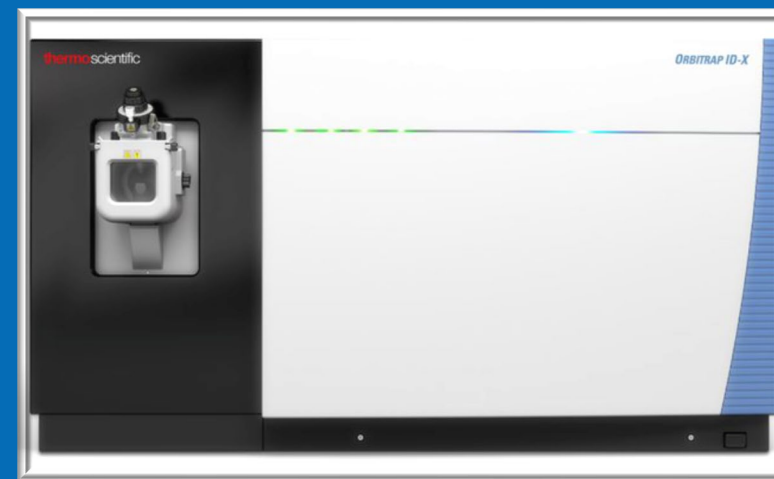


# Identification of Bioactive Contaminants Associated With Wastewater Using a Receptor Pulldown Assay and High Resolution Mass Spectrometry

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*\*The contents of this presentation neither represent nor necessarily reflect official US EPA policy\**

# Acknowledgments

## US EPA

- ORD – GLTED: J. Cavallin, A. Cole, R. Hofer, D. Villeneuve
- ORD – CEMM: D. Ekman
- Region 8 – J. Kinsey
- NEIC – J. Beihoffer, K. Keteles

## NPS

- Rebecca Weissinger

## USGS

- B. Battaglin, P. Bradley, D. Winkleman

## Univ. of Toronto

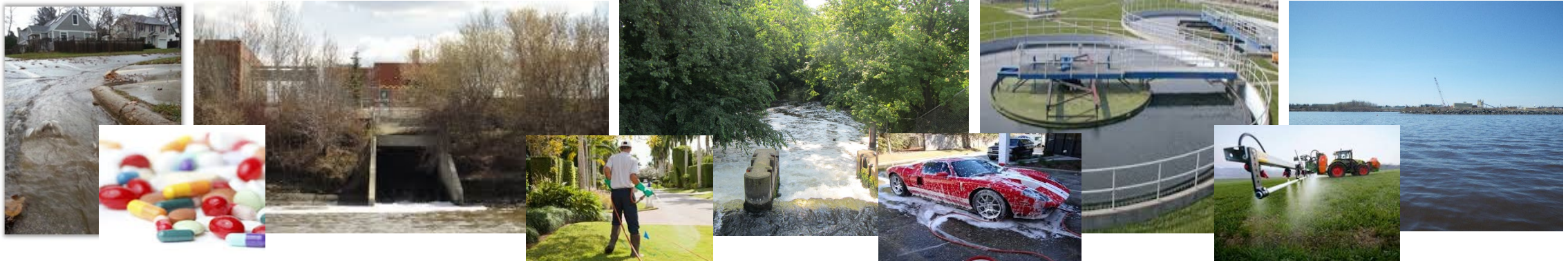
- Hui Peng





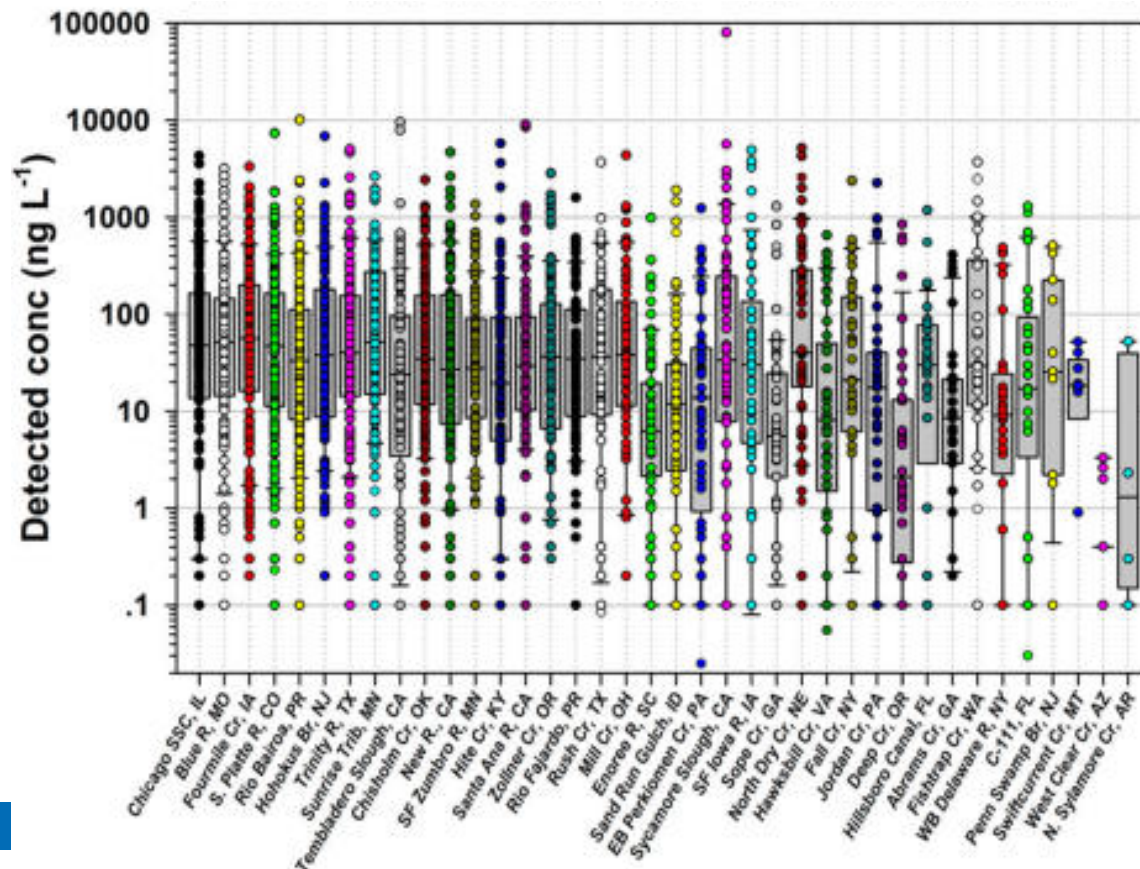
# Chemicals in the Environment

- Increasing number of compounds can be detected in the environment
- Contaminants of emerging concern (CECs)
  - Pharmaceuticals and personal care products (PPCPs)
  - Agricultural chemicals (pesticides, growth promoters)
  - Industrial chemicals (plasticizers, flame retardants, PFAS)



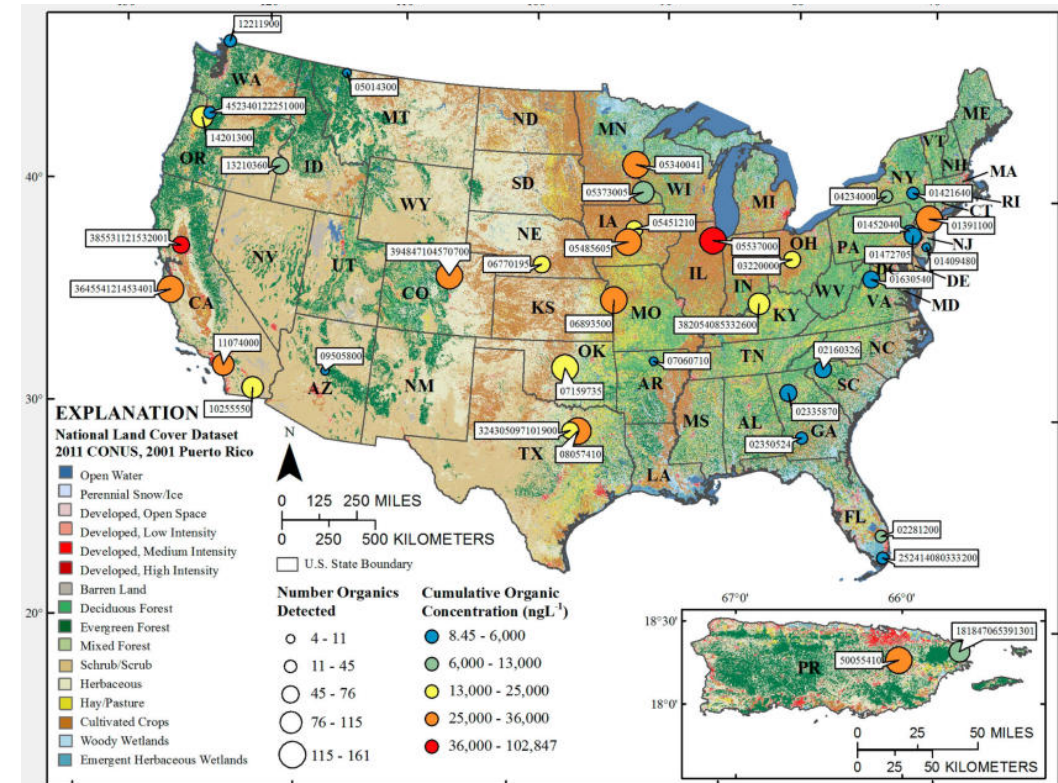
# Surface Water Contaminants in the US

- 406/719 organic compounds detected
- 4 – 161 compounds detected at a single site
- Cumulative concentrations ranged 8 – 103000 ng/L



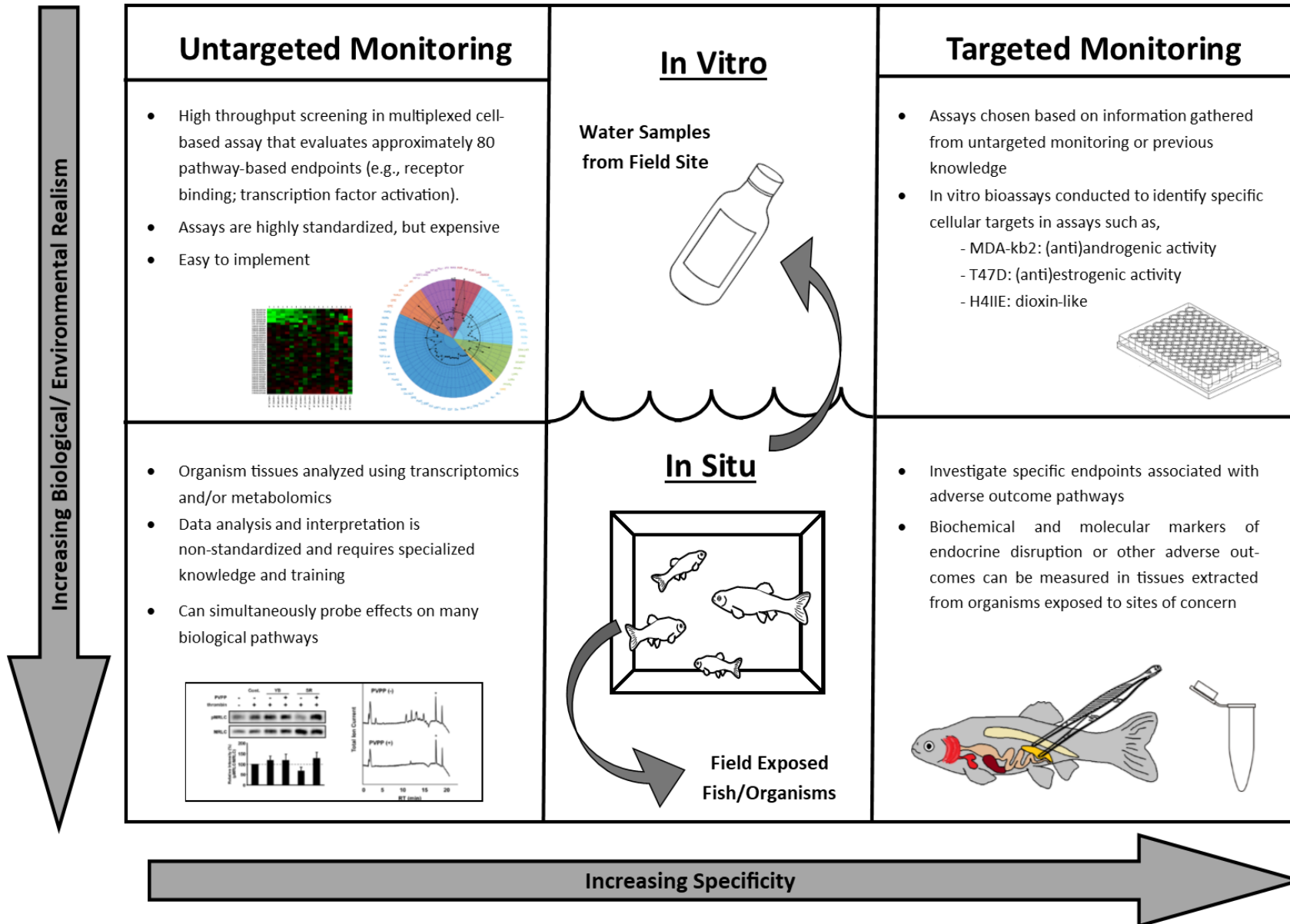
## Expanded Target-Chemical Analysis Reveals Extensive Mixed-Organic-Contaminant Exposure in U.S. Streams

Paul M. Bradley,<sup>\*,†,§</sup> Celeste A. Journey,<sup>†</sup> Kristin M. Romanok,<sup>‡</sup> Larry B. Barber,<sup>§</sup> Herbert T. Buxton,<sup>||</sup> William T. Foreman,<sup>⊥</sup> Edward T. Furlong,<sup>⊥</sup> Susan T. Glassmeyer,<sup>#</sup> Michelle L. Hladik,<sup>∇,⊙</sup> Luke R. Iwanowicz,<sup>○</sup> Daniel K. Jones,<sup>◆</sup> Dana W. Kolpin,<sup>||</sup> Kathryn M. Kuivila,<sup>⊗</sup> Keith A. Loftin,<sup>∞</sup> Marc A. Mills,<sup>#</sup> Michael T. Meyer,<sup>∞</sup> James L. Orlando,<sup>∇</sup> Timothy J. Reilly,<sup>‡</sup> Kelly L. Smalling,<sup>‡</sup> and Daniel L. Villeneuve<sup>☆</sup>





# Generalized Effects-based Approach



# 21<sup>st</sup> Century Approach: High-Throughput Toxicology (HTT)

- EPA and other federal agencies began the Toxicity Forecaster (ToxCast) and Tox21 in 2007 as part of an effort to advance chemical toxicity assessments
- Focus on in vitro testing vs traditional whole animal testing
- Greatly reduces cost of screening chemicals; ~10,000 currently screened through 100+ assays
- Provides a new source of methods and data that can be applied to assessment of environmental data

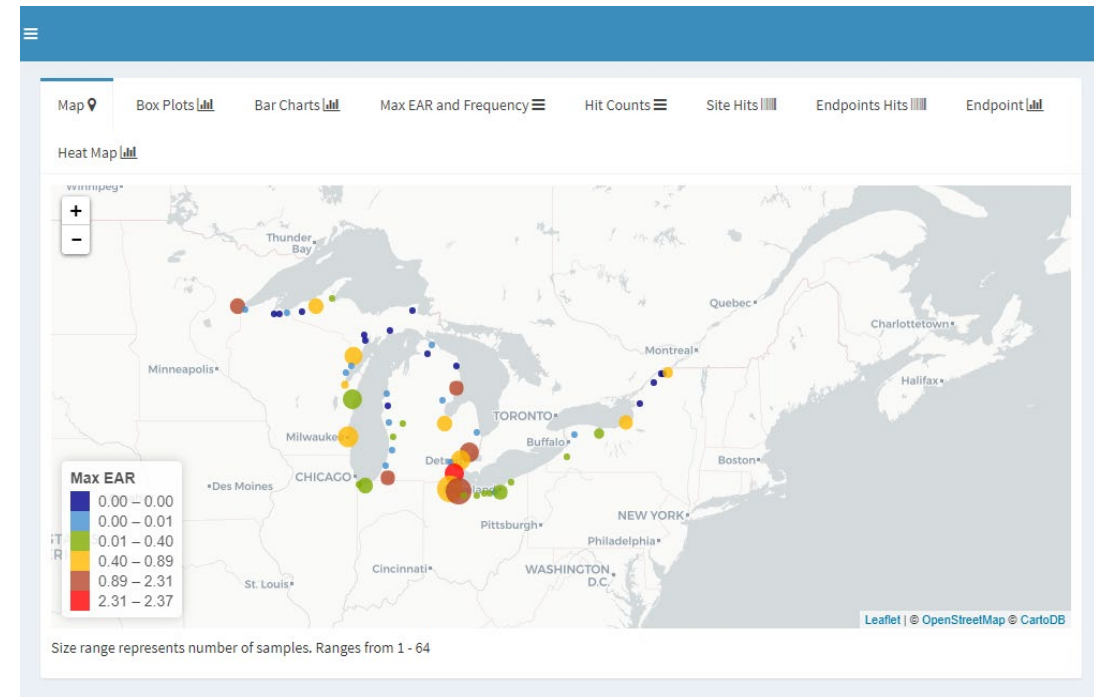
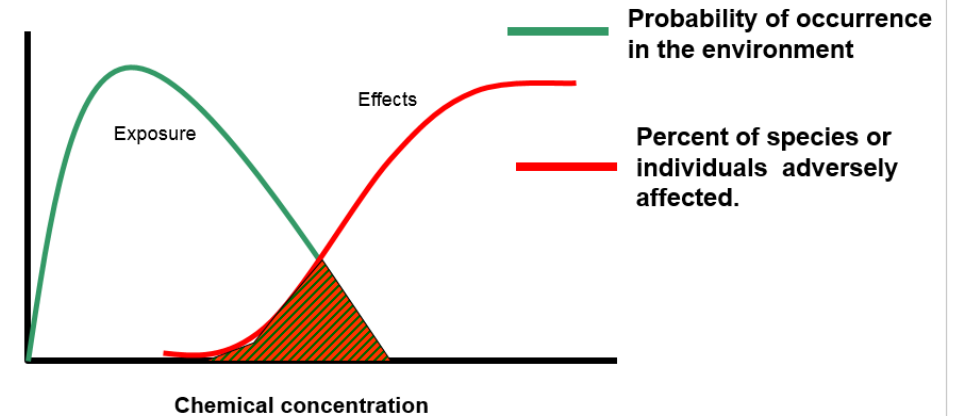


- 1536 well HTS
- 10,000 chemicals
- 25 assays per year



# Using HTT To Predict and Prioritize Chemicals

- Compare observed chemical concentrations to effects concentration in ToxCast database
  - Exposure-activity ratio (EAR)
  - Bioactivity-exposure ratio (BER)
- Simple, rapid approach to screen chemicals for potential biological effects
- Can prioritize sites or chemicals for further assessment
- R package “toxEval” allows for automated comparison of chemical monitoring data with ToxCast (<https://github.com/USGS-R/toxEval>)



# HTT-based Screening of Environmental Mixtures

- HTS assays can be used as effects-based monitoring tools to directly screen environmental extracts
- Assay responses incorporate unknowns and potential interactions of mixture components

**ENVIRONMENTAL**  
Science & Technology

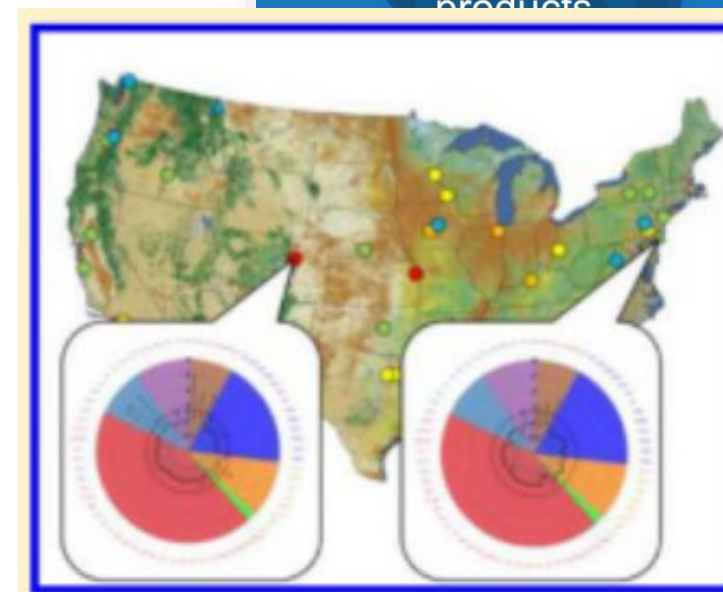
Cite This: *Environ. Sci. Technol.* 2019, 53, 973–983

pubs.acs.org/est

## Potential Toxicity of Complex Mixtures in Surface Waters from a Nationwide Survey of United States Streams: Identifying in Vitro Bioactivities and Causative Chemicals

Brett R. Blackwell,<sup>\*,†,§</sup> Gerald T. Ankley,<sup>†,§</sup> Paul M. Bradley,<sup>‡,§</sup> Keith A. Houck,<sup>§</sup> Sergei S. Makarov,<sup>||</sup> Alexander V. Medvedev,<sup>||</sup> Joe Swintek,<sup>⊥</sup> and Daniel L. Villeneuve<sup>‡,§</sup>

- Only one of 11 biological endpoint (estrogenic activity) explained by measured chemicals







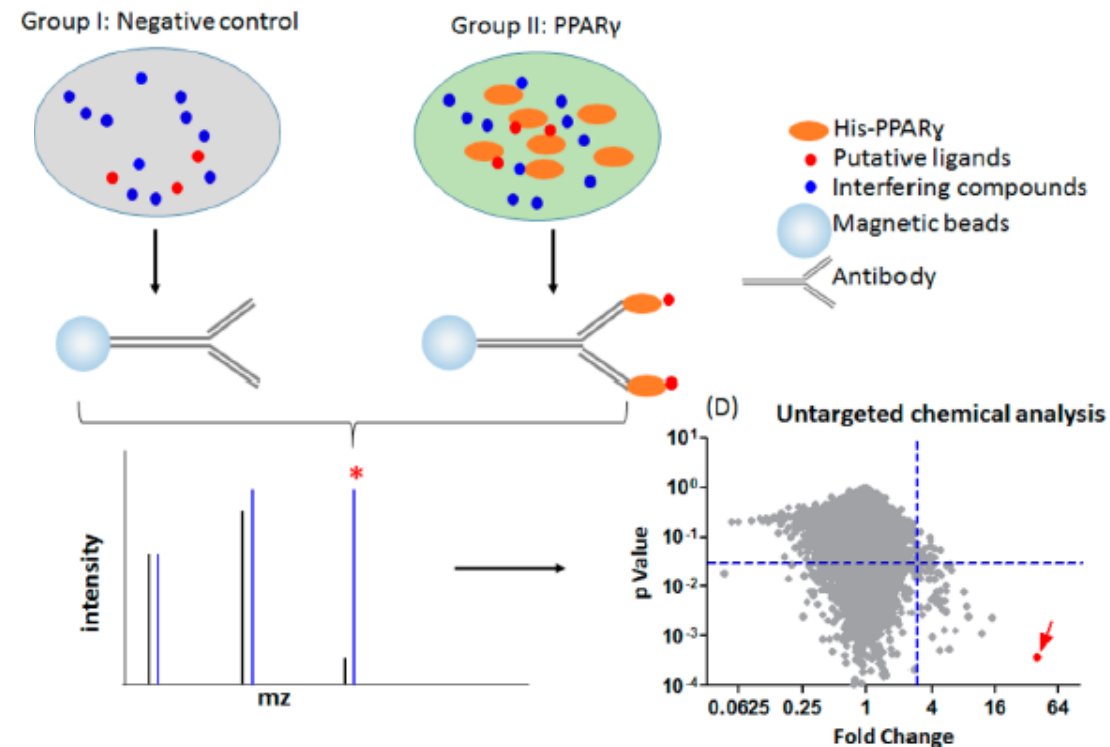
# NTA for Identification of Bioactive Chemicals

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- NTA methods can help uncover the “unknowns” of chemical exposure
- Bioactivity can prioritize samples/features for NTA analysis
- Fractionation and subsequent bioassay/NTA analysis can further define chemical space of interest (effects-directed analysis, EDA)

# Receptor Pulldown Assay and HR-MS

- Similar to bioassays, mass spectrometry techniques can utilize proteins to help elucidate bioactive compounds
- Effects-based monitoring data can guide selection of biological targets and environmental sites of interest
- Employing these methods in a case study of wastewater associated contaminants in the Colorado River



*Peng et al. 2016, Environ. Sci. Technol. 50:7816-7824*

# Colorado River Case Study

- 2013 National Park Service measured CECs along Colorado River between Arches NP and Canyonlands NP
- Detected CECs far downstream of Moab WWTP
  - Designed 1950s for 5000 – millions of visitors per year
  - Treatment plant upgraded in 2018

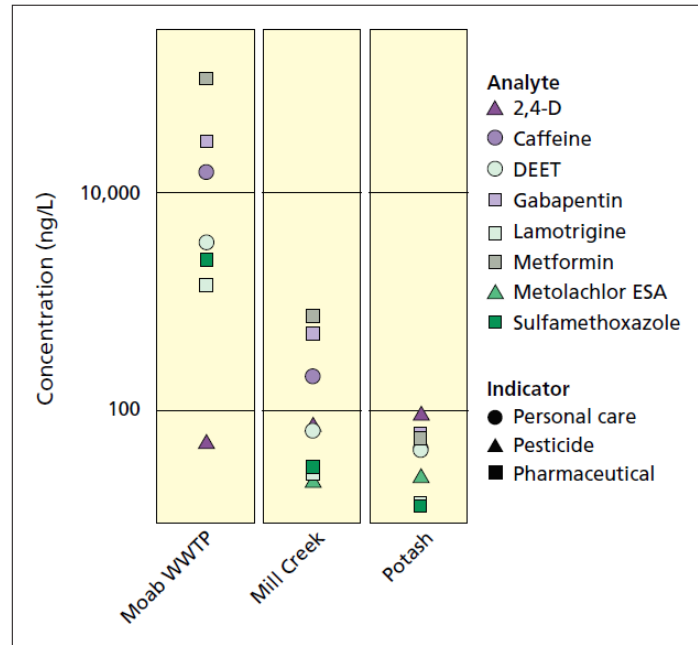
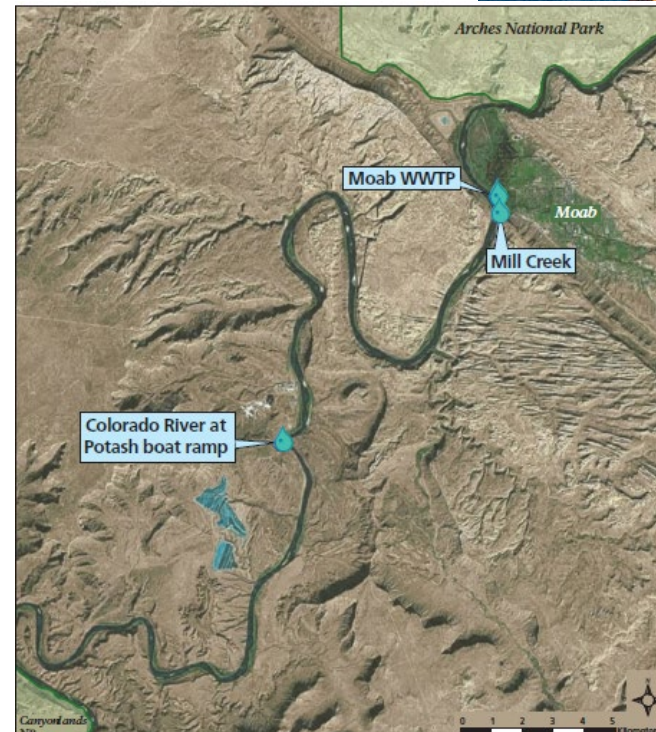


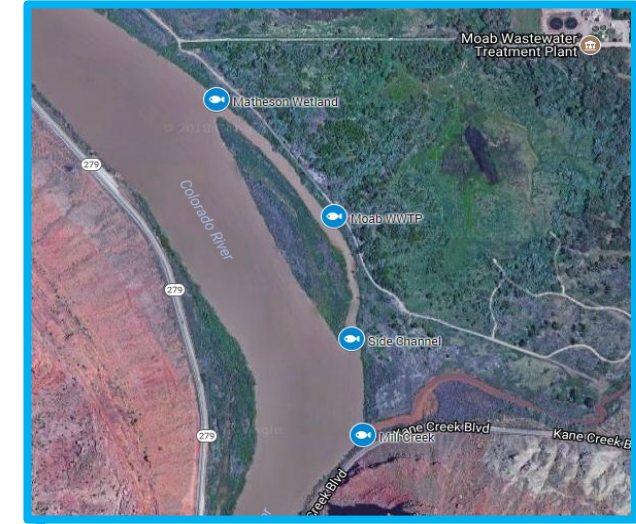
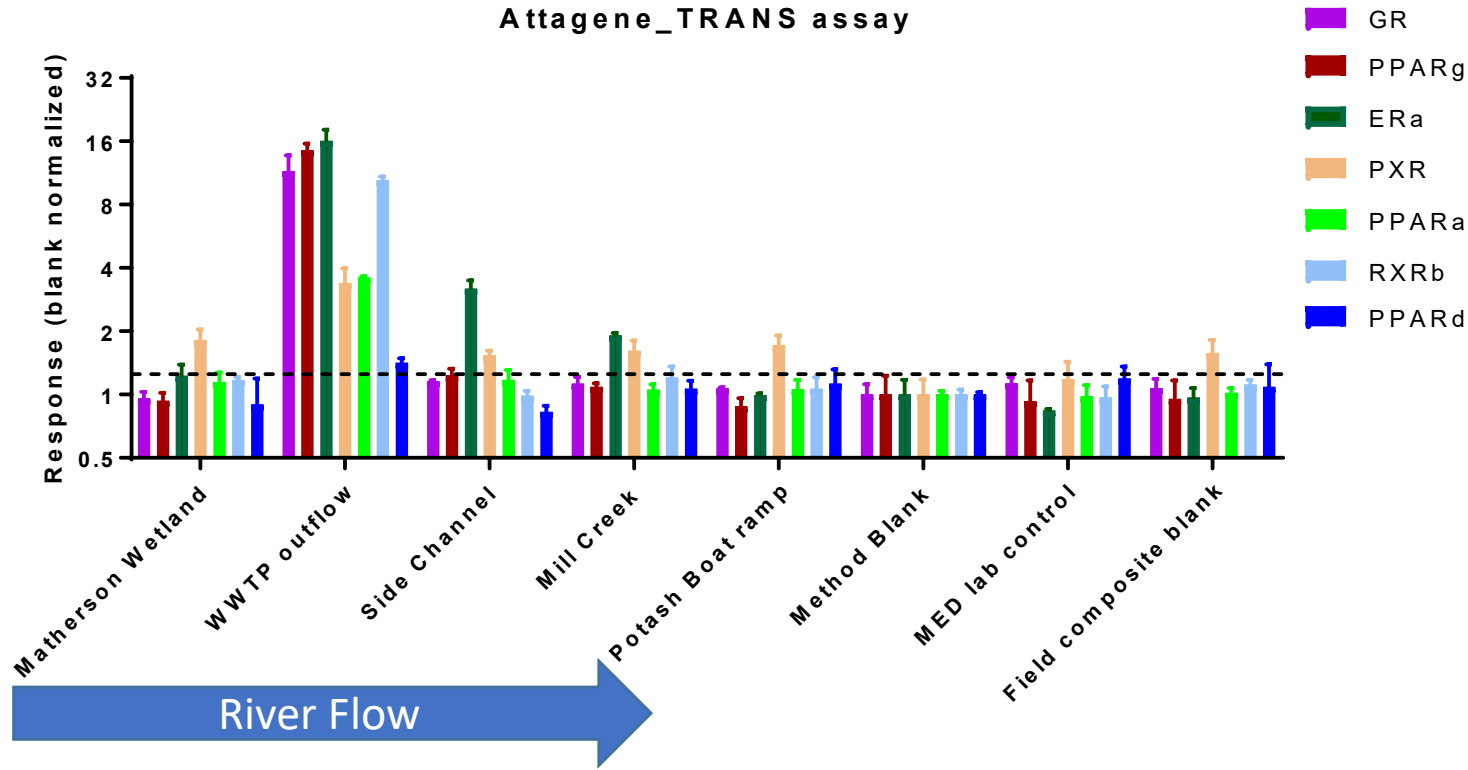
Figure 3-2. Concentrations of pharmaceuticals and personal care products decreased by orders of magnitude as samples moved downstream from the wastewater treatment plant outflow to the Colorado River at Mill Creek and to the Colorado River at the Potash boat ramp. Note the logarithmic scale on the y-axis.





# Colorado River Bioactivity

Moab April 2018  
Attagene\_TRANS assay



Target	Hazard Considerations
ER	<ul style="list-style-type: none"> <li>Well established hazard to aquatic vertebrates</li> </ul>
GR	<ul style="list-style-type: none"> <li>Hazards to aquatic life not well defined</li> </ul>
PPARg	<ul style="list-style-type: none"> <li>Hazards to aquatic life not well defined</li> </ul>

# Field Collection Methods

- Caged fathead minnows deployed at each site
- Autosampler deployed to collect composited water samples
  - Water samples align with duration of fish exposure
  - Provide representative samples for chemical or bioassay analysis
- Bimonthly grab water samples collected



## General Approach

## Supervised

## Unsupervised

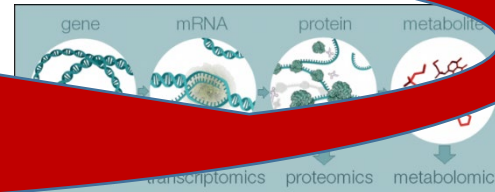
Caged fathead minnows



Endpoints associated with established AOPs  
Chemical and molecular markers  
E2, vitellogenin (VTG)



- Metabolomics



Surface Waters;  
Extracts

In vitro bioassays

- ER activity
- GR activity
- PPAR activity



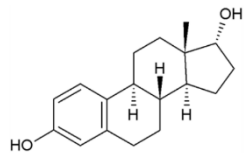
Attagene Assays

- In vitro assays
- 70 endpoints

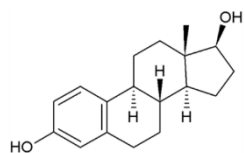


# Bioassay Workflow

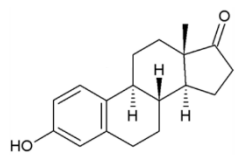
## Targeted Steroidal Estrogens



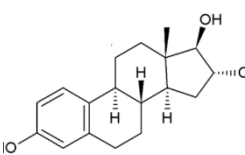
17α-estradiol



17β-estradiol

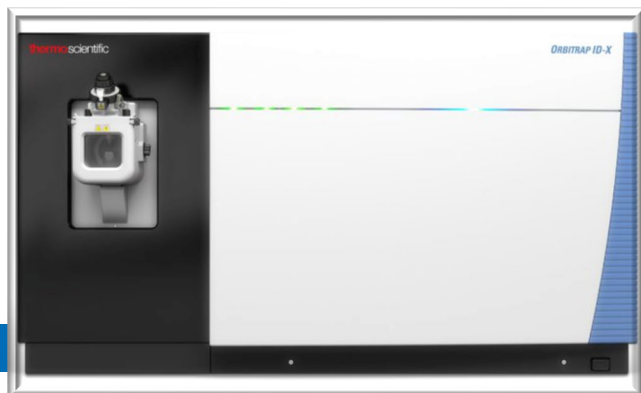


estrone



estriol

## Glucocorticoid Suspect Screening



Ambient  
Water  
Sample

Filtered

Extracted  
200mg HLB

“Unknown”  
Chemical  
Mixture

MeOH

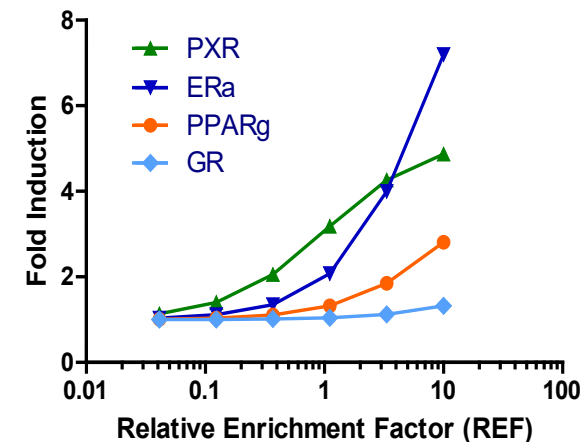
2000x conc.

DMSO

1000x conc.

Orbitrap ID-X System

## In vitro bioassays

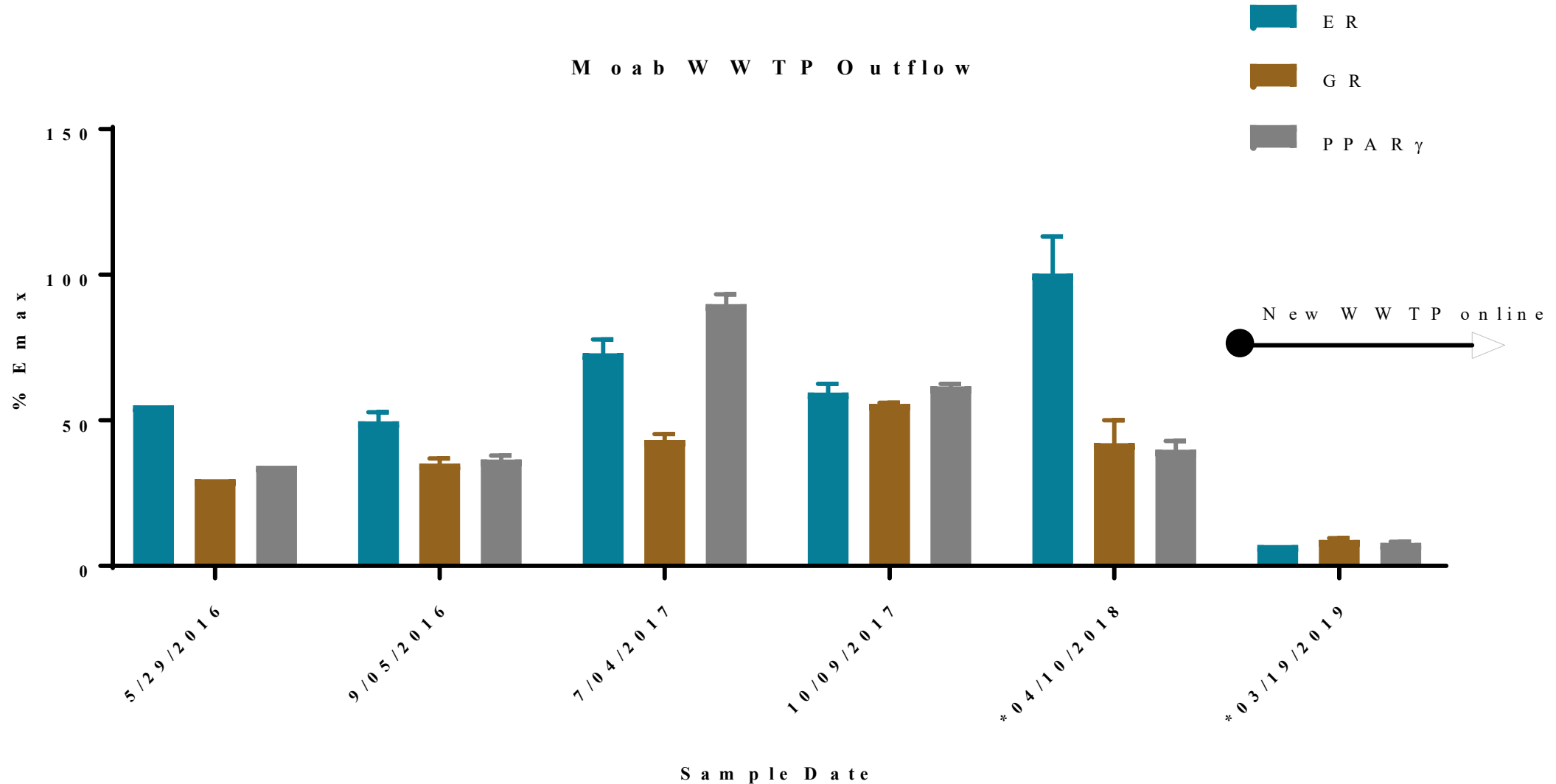


## PPARγ Pulldown Assay

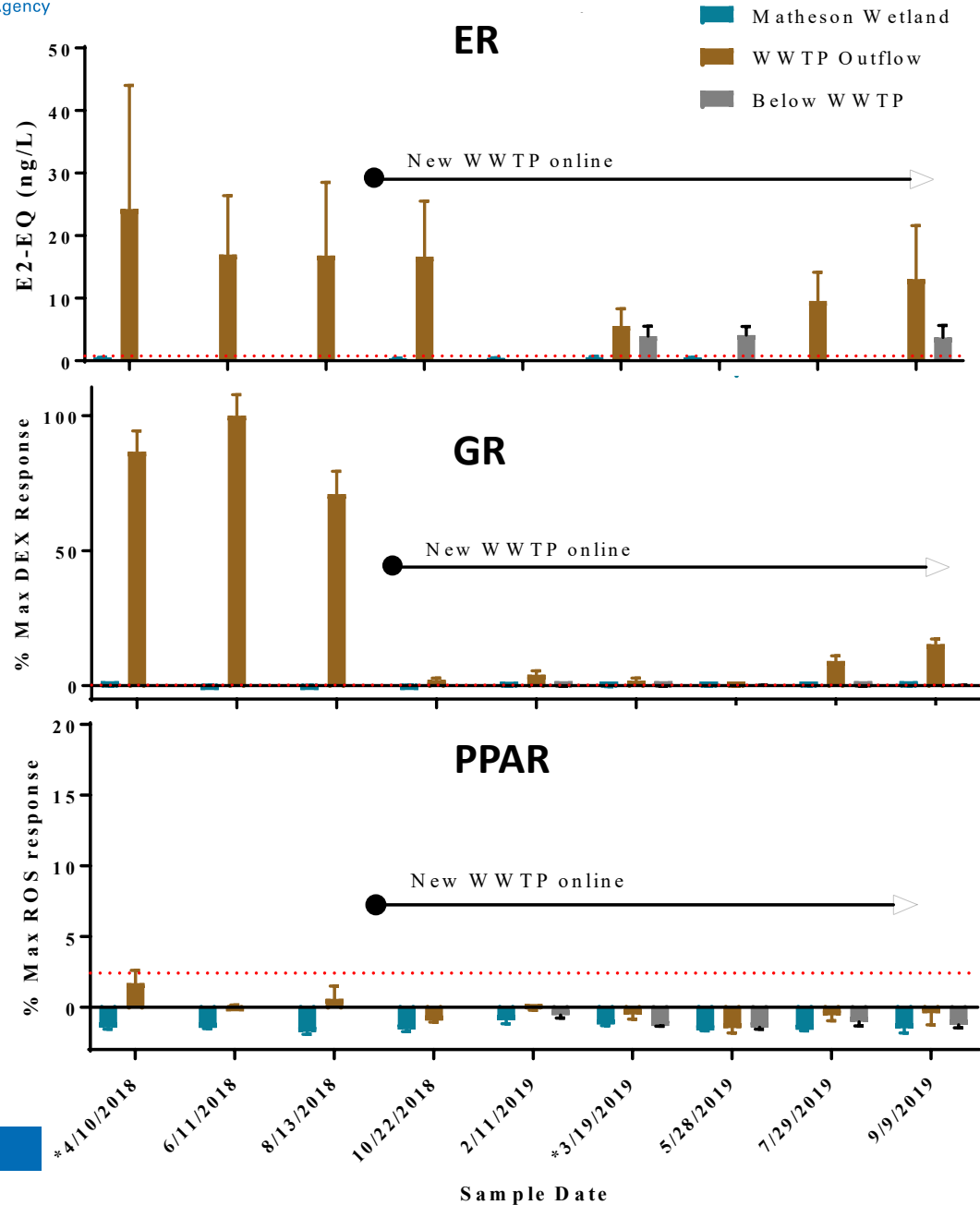




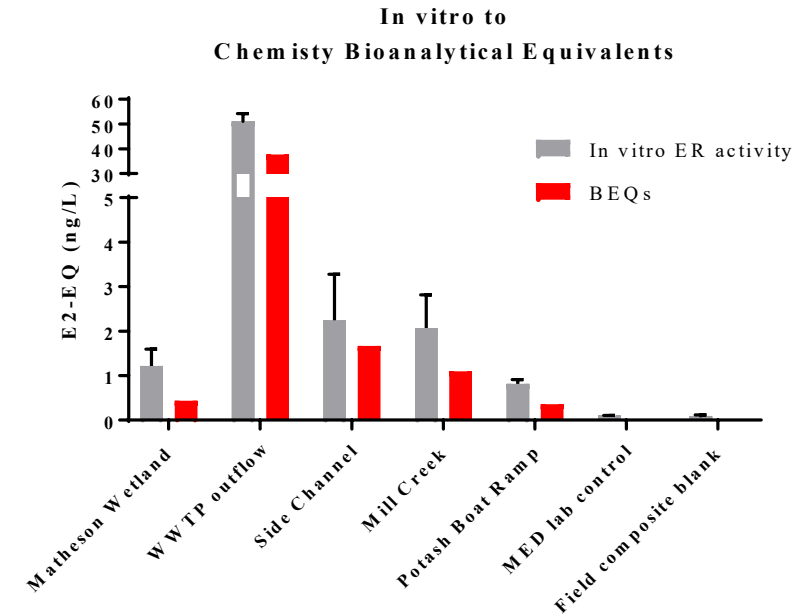
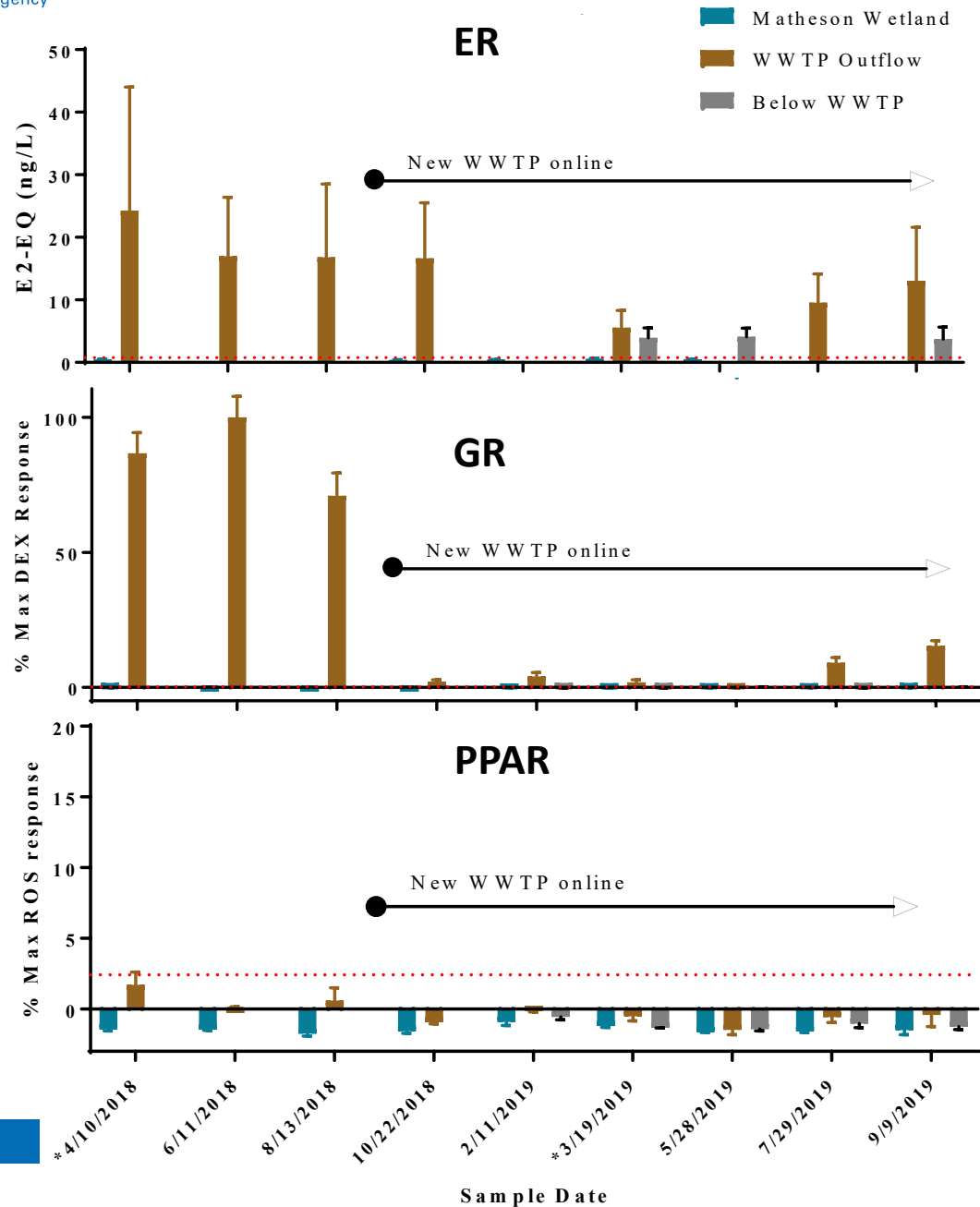
# Attagene Trans-Factorial Bioassay Results



# Targeted Bioassays



- ER and GR activity decrease greatly following WWTP upgrade
- PPAR $\gamma$  assay not sensitive enough to detect activity above baseline

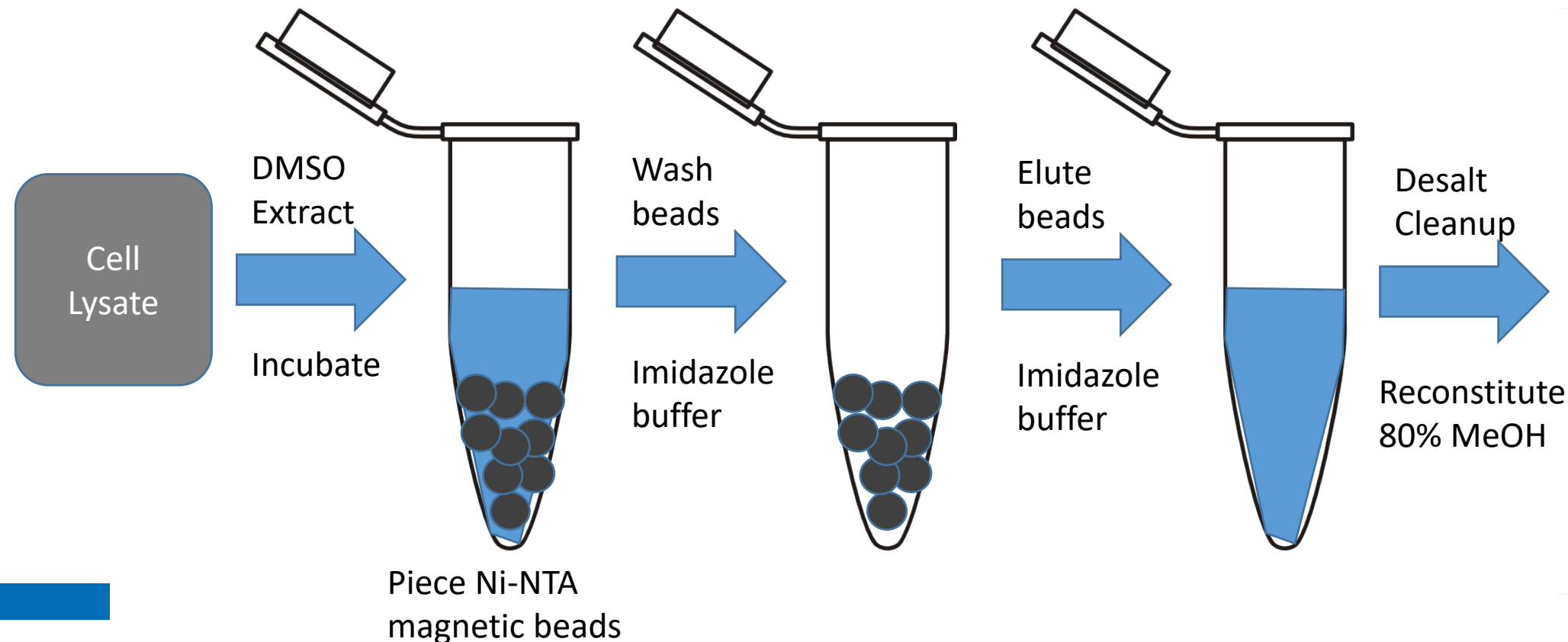


**No observed chemicals can explain GR or PPAR related activity**



# PPAR $\gamma$ Receptor Pulldown Assay

- HIS-tagged human PPAR $\gamma$  – Ligand Binding Domain (LBD) only
- Receptor overexpressed in E.Coli; cell lysate used for assay



# Pulldown Assay: LC-MS Analysis

- Vanquish Horizon LC System
- ID-X Orbitrap

Instrument Parameter	Value
Injection Volume	2 uL
Flow rate	300 uL/min
Column	C18, 1.7um, 2.1 x 150mm
Column Compartment	40°C
Polarity	Pos/Neg

Ion Source Properties	
Ion Source Type	H-ESI ▼
Spray Voltage	Static ▼
Positive Ion (V)	3500
Negative Ion (V)	2500
Sheath Gas (Arb)	50
Aux Gas (Arb)	10
Sweep Gas (Arb)	1
Ion Transfer Tube Temp (°C)	325
Vaporizer Temp (°C)	350
APPI Lamp	Not in Use ▼
Use Ion Source Settings from Tune	<input type="checkbox"/>
FAIMS Mode	Not Installed ▼

# GR-Agonist Suspect Screening

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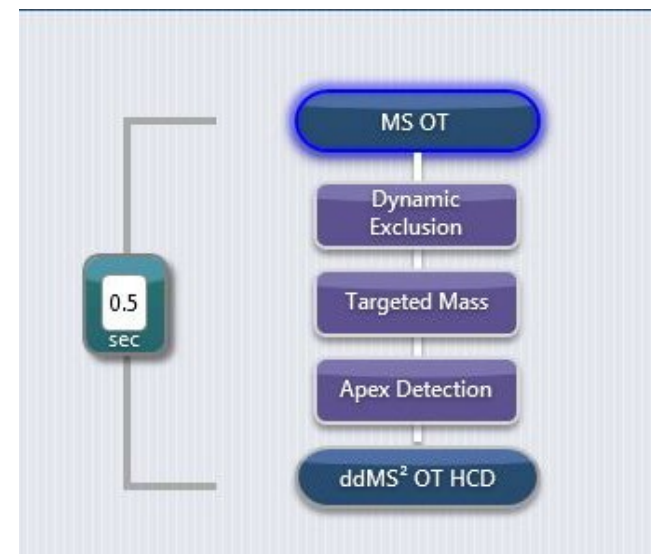
- GR receptor not readily available for pulldown assay
- GR agonist much better defined – A number have been reported in wastewater and surface waters
- Suspect screening list of 45 previously identified compounds
  - Natural corticosteroids – cortisol, cortisone, corticosterone, etc
  - Synthetic corticosteroids – dexamethasone, triamcinolone, prednisone



# Suspect Screening Analysis

- Vanquish Horizon LC System
- ID-X Orbitrap

Instrument Parameter	Value
Injection Volume	2 uL
Flow rate	300 uL/min
Column	C18, 1.7um, 2.1 x 150mm
Column Compartment	40°C
Polarity	Pos



MS Scan Properties		<a href="#">Show All</a>
Orbitrap Resolution	30000	▼
Scan Range (m/z)	300-600	
RF Lens (%)	60	
Maximum Injection Time (ms)	50	
Polarity	Positive	▼

# Suspect Screening Preliminary Results

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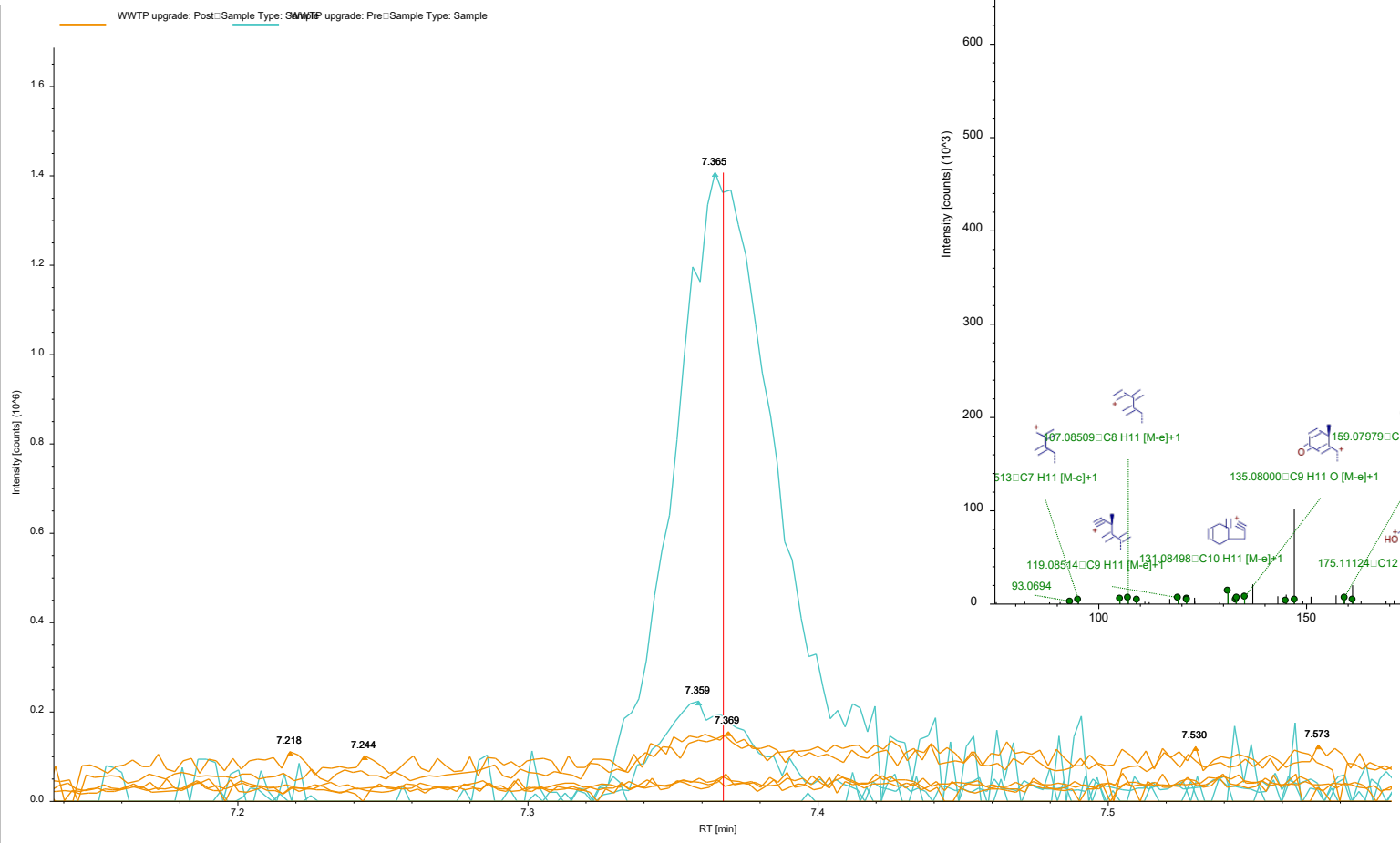
- Processing HR-MS data using Compound Discoverer v3.1
- Automated workflow: Environmental Unknown ID w Online and Local Database Searches
  - Searches against ChemSpider, MZCloud, user defined suspect list
- Data analysis very preliminary to date
- Gladly taking suggestions/recommendations for successful data analysis!

# Suspect Screening Preliminary Results

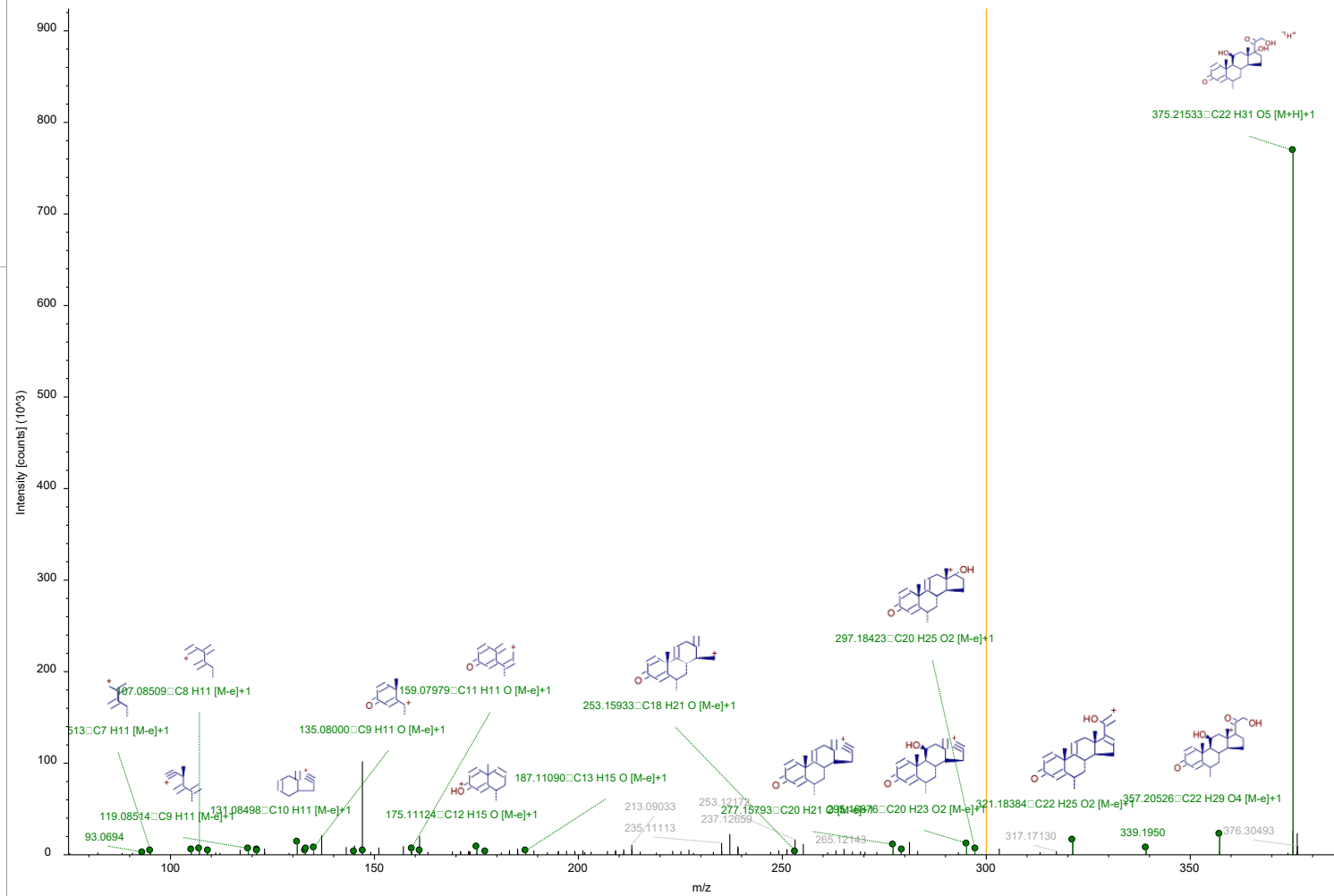
- Only WWTP site and process blanks analyzed
- 31 compounds with MS2 spectra

Compounds		Compounds per File	Merged Features	Features	mzCloud Results	ChemSpider Results	Mass List Search Results	Input Files	Specialized Traces
	Checked	Name	Formula	Annotation Sc	FISh Coverage	Molecular Weight	RT [min]	Area (Max.)	M
1	<input type="checkbox"/>		C13 H27 F N2 O7	<div><div></div><div></div><div></div></div>		342.18040	10.025	189060658	
2	<input type="checkbox"/>		C29 H64 F N7 O17	<div><div></div><div></div><div></div></div>		801.43503	5.571	3000970	
3	<input type="checkbox"/>	3,20-Dioxopregna-4,9(11),16-trien-21-yl acetate	C23 H28 O4	<div><div></div><div></div><div></div></div>		368.19894	8.226	6057044	
4	<input type="checkbox"/>		C16 H26 F2 N4 O	<div><div></div><div></div><div></div></div>		328.20730	12.571	18729871	
5	<input type="checkbox"/>		C42 H68 N3 O10 P	<div><div></div><div></div><div></div></div>		805.46484	5.774	45612841	
6	<input type="checkbox"/>		C44 H86 F2 N8 O8	<div><div></div><div></div><div></div></div>		892.65360	14.326	11642104	
7	<input checked="" type="checkbox"/>	Methylprednisolone	C22 H30 O5	<div><div></div><div></div><div></div></div>	34.57	374.20906	7.367	3358786	
8	<input type="checkbox"/>	17 $\alpha$ -Hydroxyprogesterone	C21 H30 O3	<div><div></div><div></div><div></div></div>	37.61	330.21851	7.733	34441552	
9	<input type="checkbox"/>	Hydrocortisone Valerate	C26 H38 O6	<div><div></div><div></div><div></div></div>	21.74	446.26693	10.345	15144596	
10	<input type="checkbox"/>		C18 H28 F N4 O3 P S	<div><div></div><div></div><div></div></div>		430.15993	7.646	10228015	
11	<input type="checkbox"/>	2-piperidinophenyl N-[4-(benzyloxy)phenyl]carbamate	C25 H26 N2 O3	<div><div></div><div></div><div></div></div>		402.20213	5.774	12923567	
12	<input type="checkbox"/>	N-[[[(2R,4S,5R)-5-(3-Cyclopentyl-1-methyl-1H-pyrazol-5	C22 H30 N4 O2	<div><div></div><div></div><div></div></div>		364.22515	12.760	4844295	
13	<input type="checkbox"/>		C24 H49 F O2 P2	<div><div></div><div></div><div></div></div>		450.31949	11.577	37465016	
14	<input type="checkbox"/>	Corticosterone	C21 H30 O4	<div><div></div><div></div><div></div></div>	33.62	346.21401	8.791	3391338	
15	<input type="checkbox"/>	Nor-9-carboxy- $\delta$ -9-THC	C21 H28 O4	<div><div></div><div></div><div></div></div>		344.19823	14.175	3087155	
16	<input type="checkbox"/>	Cortisol	C21 H30 O5	<div><div></div><div></div><div></div></div>	41.18	362.20897	7.534	5883962	
17	<input type="checkbox"/>	2-Octyl-3,6,9,12-tetraoxatetradecane-1,14-diol	C18 H38 O6	<div><div></div><div></div><div></div></div>		350.26582	12.180	68752768	
18	<input type="checkbox"/>	(2S,3R,4S,5S,6R)-2-[4-hydroxy-2-(3-methylbut-2-en-1-yl	C17 H24 O7	<div><div></div><div></div><div></div></div>		362.13423	10.239	3808722	
19	<input type="checkbox"/>	NP-020634	C21 H30 O6	<div><div></div><div></div><div></div></div>		400.18627	10.077	3117029	
20	<input type="checkbox"/>	Cortisone	C21 H28 O5	<div><div></div><div></div><div></div></div>	52.25	360.19383	7.619	3294584	

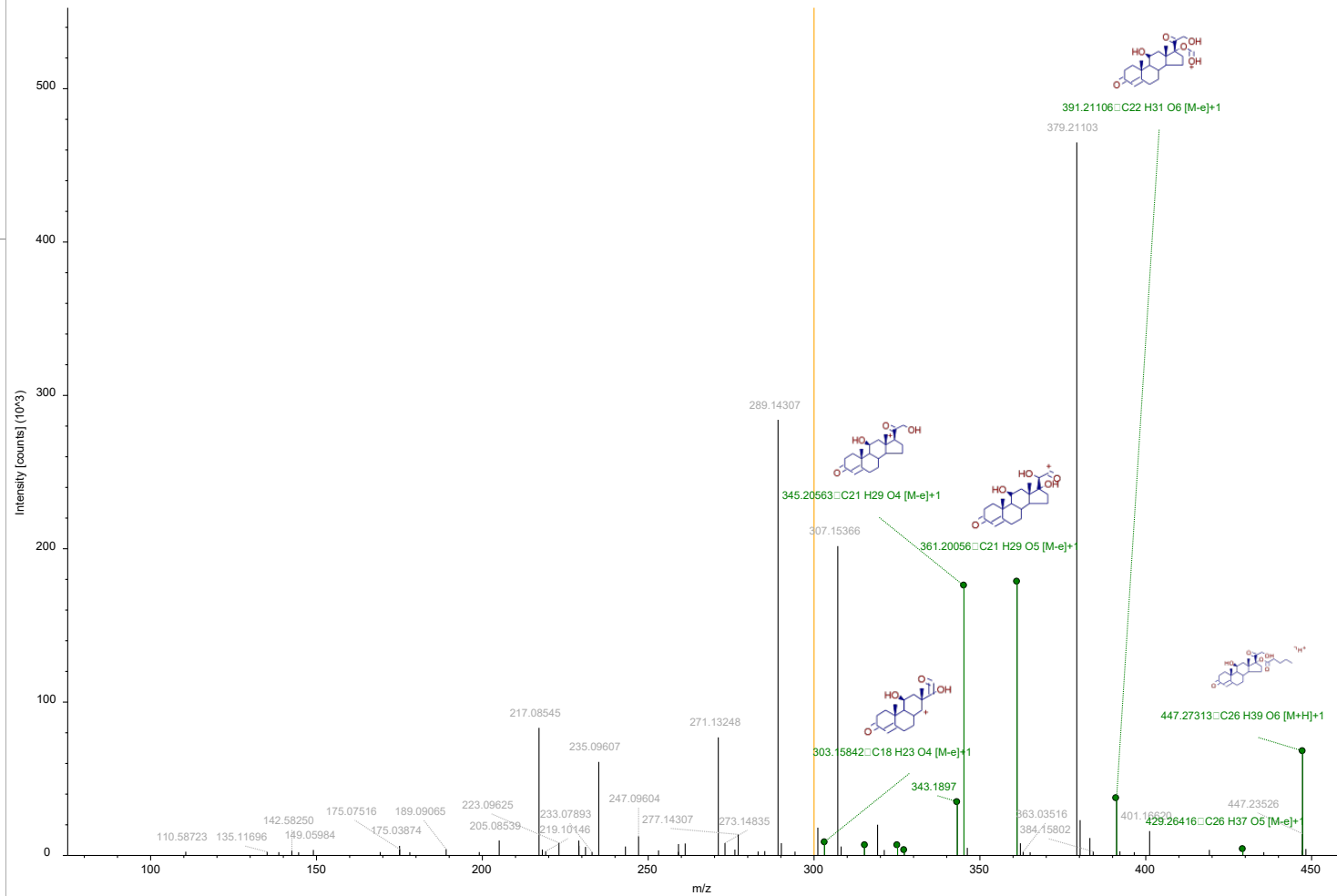
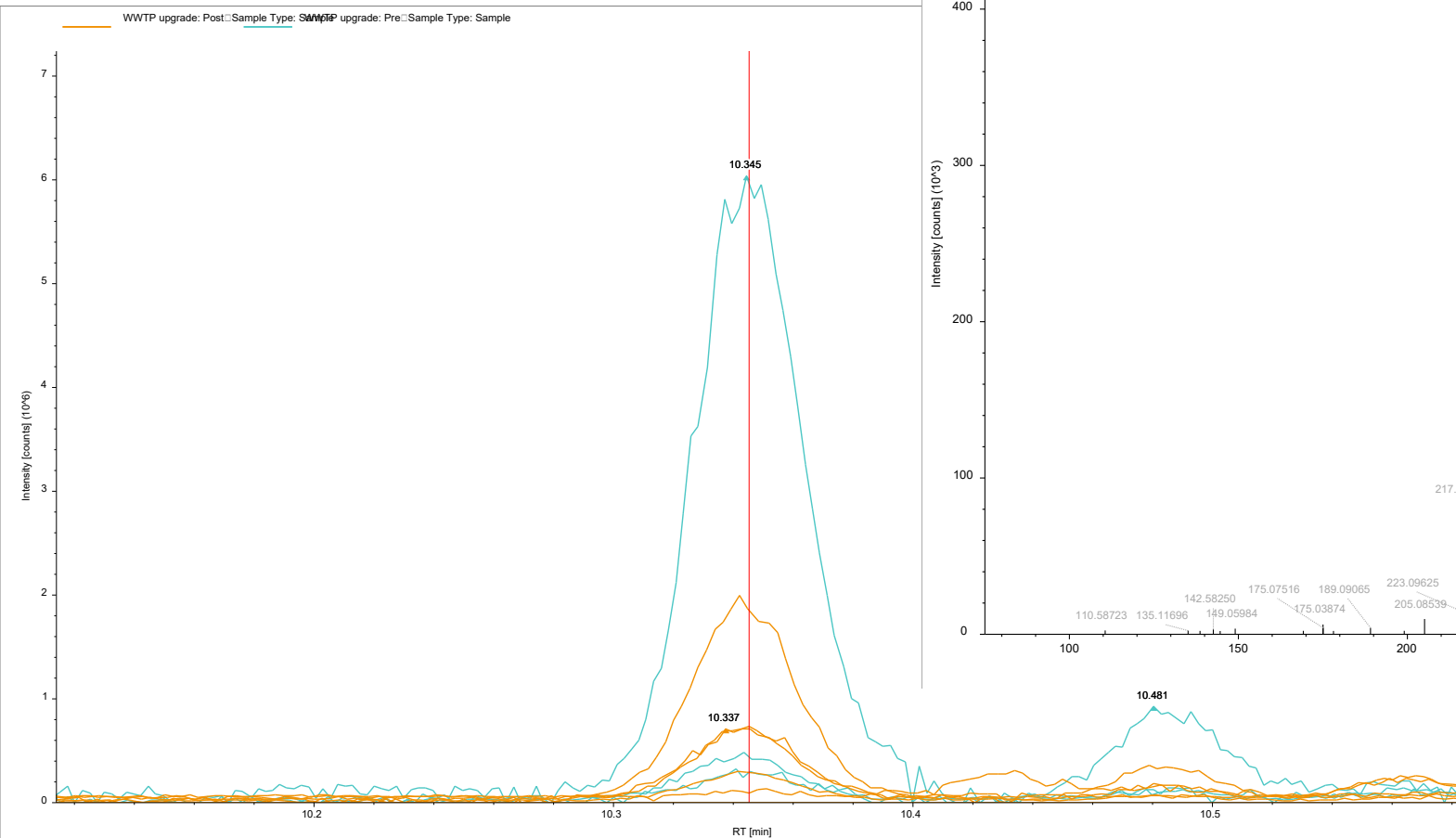




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200303\_18 (F18) #2238, RT=10.349 min, MS2, FTMS (+), (HCD, DDA, 447.2742@(15;30;45), +1)   FISH Coverage: 10 Matched, 36 Unmatched, 22 Skipped



# Summary

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- Effects-based monitoring a powerful complement to traditional chemical monitoring
- Effects-based methods can support and guide unknowns identification
- If specific targets are known, receptor pulldown assays can be a specific, targeted method to identify bioactive components of mixtures
- Suspect screening viable if receptor agonists are well defined
- Access to lab is back – Continuing pulldown assay and suspect screening work. Targeting mid-2021 for project completion



# Questions?

Brett Blackwell, PhD

USEPA Office of Research and Development

Center for Computational Toxicology and Exposure

Great Lakes Toxicology and Ecology Division

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