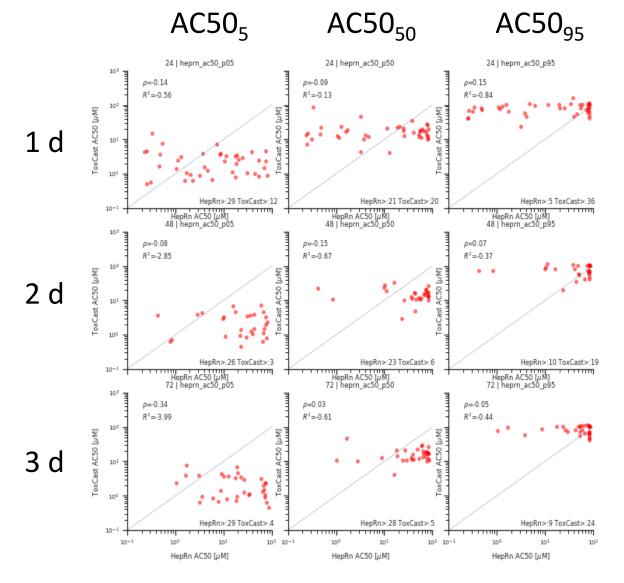
Mitochondrial function and DNA texture most sensitive assays



Hit = chemical & endpoint has high quality curve-fit

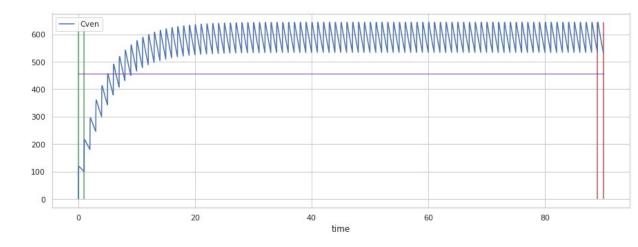
Compare HepRn with ToxCast assays

- Get the AC₅₀ values for the 51 chemicals from ToxCast using NCCT InVitroDB v3.0
- Compare the 5th,50th and 95th percentiles of AC₅₀ values between HepRn and ToxCast assays
- The ToxCast AC₅₀ values are generally lower than corresponding HepRn values



In Vitro to In Vivo Extrapolation

- Use physiologically-based toxicokinetic modeling (PBTK) extrapolate AC₅₀ values to administered dose equivalents (ADEs)
- Used the following approaches:
 - A. Concentration at steady state (Css)
 - B. Physiologically-based pharmacokinetic modeling (PBTK)
 - Clearance: restrictive
 - Dose metrics: Cmax, Cave, AUC
 - Comparing in vitro conc. to in vivo dose
 - time-initial-matched
 - time-averaged
 - time-final-matched
- Implemented using R/httk package (Pearce et al. 2017)



time-initial-matched: In vitro conc of hepatocytes = venous conc to in vivo at the same time time-averaged: In vitro conc of hepatocytes = average venous conc in rats across time time-final-matched: In vitro conc of hepatocytes is venous conc in vivo at the final time

POD_{tox} vs POD_{nam}

POD_{nam} based on:

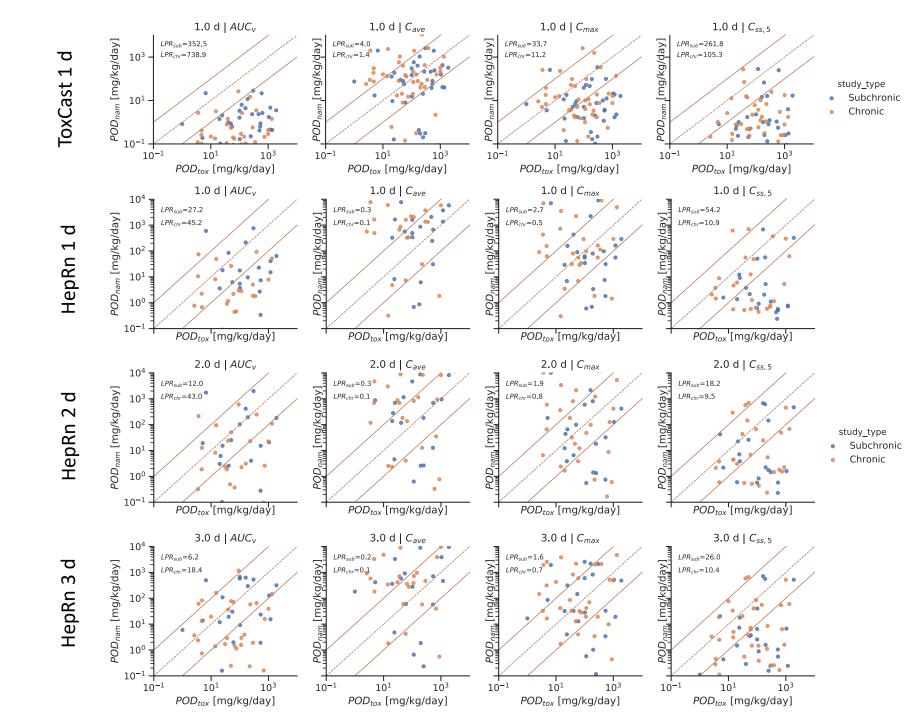
- AC_{50,5}: In vitro potency using 5th percentile of AC₅₀ from two assays:
 - HepRn: 1, 2 and 3 d exposures
 - ToxCast: using 1 d exposure
- ADE: PBTK:time-averaged using Css 5th percentile ($C_{ss,5}$), AUC_v, C_{ave} and C_{max}

POD_{tox} based on:

- LOAEL₅: 5th percentile of lowest observed adverse effect levels in liver
- Study types: Chronic and subchronic

Compare using the "POD Ratio":

- PR = mean(POD_{tox}/POD_{nam})
 - PR>1 is health protective
 - PR<1 not health protective



HepRn Subchronic POD_{nam} vs POD_{sub}

 POD_{nam} = the ADE based on 5th percentile of in vitro AC₅₀

 $POD_{tox} = 5^{th}$ percentile of lowest observed adverse effect levels (LOAEL₅) for subchronic rat liver effects

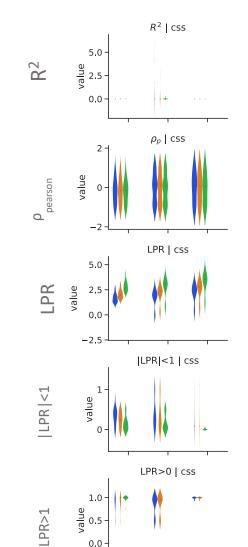
 $LPR = log10(mean(POD_{tox}/POD_{nam}))$

|LPR|<1 = fraction of chemicals for which POD_{nam} 10x greater or less than POD_{tox}

LPR>1 = fraction of chemicalsunderpredicted (healthprotective)

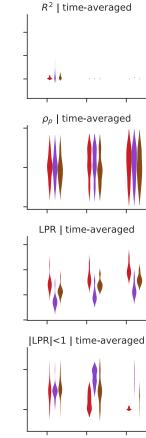
 $\rho_{p} = Corr(POD_{tox}/POD_{nam})$

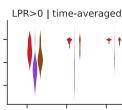
 R^2 = Coefficient of determination



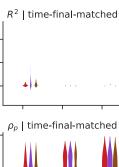
HepRn HepRnAtgToxCast

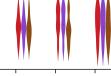
assay





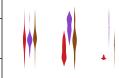
HepRn HepRnAtgToxCast assay



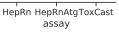


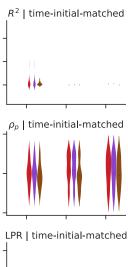
LPR | time-final-matched

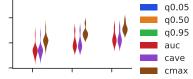
|LPR|<1 | time-final-matched



LPR>0 | time-final-matched +

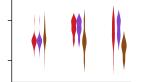


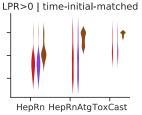




dose_metric

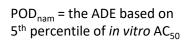
|LPR|<1 | time-initial-matched





assay

HepRn Chronic POD_{nam} vs POD_{chr}



 $POD_{tox} = 5^{th}$ percentile of lowest observed adverse effect levels (LOAEL₅) for subchronic rat liver effects

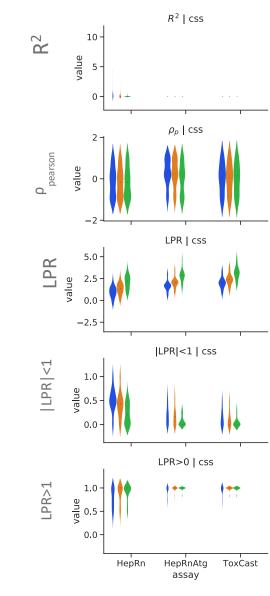
LPR = log10(mean(POD_{tox}/POD_{nam}))

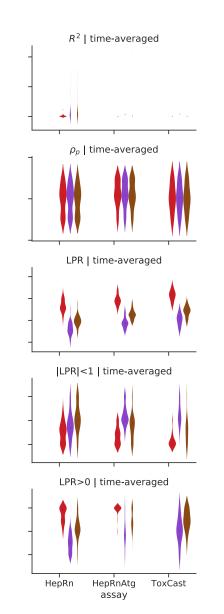
|LPR|<1 = fraction of chemicals for which POD_{nam} i10x greater or less than POD_{tox}

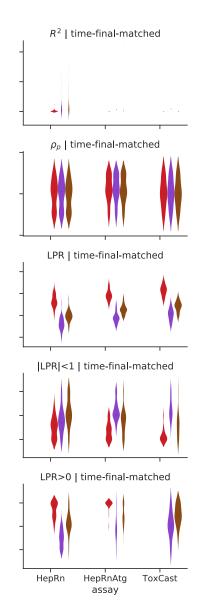
LPR>1 = fraction of chemicals underpredicted (health-protective)

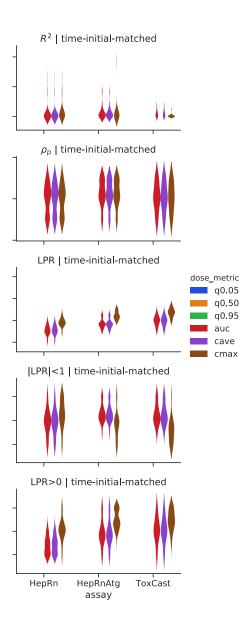
 $\rho_p = Corr(POD_{tox}/POD_{nam})$

R² = Coefficient of determination

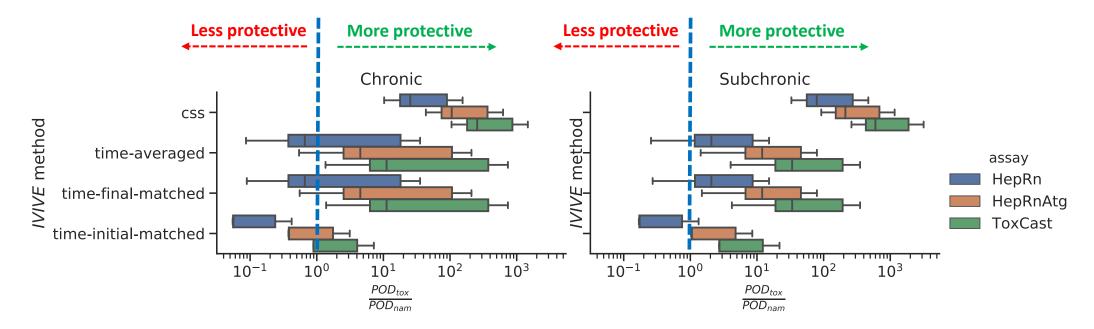








How does HepRn Compare with ToxCast?



- Compare the POD ratios (PR) across different IVIVE approaches and assays (averaged over dose-metrics)
- HepRnAtg: Combine HepRn and ToxCast:Attagene to capture non-specific (general stress responses) as well as specific chemicals (receptor-mediated pathways
- IVIVE approach matters for
 - PR for Css >> PR for PBTK
 - PR for time-averaged similar to PR for time-finaltime-matched
 - PR for time-initial-matched << PR for time-averaged
- On average, POD_{nam} from HepRn are 15-fold less than ToxCast (findings are consistent with Paul-Friedman et. al.)



Visually integrate all information about the chronic rat hepatotoxicants:

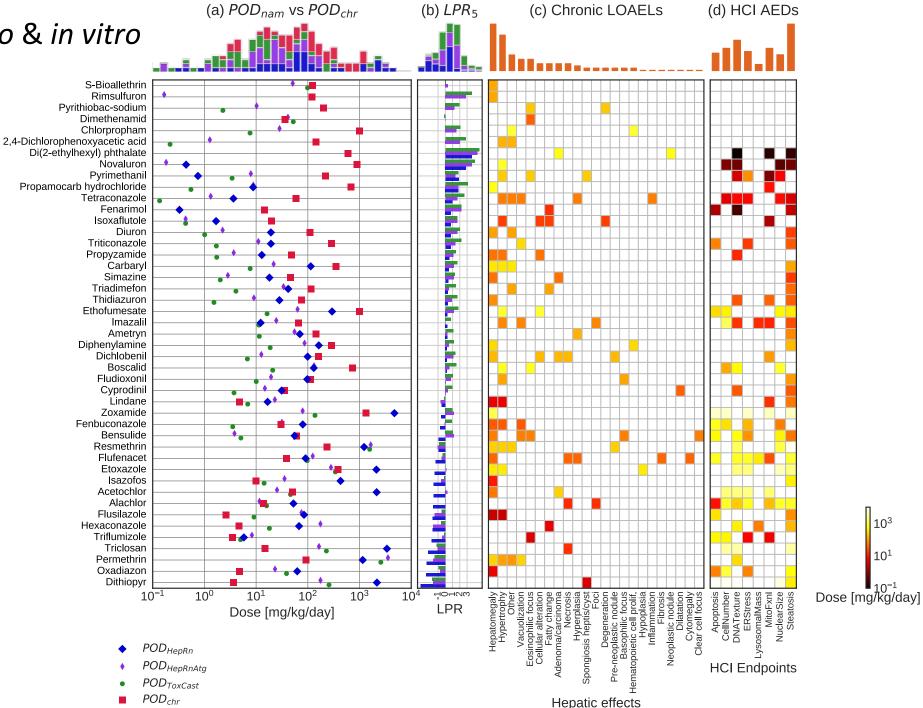
- Chronic POD values (POD_{chr})
- POD_{nam}
 - Assays: HepRn, ToxCast, HepRnAtg
 - IVIVE: time-averaged, C_{max}
- LPR for each assay type
- Chronic hepatic effects and corresponding LOEAL values
- HepRn assay endpoints and corresponding ADE₅

Highlight chemicals for which HepRn assay is protective (LPR>0), not protective (LPR<0) and reasonably predictive (|LPR|<1).

Examine the relationship between *in vivo* hepatic effects and *in vitro* cytomorphological changes

Overpredictions (LPR<0) of the HepRn assay could be due to issues with PBTK or due to specific receptor mediated mechanisms

$$\begin{split} & \mathsf{POD}_{nam} = \mathsf{the} \; \mathsf{ADE} \; \mathsf{based} \; \mathsf{on} \; \mathsf{5}^{\mathsf{th}} \; \mathsf{percentile} \; \mathsf{of} \; \textit{in vitro} \; \mathsf{AC}_{\mathsf{50}} \\ & \mathsf{POD}_{\mathsf{tox}} = \mathsf{5}^{\mathsf{th}} \; \mathsf{percentile} \; \mathsf{of} \; \mathsf{lowest} \; \mathsf{observed} \; \mathsf{adverse} \; \mathsf{effect} \; \mathsf{levels} \\ & (\mathsf{LOAEL}_{\mathsf{5}}) \; \mathsf{for} \; \mathsf{chronic} \; \mathsf{rat} \; \mathsf{liver} \; \mathsf{effects} \\ & \mathsf{LPR} = \mathsf{log10}(\mathsf{mean}(\mathsf{POD}_{\mathsf{tox}}/\mathsf{POD}_{\mathsf{nam}})) \end{split}$$



Summary

- In vitro and in silico NAM-based PODs are generally health-protective
- IVIVE assumptions matter: Steady state concentrations are more health-protective than those based on PBTK models
- In vitro assays matter: PODs produced using the same species (rat) cell type (hepatocyte) are <u>closer</u> to PODs from animal testing
- <u>Poor correlation</u> between NAM-based PODs from animal testing could be due to:
 - In vitro assays: using single cell type, narrow biological coverage, or data analysis, etc.
 - TK: *In vitro* partitioning, PBTK gut absorption, etc.
 - Toxicodynamics (TD): biology is complex!

Acknowledgements and Questions

Todor Antonijevic Rusty Thomas Josh Harrill Thomas Knudsen Katie Paul-Friedman John Wambaugh Greg Honda



Additional slides