

# Image-based high-throughput phenotypic profiling for hazard evaluation of environmental chemicals: Two applications.

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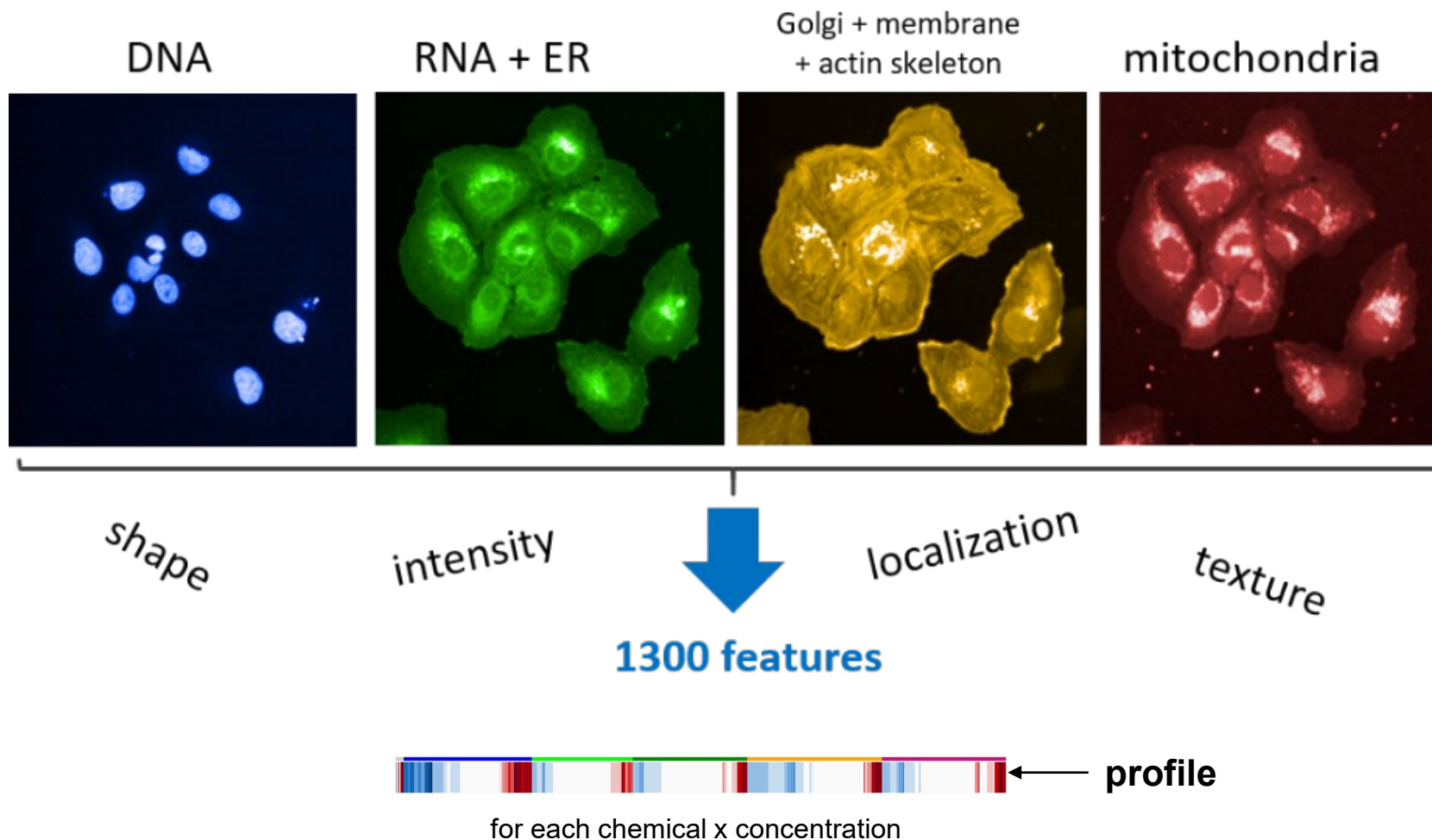
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# High-throughput phenotypic profiling (HTPP)



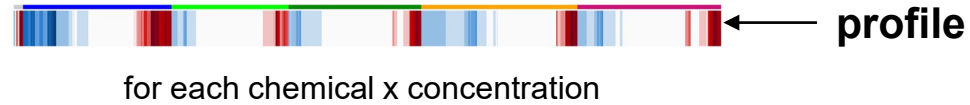
‘Cell Painting’ assay

Gustafsdottir et al. 2013

Bray et al. 2016

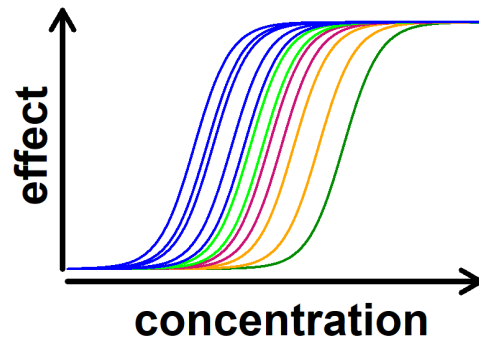
Nyffeler et al. 2020

# Two applications



## Application 1

concentration-response modelling



## Application 2

Chemical A

0	1.80	0	9.69	5.73	0	6.47	-12.84	0	0
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Chemical B

0	0	0	10.00	6.00	1.60	6.47	-15.00	0	0
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Biological similarity  
=  
Pearson correlation

Potency estimate:  
*in vitro* point-of-departure (POD)

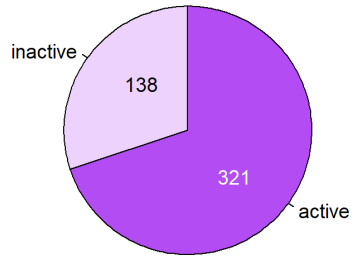
see Nyffeler et al. 2020a + 2020b

Compare profiles with annotated reference chemicals  
→ putative modes-of-action

work in progress

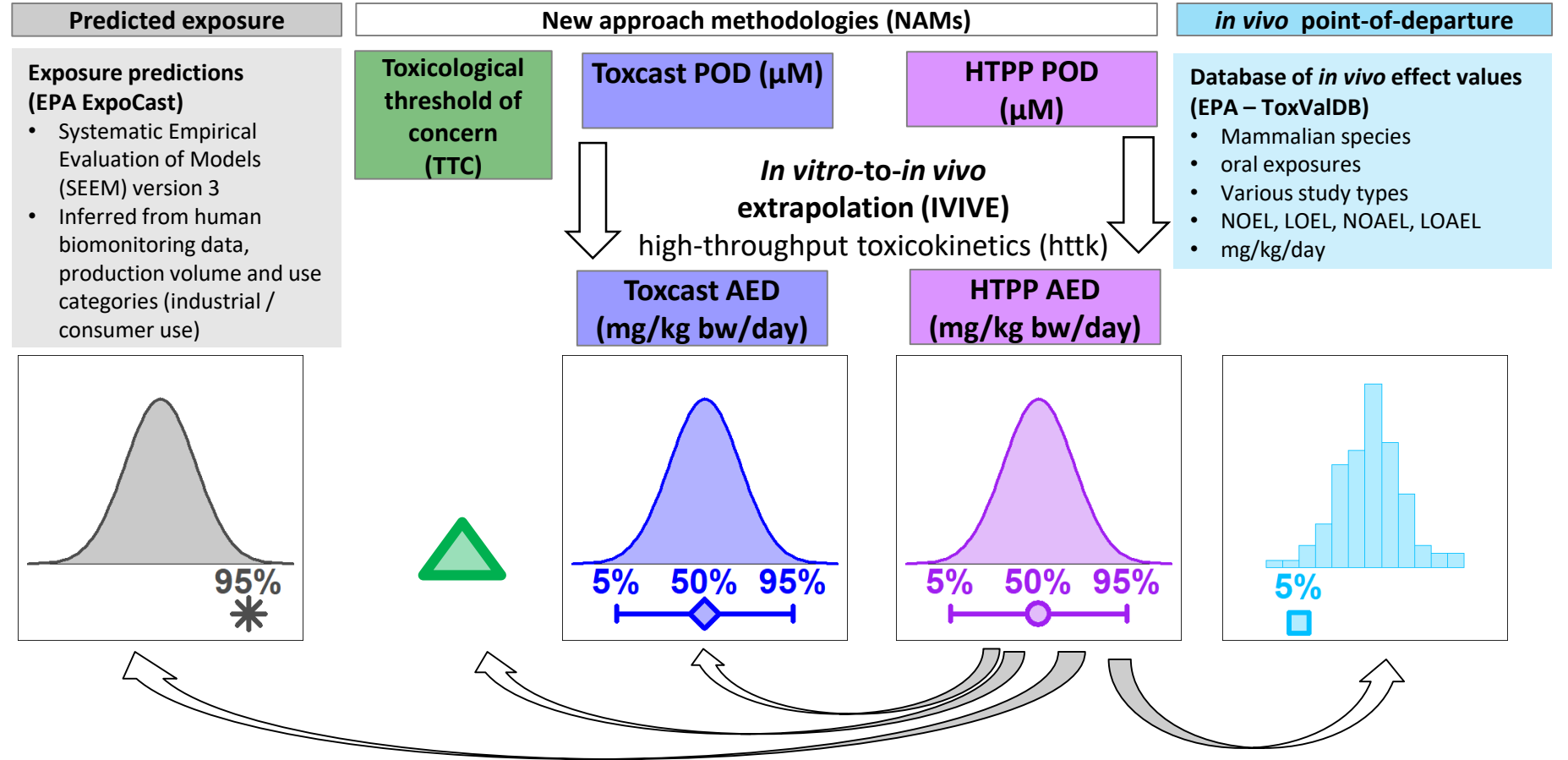
# Comparison to *in vivo* data and exposure

462 chemicals tested



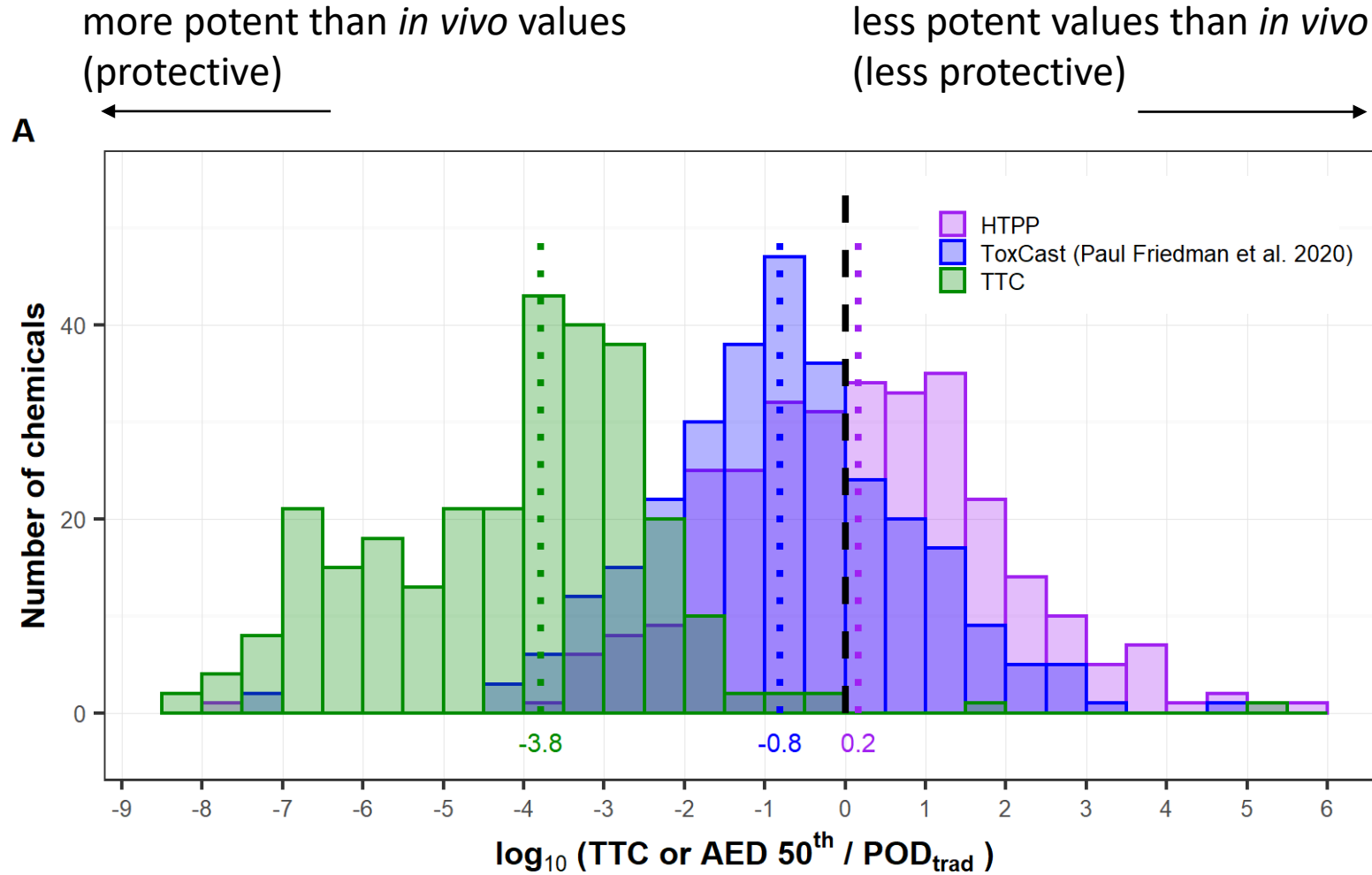
321 chemicals active

in vitro  
point-of-departure



POD: point-of-departure  
AED: administered equivalent dose

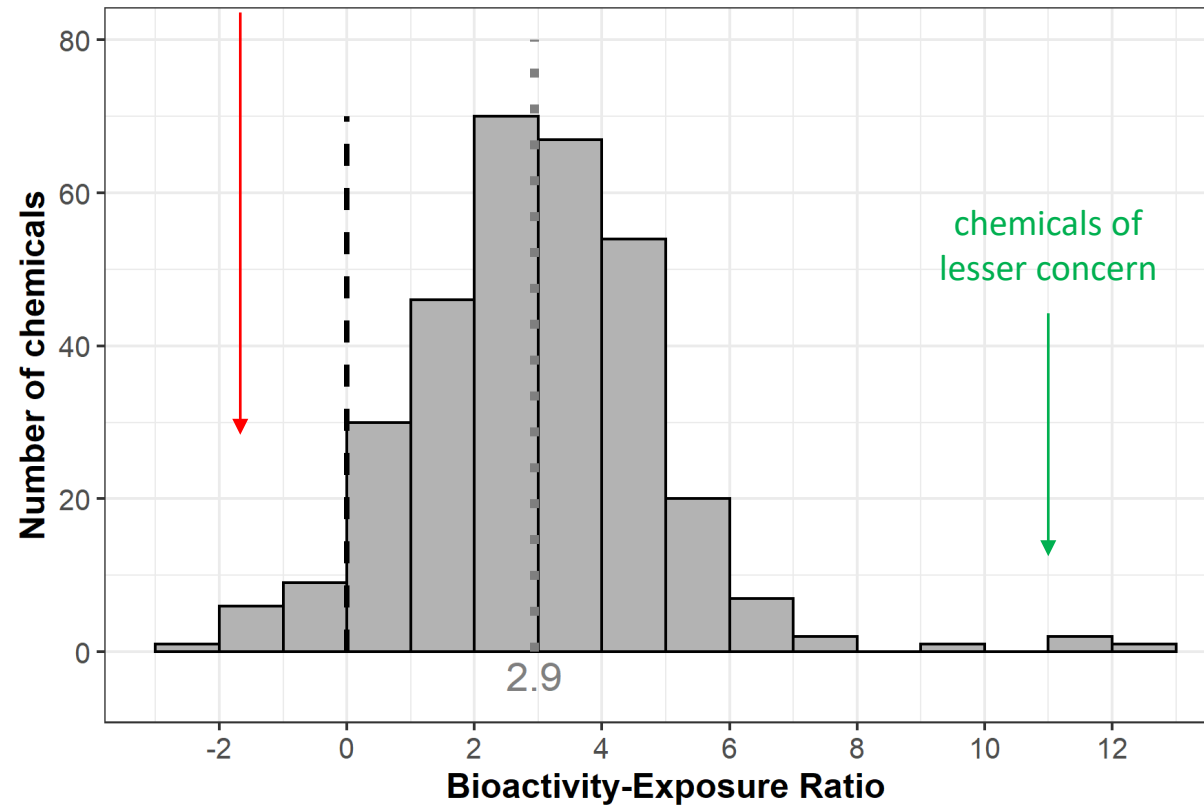
# Comparison to *in vivo* effect values & other NAMs



- ⇒ HTPP AEDs are less potent than ToxCast-derived AEDs and TTC values
- ⇒ 78% (237/303) of HTPP AED are within 2 orders of magnitude of the *in vivo* POD

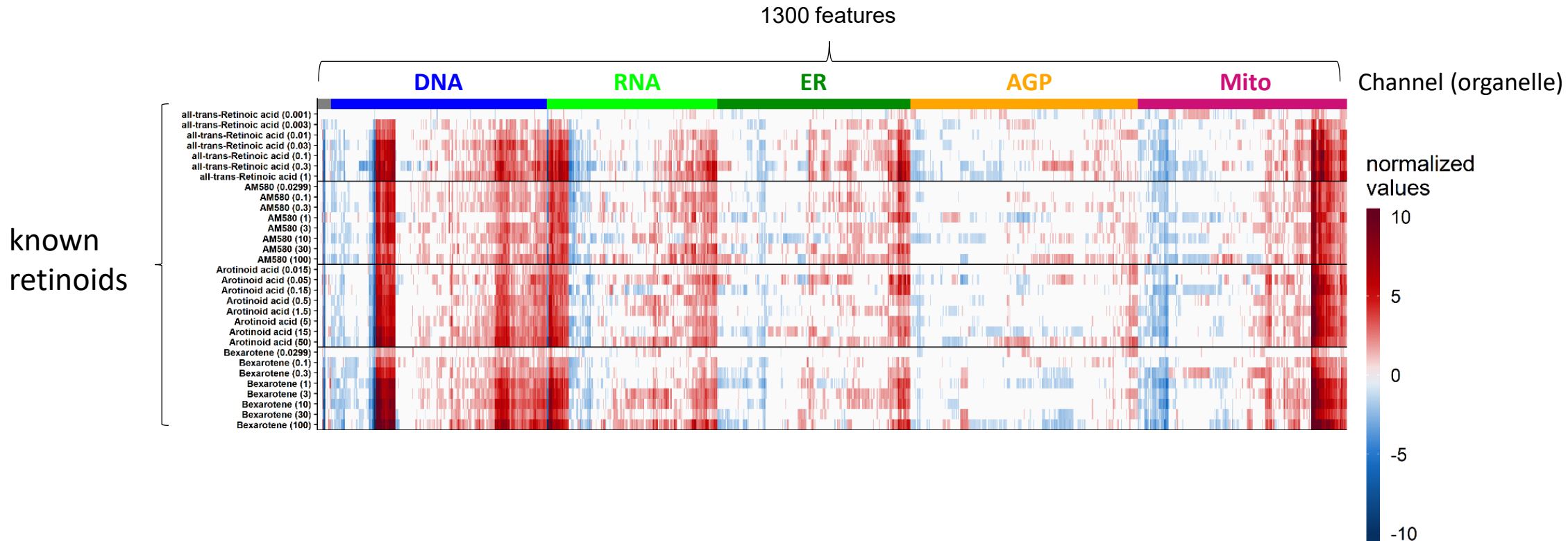
# Comparison to exposure estimates

Potential for humans  
to be exposed to  
bioactive concentrations



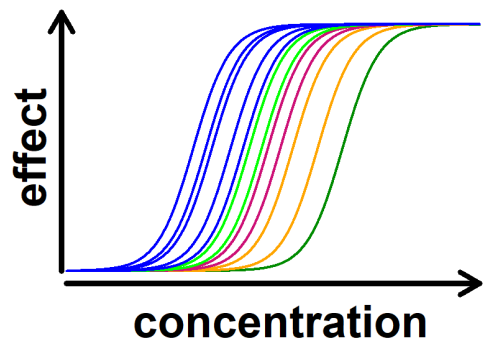
- ⇒ for 49% of chemicals, predicted exposure is > 1000x lower than estimated bioactivity
- ⇒ for 5.1% (16/316) of chemicals, the BER was negative, indicating a potential for humans to be exposed to bioactive concentrations of these chemicals

# Use of phenotypic profiles to discern putative modes-of-action (MOA)

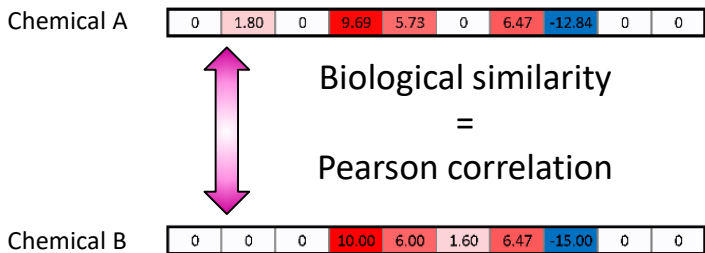


- ⇒ 5 test chemicals had similar profiles to known retinoids
- ⇒ 4 of them were inactive in ToxCast retinoic acid signaling assay  
→ potential for HTPP to give complementary results to existing assays

# Conclusions



1. HTPP *in vitro* potencies can be used for prioritizing of chemicals based on inferred bioactivity in relation to predicted human exposure



2. Potential for HTPP to give complementary information to existing assays





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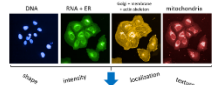
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## What is phenotypic profiling?



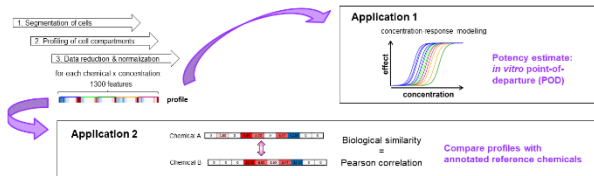
- Image-based phenotypic profiling is a chemical screening method that measures a large variety of morphological features of individual cells in *in vitro* cultures.
- No requirement for *a priori* knowledge of molecular targets.
- May be used as an efficient and cost-effective method for evaluating chemical bioactivity.

## Method: High-throughput phenotypic profiling (HTPP)

### 1. Chemical exposure & labeling

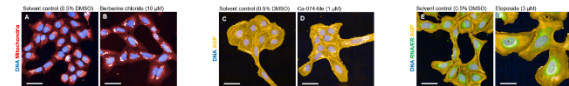


### 2. Data analysis



## Results

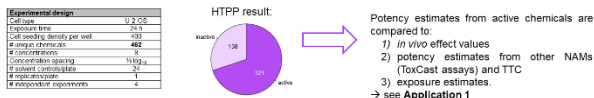
### Examples



### High-throughput screening

#### Screen 1: 462 bioactive chemicals

Chemicals from our inventory were selected that had information about *in vivo* bioactivity and for which toxicokinetic measurements and exposure estimates were available (Paul Friedman et al. 2020). A majority of chemicals are pesticides, the remaining chemicals are drugs, food additives and industrial chemicals.



#### Screen 2: 1201 ToxCast chemicals

Chemicals from the ToxCast phase 1 and 2 libraries were selected. Of the 1201 chemicals, 179 chemicals had molecular targets annotated in the RefChemDB database (Judson et al. 2019).



## References

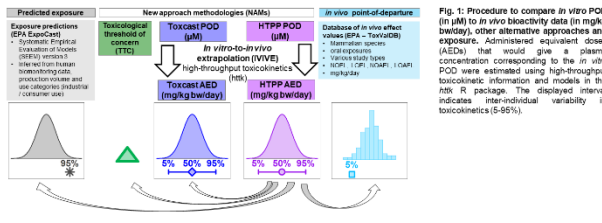
- Judson, et al. 2019. Workflow for defining reference chemicals for assessing performance of *in vitro* assays. *ALTEX* 2019;36(2):261-276.
- Nyffeler et al. 2020. Bioactivity screening using imaging-based high-throughput phenotypic profiling. *Toxicol Appl Pharmacol* 389, 114876.
- Nyffeler J, Haggard DE, Willis C, et al. Comparison of approaches for determining bioactivity hits from high-dimensional profiling data. *SLAS Discovery* August 2020.
- Paul Friedman, et al. 2020. Examining the utility of *in vitro* bioactivity as a conservative point of departure: a case study. *Toxicol Sci* 173(1): 202-225.

## Abbreviations

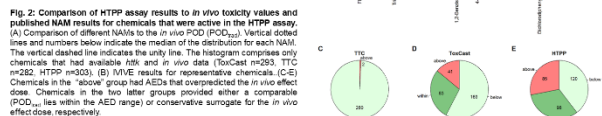
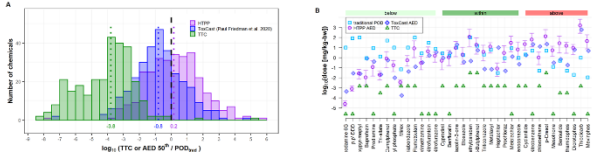
AED	Administered equivalent dose
BER	Bioactivity exposure ratio
HTPP	High-throughput phenotypic profiling
NIVE	<i>In vitro</i> to <i>in vivo</i> extrapolation
MOA	Mode-of-action
NAM	New approach methodology
POD	Point-of-departure
TTC	Threshold of toxicological concern

## Application 1: Estimation of potency thresholds for chemical bioactivity

*In vitro*-to-*in vivo* extrapolation (NIVE) was performed using reverse dosimetry to extrapolate the HTPP POD to an administered equivalent dose (AED) to compare it with *in vivo* effect values, other alternative methods and to exposure predictions:



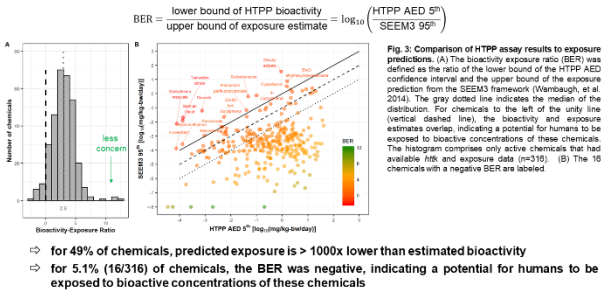
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- 78% (237/303) of HTPP AED are within 2 orders of magnitude of the *in vivo* POD

### Comparison to exposure estimates

HTPP AEDs were compared to exposure predictions and the bioactivity exposure ratio was calculated as follows:



- HTPP *in vitro* potencies can be used for prioritizing of chemicals based on inferred bioactivity in relation to predicted human exposure

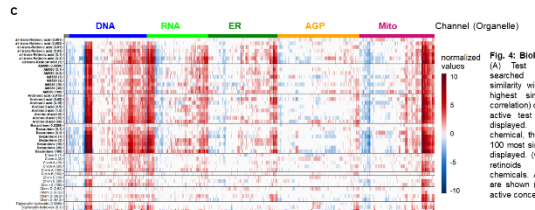
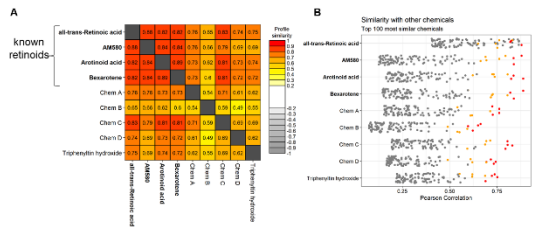
### Next steps:

- Test chemicals in multiple cell types to increase biological coverage

## Application 2: Use of phenotypic profiles to discern putative modes-of-action (MOA)

### Identify putative retinoids

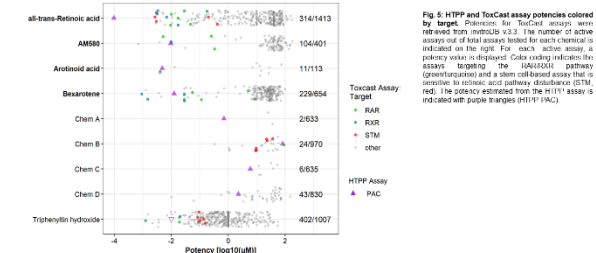
Among all 1201 tested chemicals, five chemicals displayed high biological similarity to the known retinoids:



- Five test chemicals were highly similar to the known retinoids but did not display similarity with other chemicals.
- These chemicals have similar profiles albeit a weaker effect as known retinoids

### Compare activity with ToxCast assays

For these chemicals, all *in vitro* test results from the ToxCast assay suite were retrieved:



- Only one chemical (triphenyltin hydroxide) was active in the ToxCast RAR/RXR assays. It is a known RXR agonist.
- One additional chemical was active in the STM assay
- Three chemicals were tested in RAR/RXR/STM assays but were inactive
- potential for HTPP to give complementary information to existing assays

- Five test chemicals had a similar phenotype as known retinoids

### Next steps:

- Confirm the bioactivity of these chemicals in an orthogonal assay (qPCR)



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