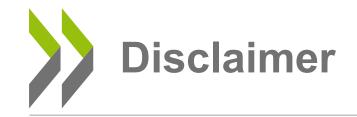
### Guidance Document for Consistent Reporting of 'Omics Data From Various Sources

-Transcriptomics Reporting Framework (TRF) -Metabolomics Reporting Framework (MRF)

Joshua Harrill (US EPA) and Mark Viant (Univ. Birmingham) on behalf of TRF & MRF

Cosmetics Europe Toxicogenomics Meeting October 30<sup>th</sup> 2020





• The views expressed in this presentation are those of the author(s) and do not necessarily represent the views or policies of the U.S. Environmental Protection Agency, nor does mention of trade names or products represent endorsement for use.





To develop frameworks for the standardisation of reporting of 'omics data generation and analysis, to ensure that all of the information required to understand, interpret and reproduce an 'omics experiment and its results are available.

**Purpose:** to ensure that sufficient information is available to enable an evaluation of the quality of the experimental data and interpretation, and support reproducibility.

**NOT** to stipulate the methods of data analysis or interpretation....**Rather**, provide guidance on reporting of information that fosters transparency and reproducibility.

Project Name	Project Leads
Metabolomics Reporting Framework (MRF)	Mark Viant (Univ. Birmingham, UK)
Transcriptomics Reporting Framework (TRF)	Joshua Harrill (USEPA) Carole Yauk (Health Canada)
Optimal Data Analysis Framework (ODAF)	Tim Gant (PHE, UK) Florian Caiment (Univ. Maastricht)
OECD Secretariat	Magda Sachana



# TRF

# TRF Document, Major Topic Areas

#### **TOXICOLOGY EXPERIMENT MODULE:**

- The experiment should be described in sufficient detail that would allow another researcher to replicate the experiment.
- Adapted from existing sources
- Information in this section is <u>independent</u> of 'omics platform

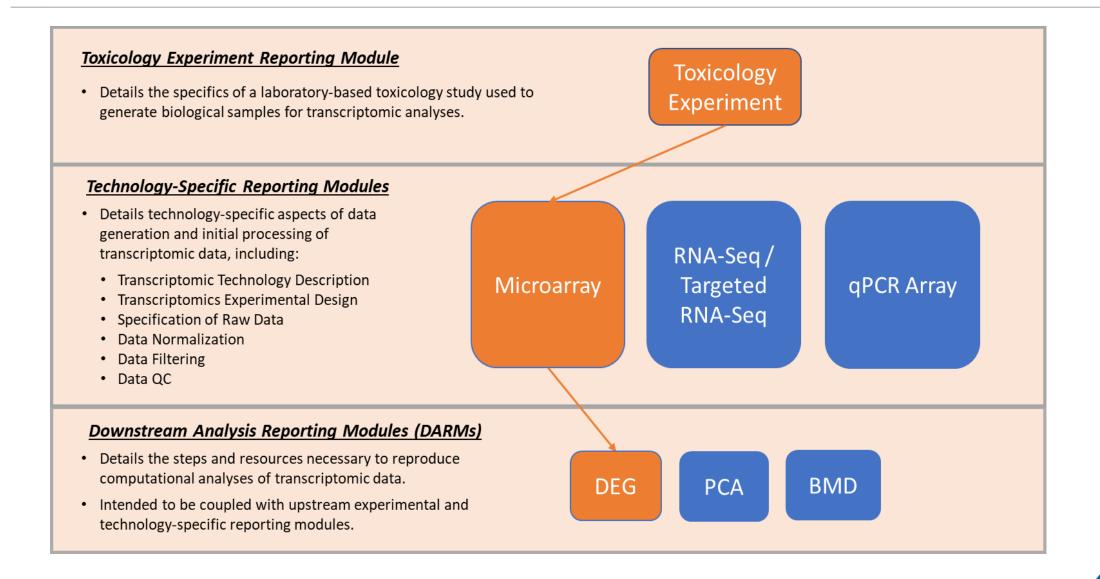
#### **PROCESSING AND ANALYSIS OF 'OMICS DATA MODULES:**

- The transcriptomics technology, sample processing procedures, methods used to collect raw data and methods used to generate processed data.
- Described in Gant et al. (2017).
- Information in this section is <u>dependent</u> on 'omics platform

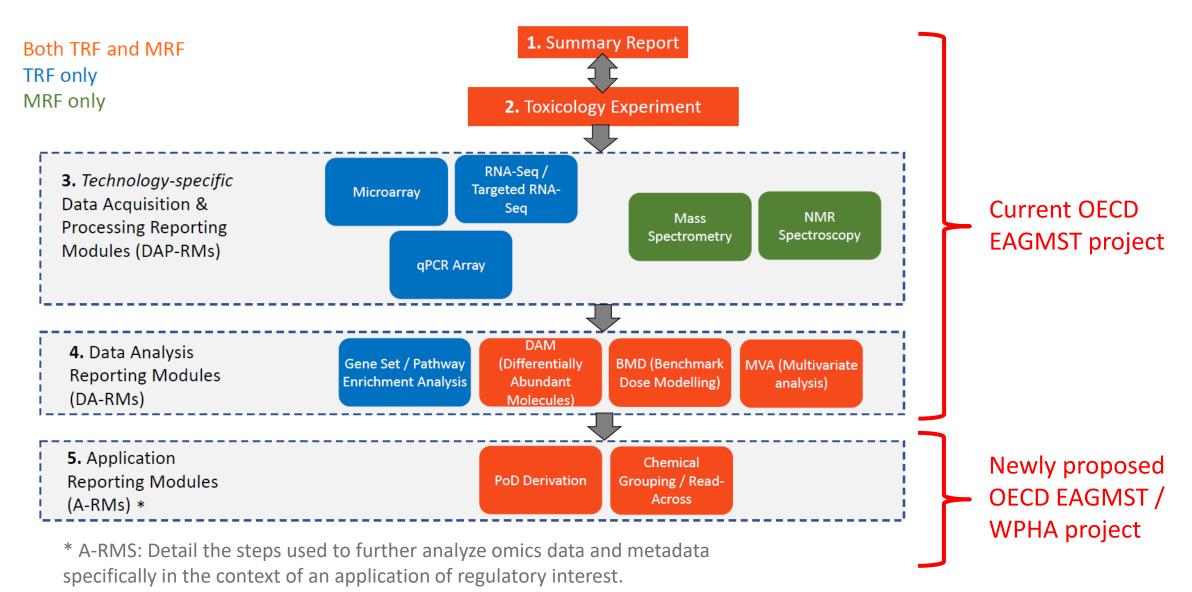
#### DOWNSTREAM ANALYSIS REPORTING MODULES [DA-RMs]

• Detail the steps and resources necessary to reproduce a computational analysis of the processed data.

TRF: Original Structure



## Modular Structure of Omics Reporting Frameworks Harmonization of TRF and MRF





#### Table of Contents

1. Introduction	4
<ol> <li>Background</li></ol>	4 6
2. Toxicology Experiment Module	
<ul> <li>2.1. Study Rationale</li></ul>	
3. Processing and Analysis of Microarray Data	29
<ul> <li>3.1. Technology</li></ul>	
4. Processing and Analysis of RNA Sequencing Data	45
<ul> <li>4.1. RNA-Seq Technology</li></ul>	
5. Processing and Analysis of Quantitative Reverse-Transcription PCR Data	
<ul> <li>5.1. Sample Processing</li></ul>	57 58

• Stylistic alignment:

- Previous OECD guidance in the biological sciences (where applicable)
- MERIT Project / Metabolomics Reporting Framework (MRF) – *In Progress*

#### • Reporting Format

- Narrative text followed by Reporting Fields
- Excel spreadsheet for reporting
- Consistent vocabulary across modules



1	A		В	C	D
	RELEVANT MODULE	REPORTING	CATEGORY	REPORTING ELEMENT	DESCRIPTION OF ELEMENT
	Summary Report	Study identi	fier	Unique study identifier	Unique code
4					
5		Study ration	ale	Objective of study	Controlled vocabulary: point-of-departure; hazard
6				Background information (supporting the objective)	Free text elaboration of above
7					
8		Links to rela	ted study records	Standardised toxicology dataset (e.g., linked to OE	
9				Omics complete dataset (e.g., linked to MetaboLig	hts, e.g. MTBLSxx
10					
11	Toxicology Experiment Module	Test item (c	hemical) and vehicle	e Test item name	
12				Test item identifier	SMILES or InChIKey
13				Vehicle name	
14					
15		Test system	characteristics		
16		EITHER in	vivo	a. Species	
17				b. Strain	
18				c. Sex	
19				d. Age	
20		OR in vitr	0	a. Cell type	
21				b. Species of origin	
22					
23		Exposure co	nditions	Test item concentration(s)	
24				Route of administration	
25				Schedule (single dose or repeated dosing)	
26				Frequency of repeated dosing	
27				Exposure duration(s)	
28					
		Biological sa	mples	Type (e.g., in vitro: cells, media, etc.; in vivo: cells,	
29				tissue, biofluid, whole organism, etc.)	
				Sample preservation method (e.g., fresh, frozen,	
30 31				paraffin-embedded, etc.)	
32				Number of biological replicates per treatment	
202	TRF platform-specific data	Sample prep	aration	Sample preparation method (e.g., RNA extraction	
	acquistion and processing Reportin			method, cell lysis, etc.)	
33	Module	-6			
34					
	Technology		Type (e.g., data acquisition and processing module	6	
35				used - DNA microarray, RNA-seq, etc.)	
				Manufacturer(s) and model(s) (e.g., Agilent microa	rray,
36				etc.)	
	Study Summar	v revised	Experiment	Processing microarray data DARM DEG	÷

#### • Executive summary included

#### Modular

- Flexible/expandable
- Add new modules as new platforms or analytical approaches released
- Mandatory/optional reporting fields

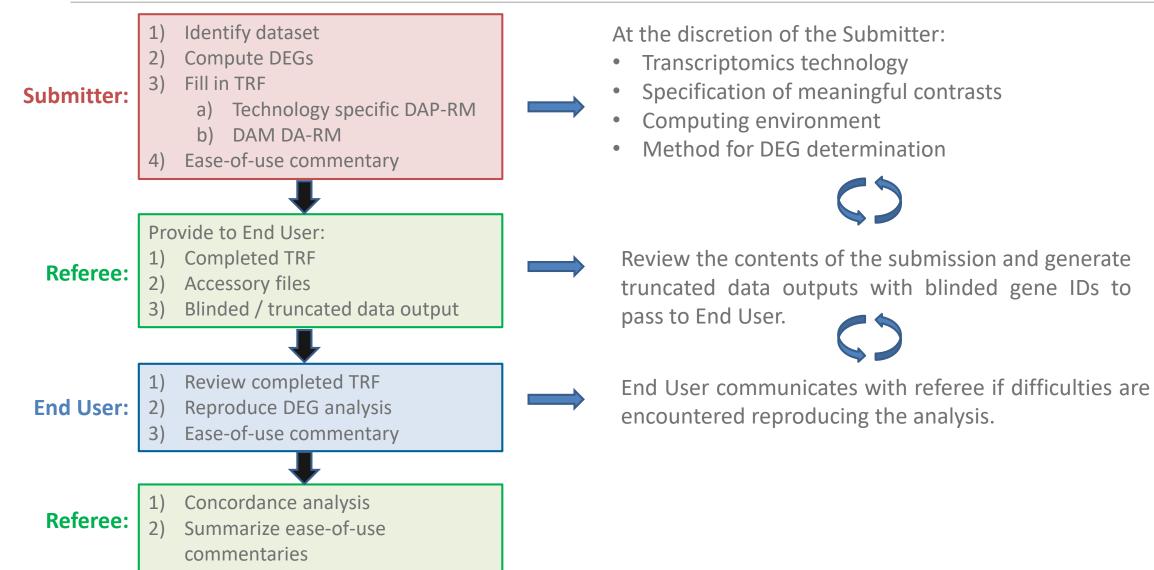
#### • Database compatibility

 Summary, Experiment and DAM DARM harmonized with MRF



Module Name	Module Development Lead	Status					
Introduction	Joshua Harrill (US EPA) Carole Yauk (U Ottawa)	Complete					
Study Summary	Carole Yauk (U Ottawa)	Complete					
Toxicology Experiment Module (TEM)	Raffaella Corvi (JRC)	Complete					
Technology Specific Data Acquisition and Processing Reporting Modules (DAP-RM)							
Microarray	Vikrant Vijay (NCTR)	Complete					
RNA-Seq / Targeted RNA-Seq	Florian Caiment (U Maastricht)	Complete					
qPCR Array	Jason O'Brien (ECCC)	In Process					
Data Analysis	Data Analysis Reporting Modules (DA-RM)						
Differentially Abundant Molecules (DAM)	Lyle Burgoon (ERDC)	Complete					
Benchmark Dose Modeling (BMD)	Scott Auerbach (NIH DNTP)	In Process					
Gene Set / Pathway Enrichment	TBD	Pending					



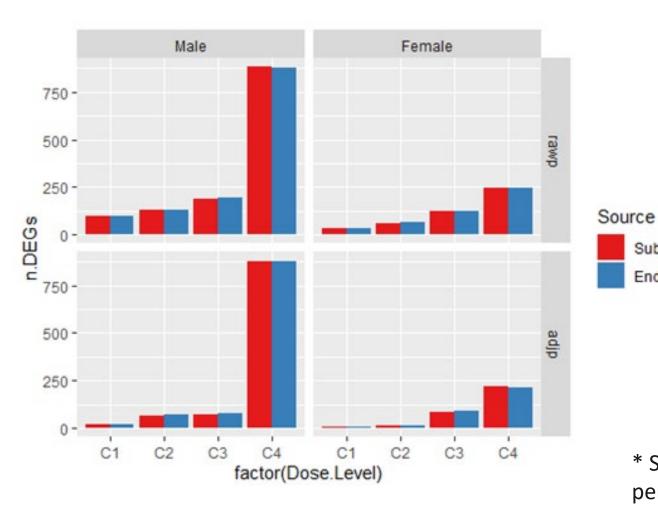




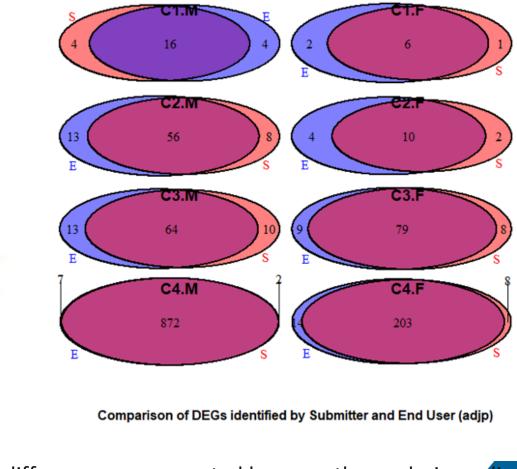
Platform	Study Description	DAM Method	Computing Environment	Submitter	End User	Status
Agilent Microarray	Four point concentration-response of furan in male and female Fisher rat liver (GEO GSE62805)	Submitter's Choice	R	Andrew Williams (Health Canada)	Leah Wehmas (US EPA)	Complete
Affymetrix Microarray	Comparison of PFOA responses in livers of 129S1/SvImJ wild-type and PPAR-alpha null mice (GEO GSE9786)	Submitter's Choice	Partek Flow	Beena Vallanat (US EPA)	Alison Harrill (HHS DNTP	In Process
RNA-Seq	Three point concentration- response of hexabromocyclododecane in male and female Fisher rat liver (PRJNA395549)	ODAF	R	Matt Meier (Health Canada)	Brian Chorley (US EPA)	In Process
RNA-Seq	TBD	Submitter's Choice	R	Natalia Garcia- Reyero (MS State IGBB)	Andrew Williams (Health Canada)	In Process



#### **Number of Differentially Expressed Genes**



#### **Overlap of Differentially Expressed Genes**



Submitter

End User

\* Small differences are expected because the analysis applied permutation-based *p*-value calculations



#### General findings relating to ability to reproduce analyses:

1. Analyses in open source computing environments (R, Python, etc.)

- a) Much easier for an end user knowledgeable in coding languages to reproduce because they come with an "instruction manual" (i.e. the analysis script or notebook)
- b) Details in the reporting fields become somewhat less critical for reproducing the analysis secondary to the scripts
- c) There are also no financial or licensing barriers with regards to accessing the tools
- d) Issue: users may not have sufficient expertise with open source computing environments

#### 2. Analyses using **freeware analysis softwares or web applications** (BMDExpress; iDep; kallisto)

- a) These types of software are more user friendly and require less technical or statistical expertise to use
- b) No "pay wall" barrier that would prevent an end user from accessing such tools.
- c) Reproducibility depends on clear and precise reporting in the TRF documentation as well as provision of a configuration file or some other configuration snapshot that the end user could follow. **NEEDS TO BE TESTED**
- 3. Analyses using **proprietary software** (Partek, Ingenuity, etc.):
- a) End user needs access to the same software (and maybe even version)
- b) "pay wall" issues.
- c) Reproducibility depends on precise reporting in the TRF documentation as well as provision of a configuration or workflow that the end user could follow.



### Additional case studies:

- More developer and user feedback and participation.
- Testing different analytical platforms:
  - Open computing environments versus
  - Web applications
- "Test Driving" reporting modules in development
  - qPCR case studies
  - BMD case studies



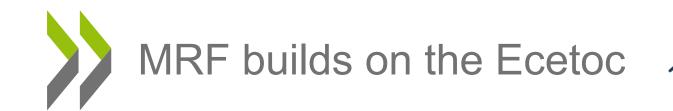


# MRF



# OECD MRF Expert Group

Title	Identity	Roles
Expert Group	Tripartite Industry Government / Regulator Academic ca. 15 very active members, ca. 10 'observers'	-Contribute expert knowledge wherever possible across the whole MRF guidance document -Ensure consistency of whole document
Facilitator	Mark Viant [ Univ. Birmingham ]	<ul> <li>Foster discussion</li> <li>Monitor progress in accordance with project timeline</li> <li>Ensure consistency with TRF</li> </ul>
Administrator	David Epps [ Univ. Birmingham ]	-Meeting organisation
OECD Secretariat	Magda Sachana	-Project administration / OECD liaison







Published July 2019

#### PERSPECTIVE

https://doi.org/10.1038/s41467-019-10900-y

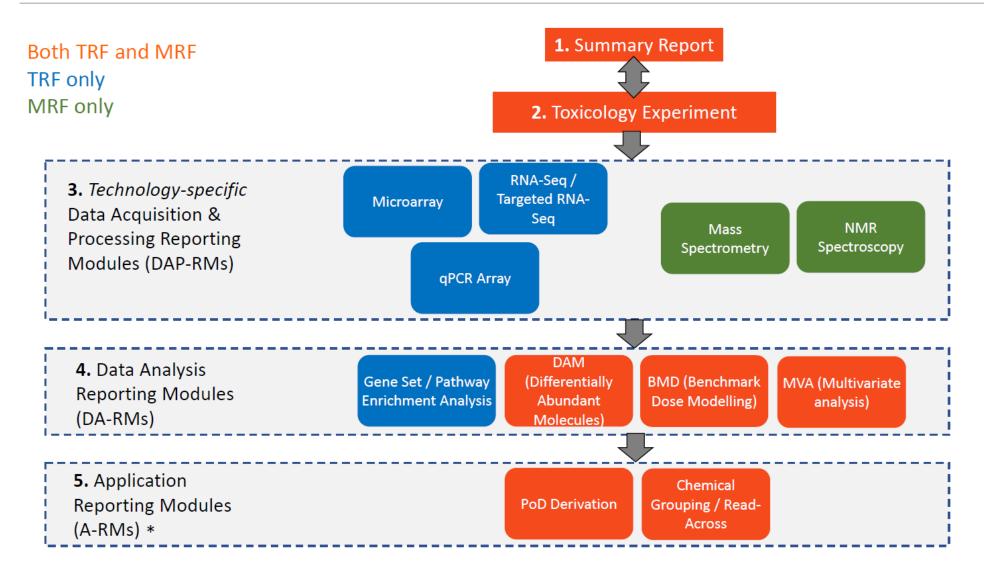
OPEN

# Use cases, best practice and reporting standards for metabolomics in regulatory toxicology

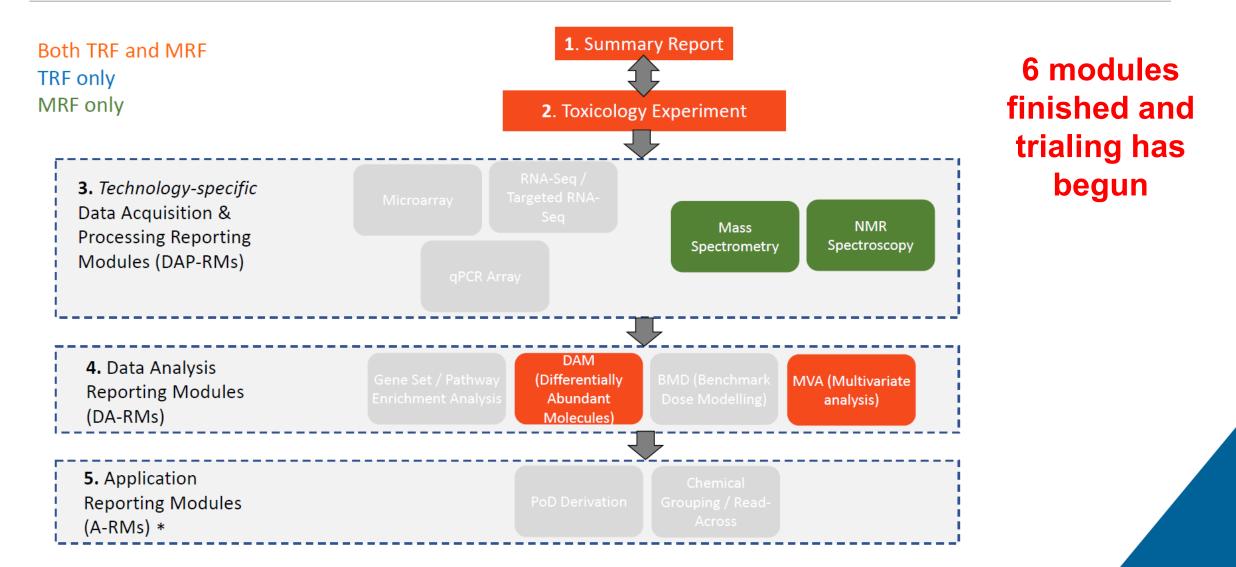
```
Mark R. Viant <sup>1,12</sup>, Timothy M.D. Ebbels <sup>2,12</sup>, Richard D. Beger <sup>3</sup>,
Drew R. Ekman <sup>4</sup>, David J.T. Epps<sup>1</sup>, Hennicke Kamp <sup>5</sup>, Pim E.G. Leonards <sup>6</sup>,
George D. Loizou <sup>7</sup>, James I. MacRae <sup>8</sup>, Bennard van Ravenzwaay<sup>5</sup>,
Philippe Rocca-Serra <sup>9</sup>, Reza M. Salek <sup>10</sup>, Tilmann Walk <sup>11</sup> &
Ralf J.M. Weber <sup>1</sup>
```

https://www.nature.com/articles/s41467-019-10900-y

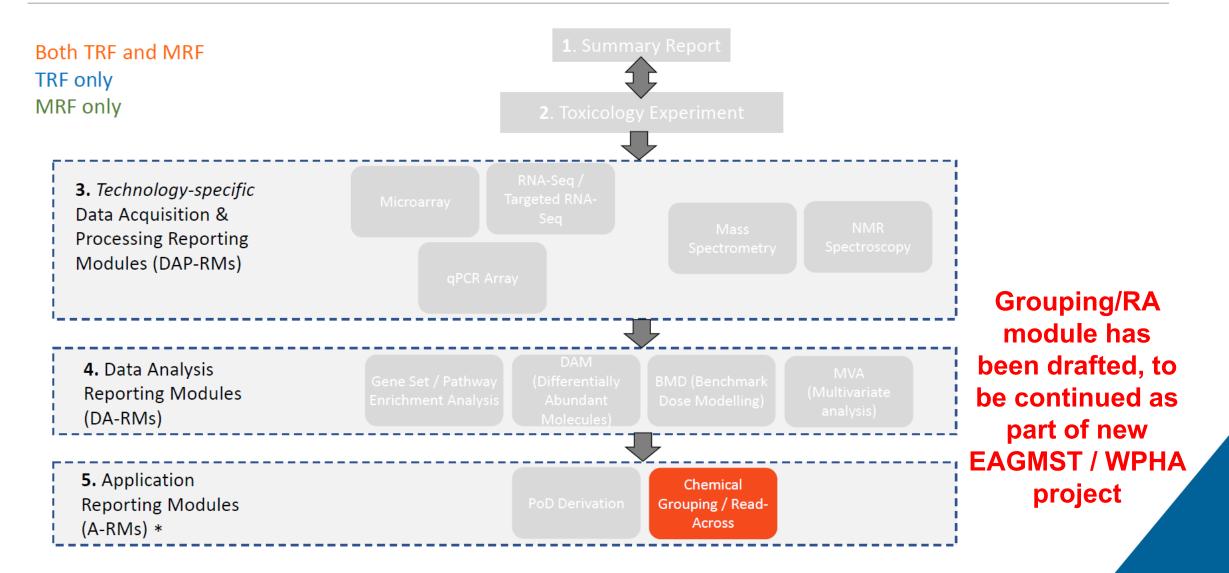


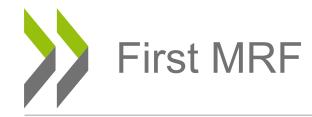












#### **Table of Contents**

#### 1. Introduction

- 1.1 Background, Objective and Scope
- 1.2 Modular Structure of MRF
  - 1. Summary Report (SR)
  - 2. Toxicology Experiment Module (TEM)
  - 3. Data Acquisition & Processing Reporting Modules (DAPRMs)
  - 4. Data Analysis Reporting Modules (DARMs)
  - 5. Application Reporting Modules (ARMs)
- 1.3 Example Use Cases using Modular Reporting
- 2. Summary Report
- 3. Toxicology Experiment Module
- 4. MRF Technology-specific Data Acquisition & Processing Reporting Modules
  - 4.1 Mass Spectrometry Metabolomics Module
  - 4.2 NMR Spectroscopy Metabolomics Module
- 5. Data Analysis Reporting Modules
  - 5.1 Discovery of Differentially Abundant Molecules (using univariate analysis) Module
  - 5.2 Multivariate Statistical Analysis Module
- 6. Application Reporting Modules
  - 6.1 Chemical Grouping for Read-Across Module
- 7. References

- Reporting Format
  - Narrative text followed by Reporting Fields
  - Excel spreadsheet for reporting
- Consistent vocabulary across modules
- Database compatibility
- ca. 80 page document



#### MASS SPECTROMETRY METABOLOMICS MODULE:

- Describes the acquisition and data processing for mass spectrometry based metabolomics studies.
- For the first time:
  - Integrated untargeted and targeted metabolite analysis into consistent reporting framework
  - Integrated a range of platforms (LC-MS, GC-MS, direct infusion MS) into one module
  - Developed consistent terminology describing relative quantification, semi-quantification and absolute metabolite quantification, and the description of reference standards
  - Defined new terminology for MS assay names including "hybrid" assays that combine targeted and untargeted analyses.



#### MASS SPECTROMETRY METABOLOMICS MODULE:

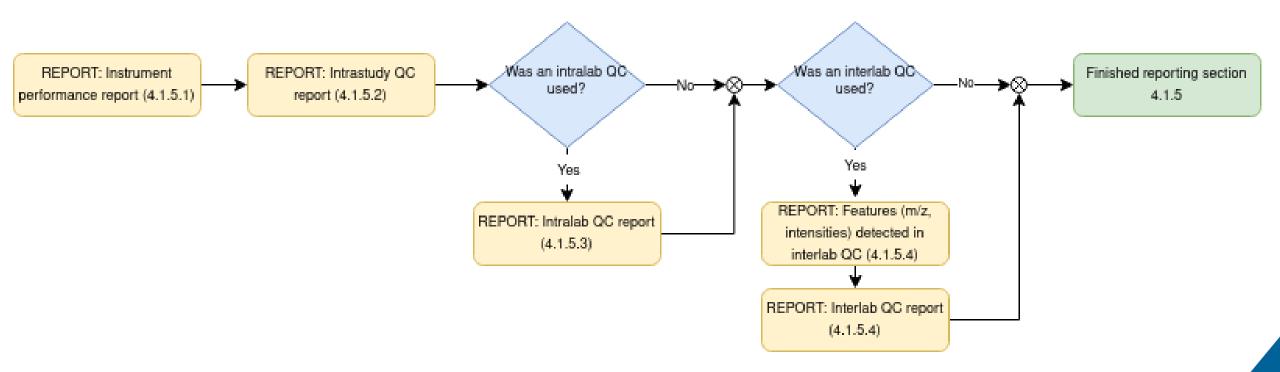
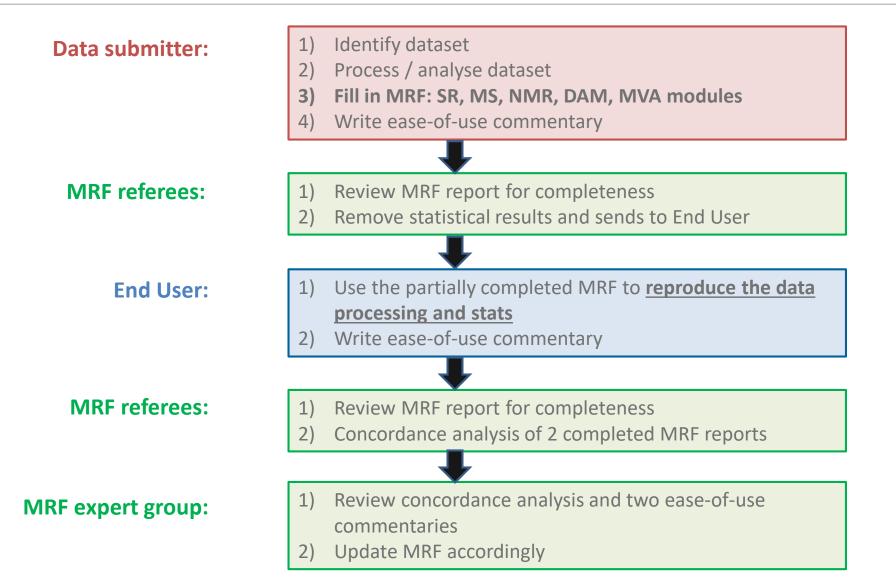


Figure 5: Workflow for mass spectrometry metabolomics reporting -Demonstration of quality of metabolomics analysis







Mass spectrometry metabolomics trial - Underway

- Data submitter: David Crizer (National Toxicology Program, US)
  - 5-day rodent assay, plasma samples, thujone exposure



- MRF referees: Oliver Schmitz (BASF, DE), Pim Leonards (VU University, NL), Aniko Kende (Syngenta, UK)
- End user: Tom Lawson (Michabo Health Science, UK)

#### NMR spectroscopy metabolomics trial - Now starting

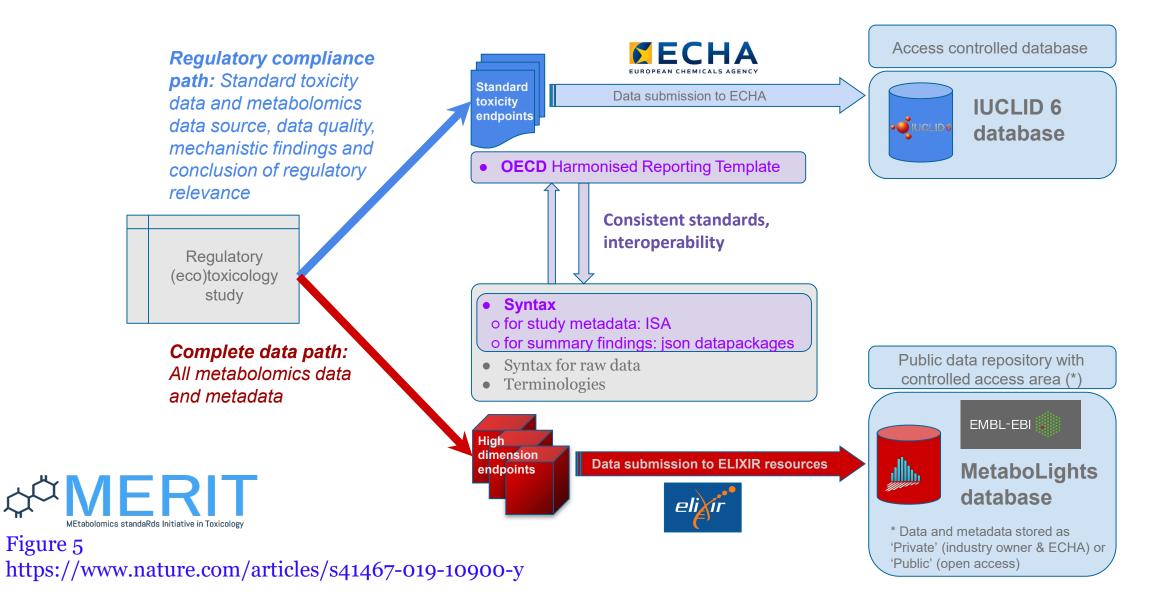
- Data submitter: Fabien Jourdan, Nicolas Cabaton, Cécile Canlet (INRA, FR)
  - Mouse study, brain tissue, bisphenol A exposure
- MRF referees: Drew Ekman (EPA, US), Mark Viant (University of Birmingham, UK)
- End user: Tracey Schock (NIST, US)

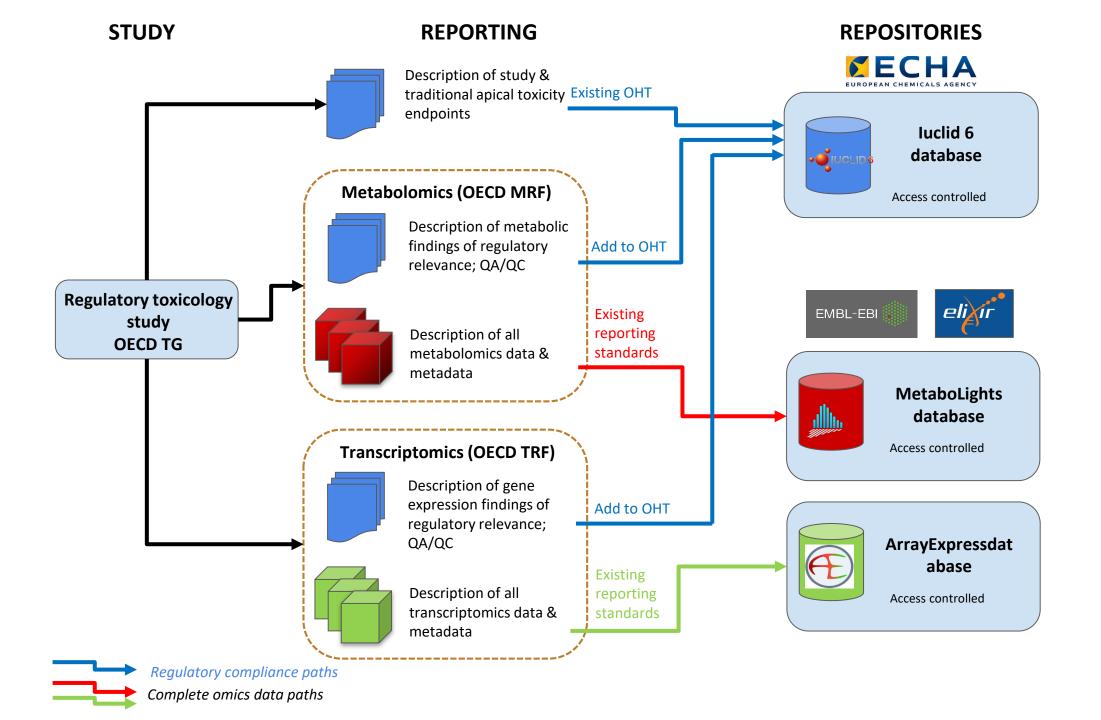


How and where to report (metabol)omics data from a regulatory toxicology study in Europe?

#### **Chemical Safety Study & Assessment**

Sponsor:Chemical companyProcess:In-house or outsourced study encompassing several data acquisition modalitiesOutput:Dataset [Study Metadata Descriptions, Raw Data, Processed Data, Findings and Reports]





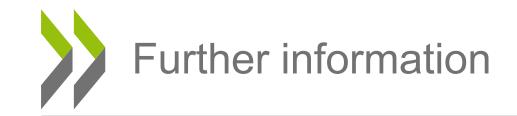
### Michabo Health Science – ECHA project

### Mapping to IUCLID and MetaboLights

REPORTING ELEMENT	MANDATORY / OPTIONAL	DESCRIPTION OF ELEMENT AND CONTROLLED VOCABULARY	REPORT TO SPECIALIST REPOSITORY? (termed 'Complete Data path', e.g. to EBI MetaboLights or NIH Metabolomics Workbench)	l i i i i i i i i i i i i i i i i i i i	REPORT TO REGULATOR? POTENTIAL COMPONEN (termed 'Regulatory FOR IUCLID Compliance path', e.g. to IUCLID)
Extraction method accord description	Mandatas	Eroo tout	v	Extraction material	N (included in Summer-
Extraction method general description	Mandatory	Free text	- V	Extraction protocol	N (included in Summary
Solvent(s) used,	Mandatory	Free text	T V	Extraction protocol	N
Means of agitation/maceration	Mandatory	Free text	V	Extraction protocol	N
Temperatures and times Post extraction handling, e.g., storage	Mandatory Mandatory	Free text Free text	T V	Extraction protocol Extraction protocol	N
Derivatization method general description	Mandatory if used	Free text	Y	Extraction protocol	Ν
Reagents and reaction (including incubation	Mandatory if used	Free text	Y	Extraction protocol	N
Clean-up/partitioning (if used).	Mandatory if used	Free text	Y	Extraction protocol	N
Evaporation and reconstitution method general	Mandatory	Free text	Υ	Extraction protocol	Ν
Final reconstitution solvent(s) and final volume (if	Mandatory if used	Free text	Y	Extraction protocol	Ν
Storage temperature (if relevant)	Mandatory if used	Free text	Y	Extraction protocol	N
Duration of reconstituted extracts (if relevant)	Mandatory if used	Free text	Y	Extraction protocol	Ν
Quality assessment reference standard general	Mandatory if used	col	Y	Extraction protocol	Ν
Report in Table A4.1(1): Quality assessment	Mandatory if used	Table	Y	Separate file	Ν



- MRF draft (version 1): completed
- MRF trialling: on-going, deadline April 2021
- Revised MRF submitted to OECD EAGMST for formal review: June 2021
- Extension of TRF + MRF to include *Application Reporting Modules*: new OECD EAGMST-WPHA proposal asap



- TRF: Josh Harrill (<u>Harrill.Joshua@epa.gov</u>),
   Carole Yauk (<u>Carole.Yauk@uottawa.ca</u>)
- MRF: Mark Viant (<u>M.Viant@bham.ac.uk</u>)

OECD: Magda Sachana (<u>Magdalini.Sachana@oecd.org</u>)

