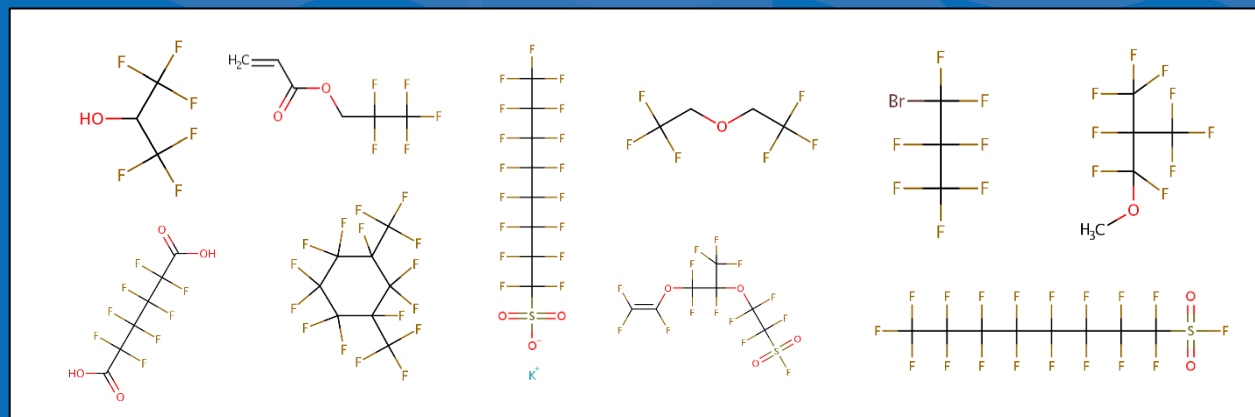


A Chemical Category-Based Approach for Selecting and Screening PFAS for Toxicity and Toxicokinetic Testing



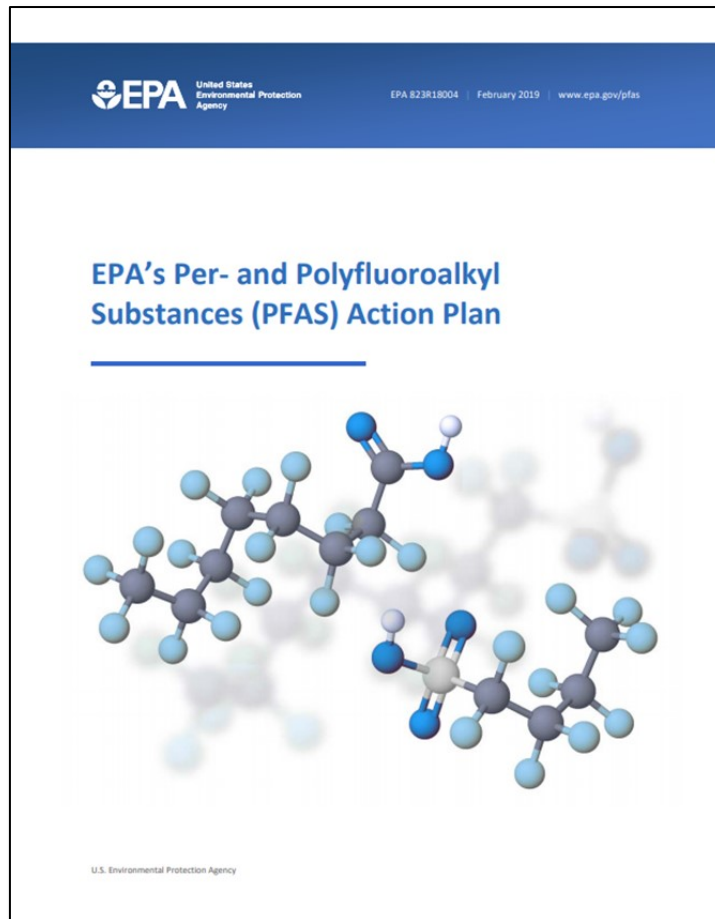
Grace Patlewicz
Center for Computational Toxicology & Exposure (CCTE), US EPA

Background and Importance of the Problem



Bottom line is that we cannot readily dig our way out using only traditional testing approaches...

EPA is Using New Approach Methods (NAMs) to Help Fill Information Gaps



Research Area 1: What are the human health and ecological effects of exposure to PFAS?


- **Using computational toxicology approaches to fill in gaps.** For the many PFAS for which published peer-reviewed data are not currently available, the EPA plans to use new approaches such as high throughput and computational approaches to explore different chemical categories of PFAS, to inform hazard effects characterization, and to promote prioritization of chemicals for further testing. These data will be useful for filling gaps in understanding the toxicity of those PFAS with little to no available data. *In the near term*, the EPA intends to complete assays for a representative set of 150 PFAS chemicals, load the data into the [CompTox Chemicals Dashboard](#) for access, and provide peer-reviewed guidance for stakeholders on the use and application of the information. *In the long term*, the EPA will continue research on methods for using these data to support risk assessments using New Approach Methods (NAMs) such as read-across and transcriptomics, and to make inferences about the toxicity of PFAS mixtures which commonly occur in real world exposures. The EPA plans to collaborate with NIEHS and universities to lead the science in this area and work with universities, industry, and other government agencies to develop the technology and chemical standards needed to conduct this research.


But, It All Starts With Chemistry...

Curating Names, Structures, and Identifiers

November 26, 2015

Dataset Open Access

 **OECD**
Organisation for Economic Co-operation and Development

 **EPA**
United States
Environmental Protection
Agency

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4 May 2018

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List Acronym	List Name	Last Updated	Number of Chemicals	List Description
EPAPFASNONDW	PFAS[EPA: New EPA Method Non-Drinking Water	2019-04-17	24	EPA is developing and validating a new method for detecting these PFAS in non-drinking water sources.
EPAPFASRESEARCH	PFAS[EPA: EPA PFAS Research List	2019-05-03	165	The list of PFAS EPA is currently researching using various scientific approaches.
EPAPFASRL	PFAS[EPA: Cross-Agency Research List	2017-11-16	199	EPAPFASRL is a manually curated listing of mainly straight-chain and branched PFAS (Per- & Poly-fluorinated alkyl substances) compiled from various internal, literature and public sources by EPA researchers and program office representatives.

10,776 PFAS (as of August 2021) captured on the PFASSTRUCT list

PFASMASTER	PFAS Master List of PFAS Substances	2019-11-11	7866	PFASMASTER is a consolidated list of PFAS substances spanning and bounded by the below lists of current interest to researchers and regulators worldwide.
PFASNTREV19	PFAS: PFAS in Non-Target HRMS Studies (Liu et al 2019)	2019-04-17	127	List of PFAS substances detected in non-target HRMS reviewed by Liu et al 2019
PFASOECD	PFAS: Listed in OECD Global Database	2018-05-16	4729	OECD released a New Comprehensive Global Database of Per- and Polyfluoroalkyl Substances, (PFASs) listing more than 4700 new PFAS
PFASOECDNA	PFAS[NORMAN: List of PFAS from the OECD Curated by Nikiforos Alygizakis	2019-05-19	3213	List of PFAS released by the OECD, provided by Zhanyun Wang, curated and mapped to structures by Nikiforos Alygizakis

<< < 1 2 3 > >>

Showing 11 to 20 of 22 records

e_Acronym (correct or uious)	Unique_Acronym
d	5:3 PFQA
YSE, MeFOSE	NMeFOSE
Byproduct 2	PFESA Byproduct 2
	PFPeS_ion
	APFPeS

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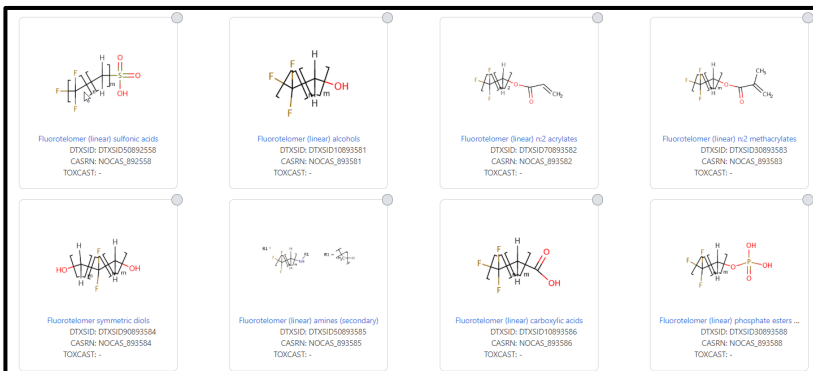
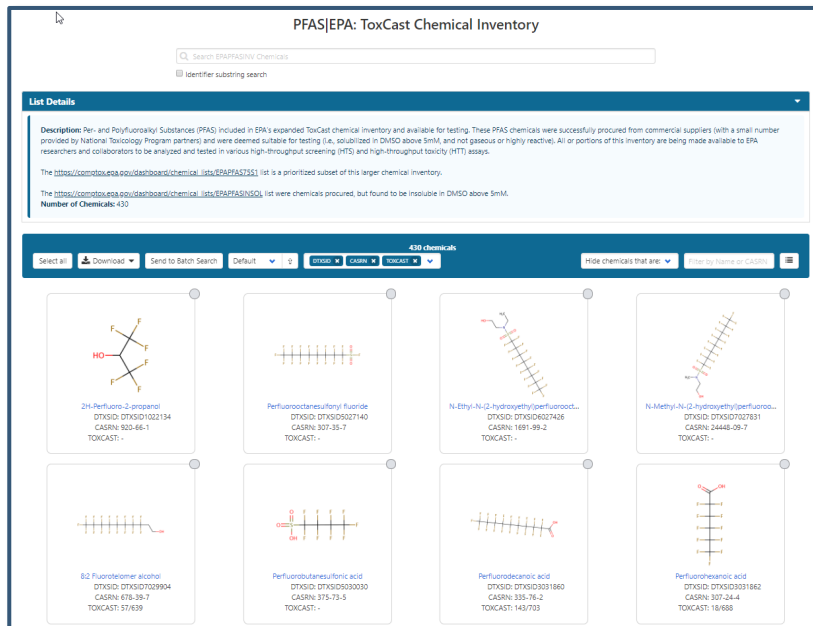


kemikalieinspektionen.se

DTXSID40881350	4,8-Dioxo-3H-perfluorononanoic acid	919005-14-4	2,2,3-Trifluoro-3-(1,1,2,2,3,3-hexafluoro-3-yl)propanoic acid	919005-14-4	ADONA	ADONA parent acid
DTXSID00874026	Ammonium 4,8-dioxo-3H-perfluorononanoate	919005-14-4	Ammonium 2,2,3-trifluoro-3-(1,1,2,2,3,3-hexafluoro-3-yl)propanoate	919005-14-4	ADONA	ADONA
DTXSID3037707	Potassium perfluorobutanesulfonate	29420-49-3	Potassium perfluoro-1-butanedisulfonate	375-73-5	PFBS	PFBS-K
DTXSID5030030	Perfluorobutanesulfonic acid	375-73-5	Perfluorobutanesulfonic acid	375-73-5	PFBS	PFBS
DTXSID60873015	Perfluorobutanesulfonate	45187-15-3	Perfluorobutanesulfonate	375-73-5	PFBS	PFBS_ion
DTXSID3040148	Perfluorodecanesulfonic acid	335-77-3	Perfluorodecanesulfonic acid	335-77-3	PFDS	PFDS
DTXSID00873014	Perfluorodecanesulfonate	126105-34-8	Perfluorodecanesulfonate	335-77-3	PFDS	PFDS_ion
DTXSID60892443	Sodium perfluorodecanesulfonate	2806-15-7	Sodium perfluoro-1-decanedisulfonate	335-77-3	PFDS	PFDS-Na

Offices and Regions

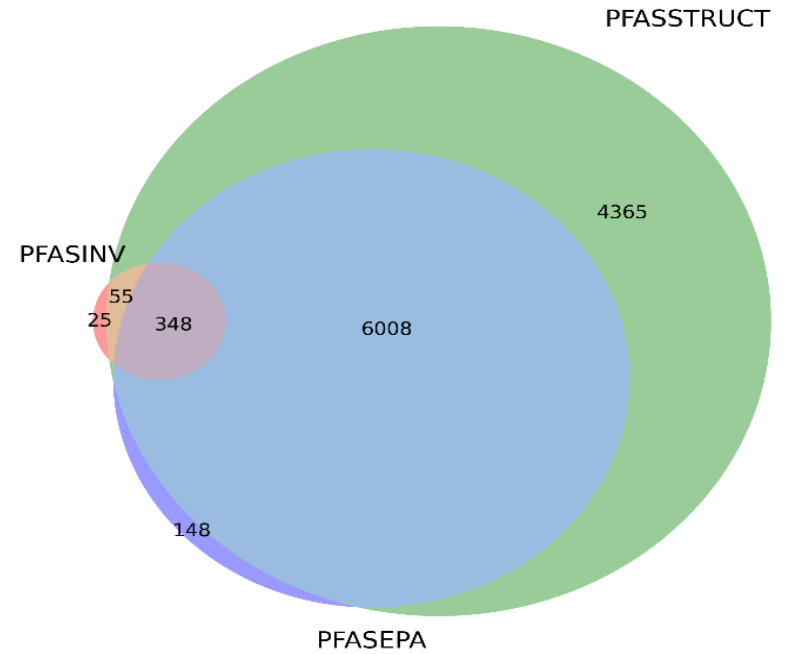
Assembled a PFAS Chemical Library for Research and Methods Development



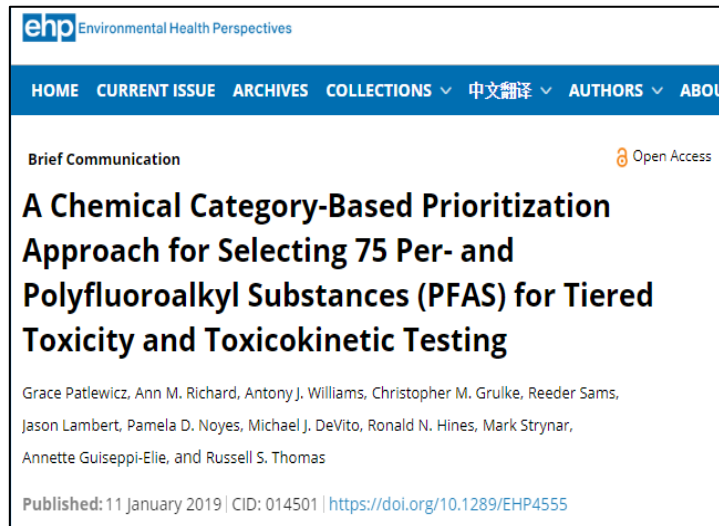
- Attempted to procure ~3,000 based on chemical diversity, Agency priorities, and other considerations
- Obtained 480 total unique chemicals
 - 430/480 soluble in DMSO (90%)
 - 54/75 soluble in water (72%) (incl. only 3 DMSO insolubles)
- Issues with sample stability and volatility
- Categories initially assigned based on three approaches
 - Buck et al., 2011 categories
 - Markush categories
 - OECD categories

PFAS List Overlap

	OECD	PFAS STRUCT	PFAS 430INV	PFAS150
OECD	4729			
PFASSTRUCT	3723	10776		
PFAS430INV	310	407	428	
PFAS150	119	139	146	146



Selecting a Subset of PFAS for Tiered Toxicity and Toxicokinetic Testing

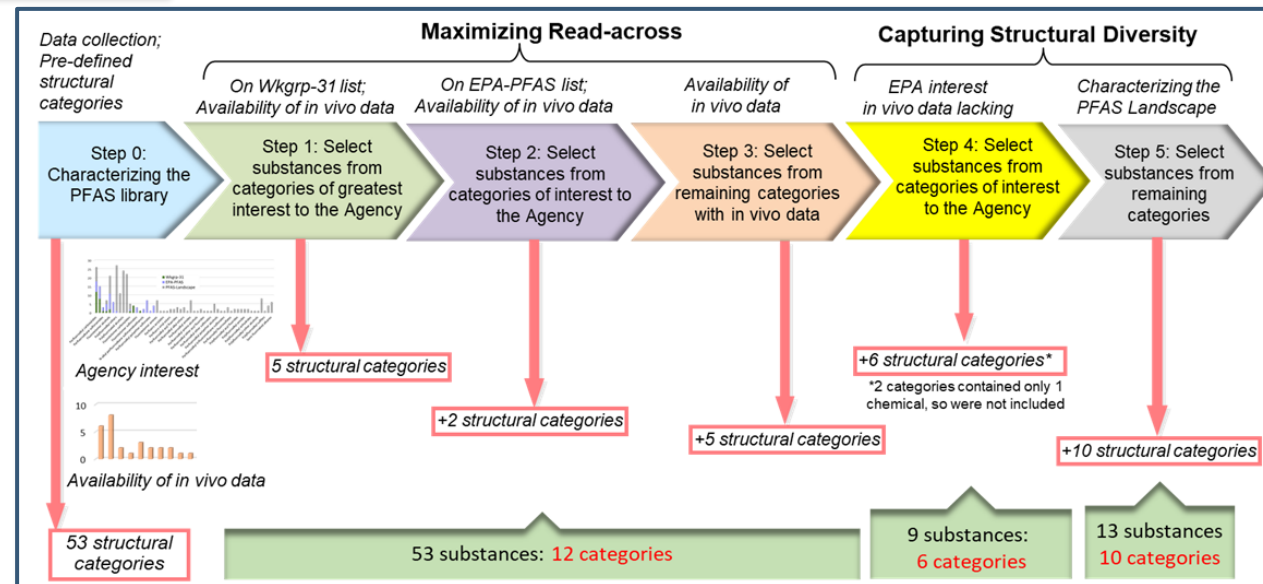


Goals:

- Generate data to support development and refinement of categories and read-across evaluation
- Incorporate substances of interest to Agency
- Characterise mechanistic and toxicokinetic properties of the broader PFAS landscape

Selected 150 PFAS in two phases representing 83 different categories

- 9 categories with > 3 members
- Lots of singletons



In Vitro Toxicity and Toxicokinetic Testing

Toxicological Response	Assay	Assay Endpoints	Purpose
Developmental Toxicity	Zebrafish embryo assay	Fertilisation, lethality, and structural defects	Assess potential teratogenicity
Immunotoxicity	Bioseek Diversity Plus	Protein biomarkers across multiple primary cell types	Measure potential disease and immune responses
Mitochondrial Toxicity	Mitochondrial membrane potential (HepaRG)	Mitochondrial membrane potential	Measure mitochondrial health and function
Developmental Neurotoxicity	Microelectrode array assay (rat primary neurons)	Neuronal electrical activity	Impacts on neuron function
Endocrine Disruption	ACEA real-time cell proliferation assay (T47D)	Cell proliferation	Measure ER activity
General Toxicity	Attagene cis- and trans-Factorial assay (HepG2)	Nuclear receptor and transcription factor activation	Activation of key receptors and transcription factors involved in hepatotoxicity
	High-throughput transcriptomic assay (multiple cell types)	Cellular mRNA	Measures changes in important biological pathways
	High-throughput phenotypic profiling (multiple cell types)	Nuclear, endoplasmic reticulum, nucleoli, golgi, plasma membrane, cytoskeleton, and mitochondria morphology	Changes in cellular organelles and general morphology

Toxicokinetic Parameter	Assay	Assay Endpoints	Purpose
Intrinsic hepatic clearance	Hepatocyte stability assay (primary human hepatocytes)	Time course metabolism of parent chemical	Measure metabolic breakdown by the liver
Plasma protein binding	Ultracentrifugation assay	Fraction of chemical not bound to plasma protein	Measure amount of free chemical in the blood

Objectives

- To inform
 - Chemical Category and Read-across approaches
 - Bioactive Dose Level (BDL) Approach (*in vitro* to *in vivo* extrapolation to define administered dose equivalent (ADE) values)

In order to:

Translate learnings to make inferences for a broader landscape of PFAS

Initially use structural categories to evaluate the degree of concordance in NAM results (per technology) within categories and across categories as a means to qualitatively and quantitatively infer *in vivo* toxicity

Characterising PFAS using structural categories

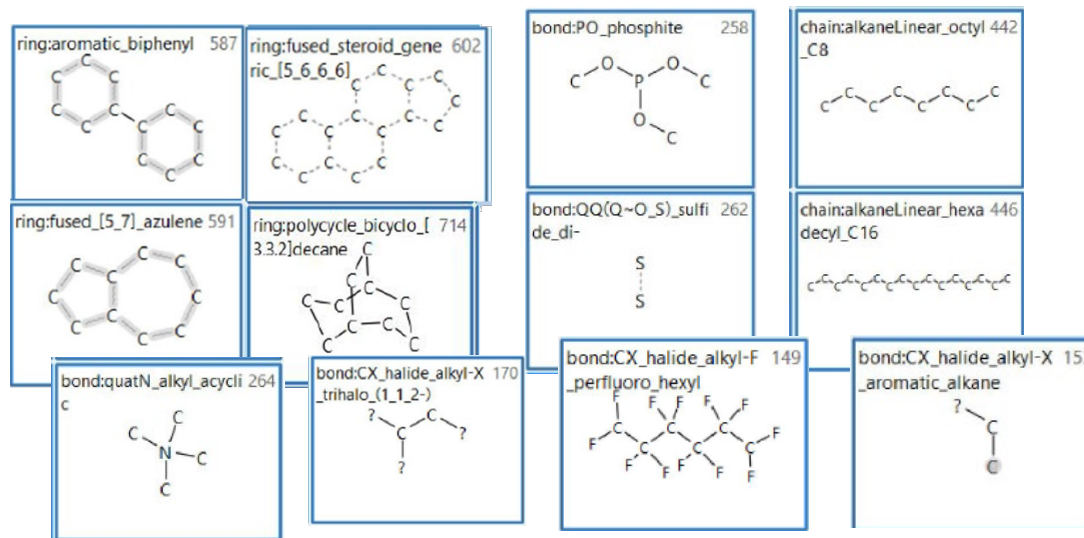
- Structural categories were assigned by visual inspection and whilst nominally consistent since only one individual was making the assignments, the approach was prone to error and not easily reproducible.
- The assignments provided by OECD were similar in their genesis - they were manually assigned by the same person.
- Indeed, authors of many of the published literature studies on PFAS have often ended up deriving bespoke naming conventions for categories which leads to the generation of a lot of parallel nomenclature that differ, creating unintended barriers to effective communication among scientists
- Urgent need exists to develop a reproducible & objective means of developing structure-based categories

PFAS Structure-based Categorisation

- Reconcile the different structural categories schemes initially used by creating a harmonised set of structure-based categories
- Category assignments should be computationally generated from structure only → reproducible, transferable, standardised, extendable
- Permits nested & overlapping categories such that categories can be tailored to different datasets (i.e. the various NAM data streams being generated) and decision contexts

PFAS Structure-based Categorisation: ToxPrints

- Publicly available tools exist to generate & download ToxPrints e.g. ChemoTyper, CompTox Chemicals Dashboard
- Provides excellent coverage of PFAS chemical space
- Nested, hierarchical nature lends itself to creating flexible categories tailored to problem at hand, i.e., “fit for purpose”
- Can augment with computed structure properties (s.a., MW, size, etc.)
- Intuitive, easy to work with



ToxPrints:

- ✓ 729 chemical features
- ✓ Chemically interpretable
- ✓ Coverage of diverse chemistry
- ✓ Includes scaffolds, functional groups, chains, rings, bonding patterns, atom-types

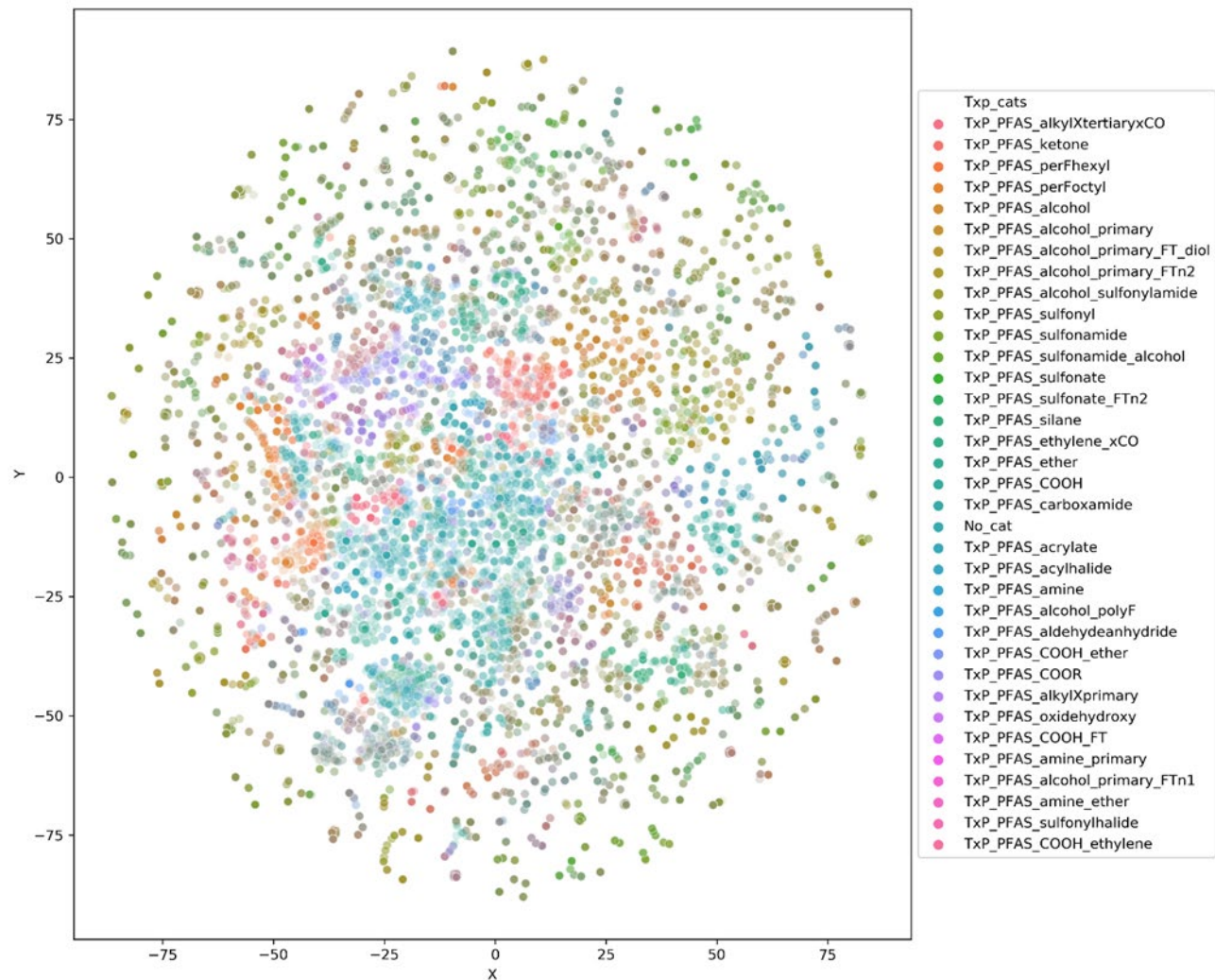
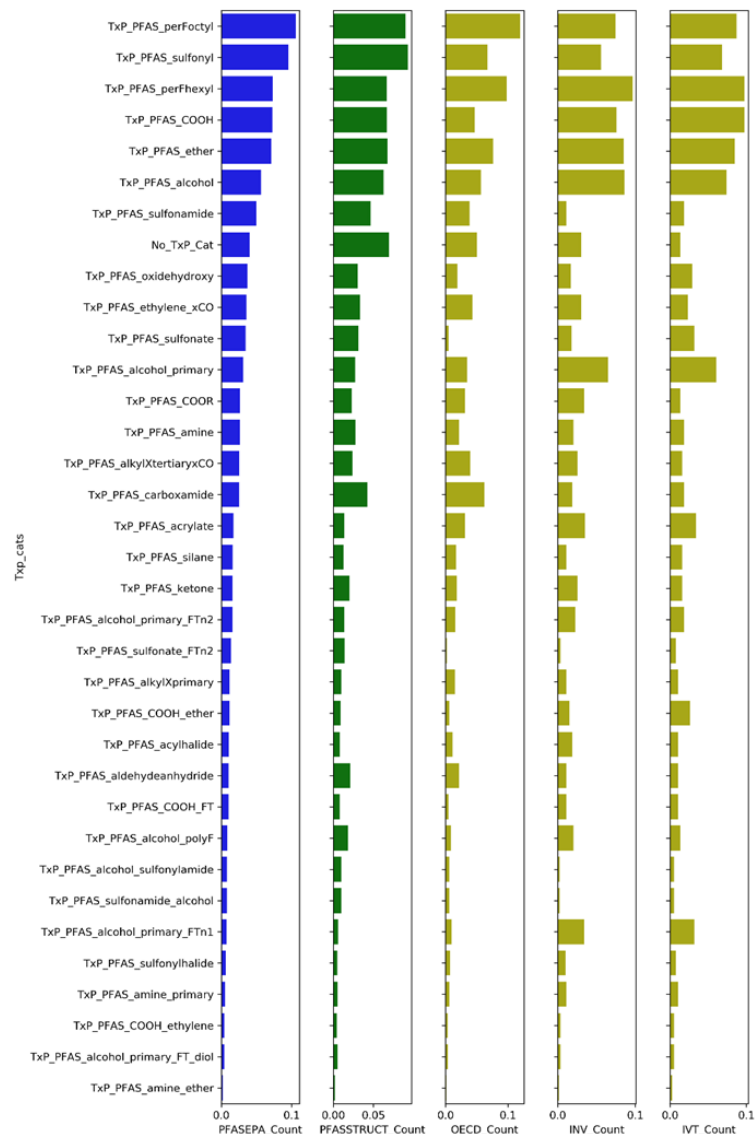
→ Clear, reproducible means for defining regions of local chemistry, i.e. categories!!

PFAS Structure-based Categorisation

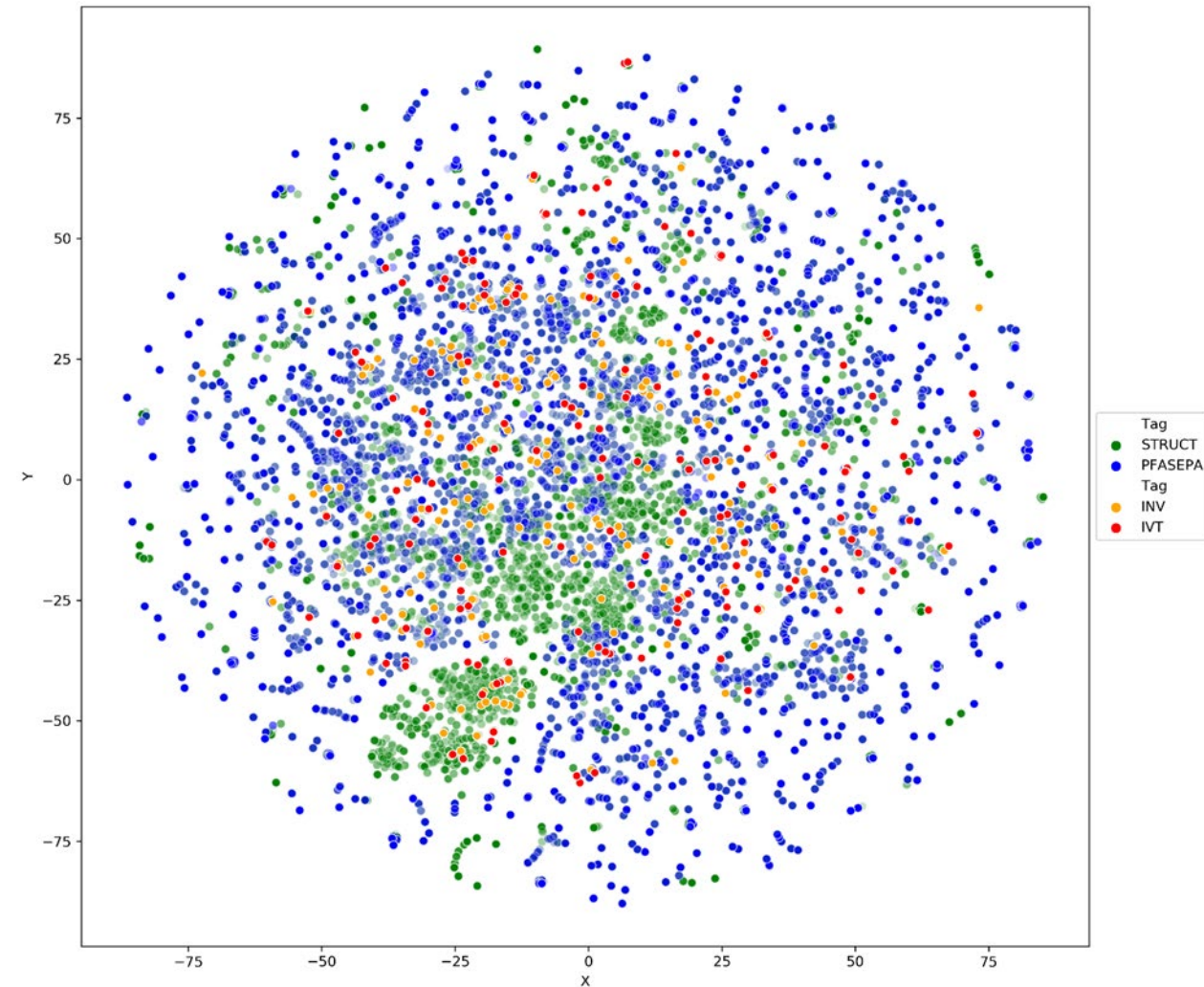
- Reconcile the different structural categories schemes initially used - by creating a harmonised set of structure-based categories
- Category assignments should be computationally generated from structure only → reproducible, transferable, standardised, extendable
- Permits nested & overlapping categories such that categories can be tailored to different datasets and decision contexts
- ToxPrints were used to develop 34 structural categories (TxP Categories) which cover >90% of the different PFAS inventories

PFAS Structure-based Categorisation

Comparison of different inventories (PFASSTRUCT, OECD & the PFAS430INV) using the TxP Categories

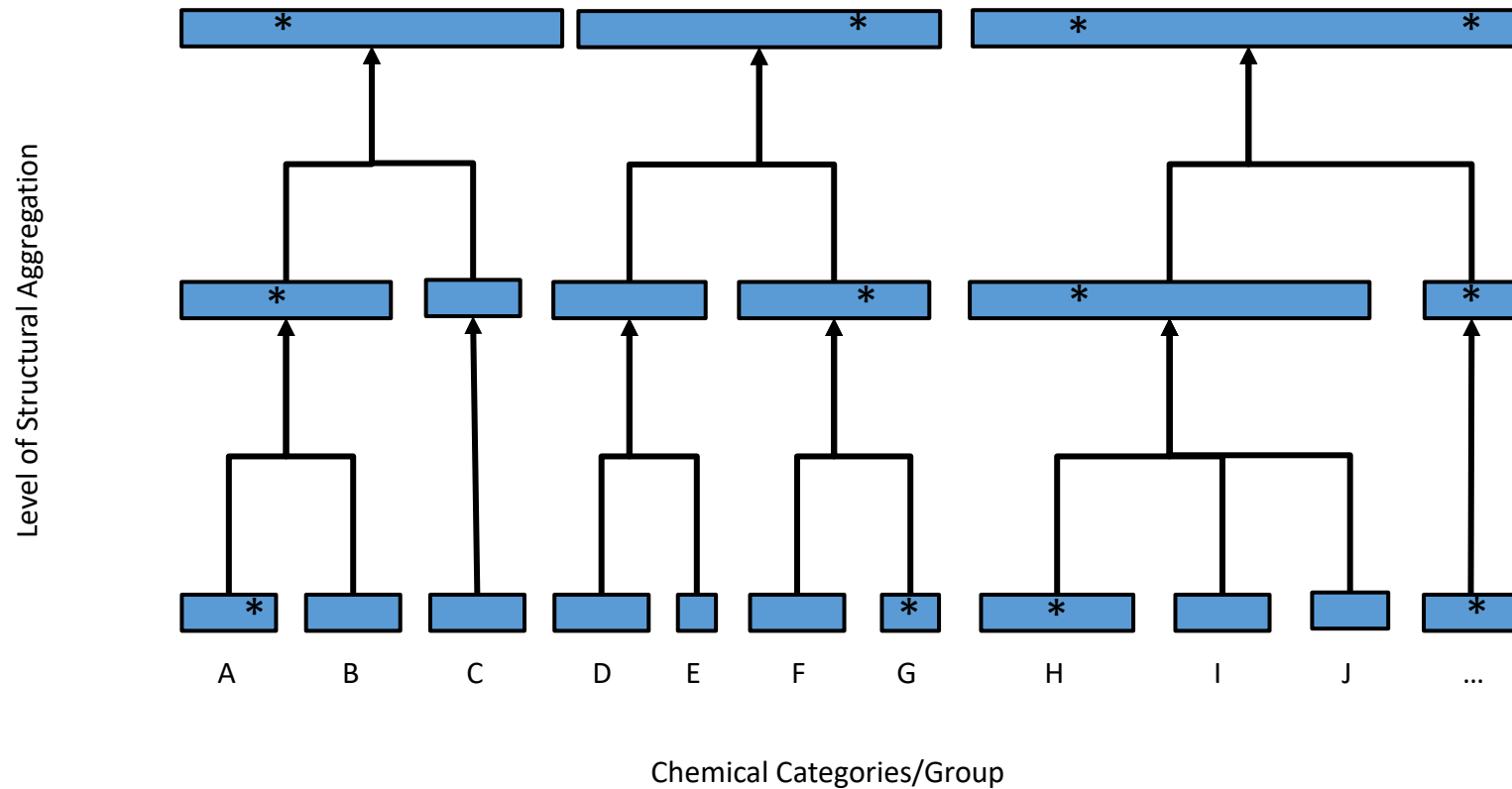


PFAS Coverage based on structure



- A 2D representation constructed using t-Distributed Stochastic Neighbour Embedding (t-SNE) based on 729 ToxPrints as chemical fingerprints
- PFAS430 inventory well distributed across the PFASSTRUCT inventory

Current PFAS Structural Grouping Approaches Use Different Levels of Aggregation



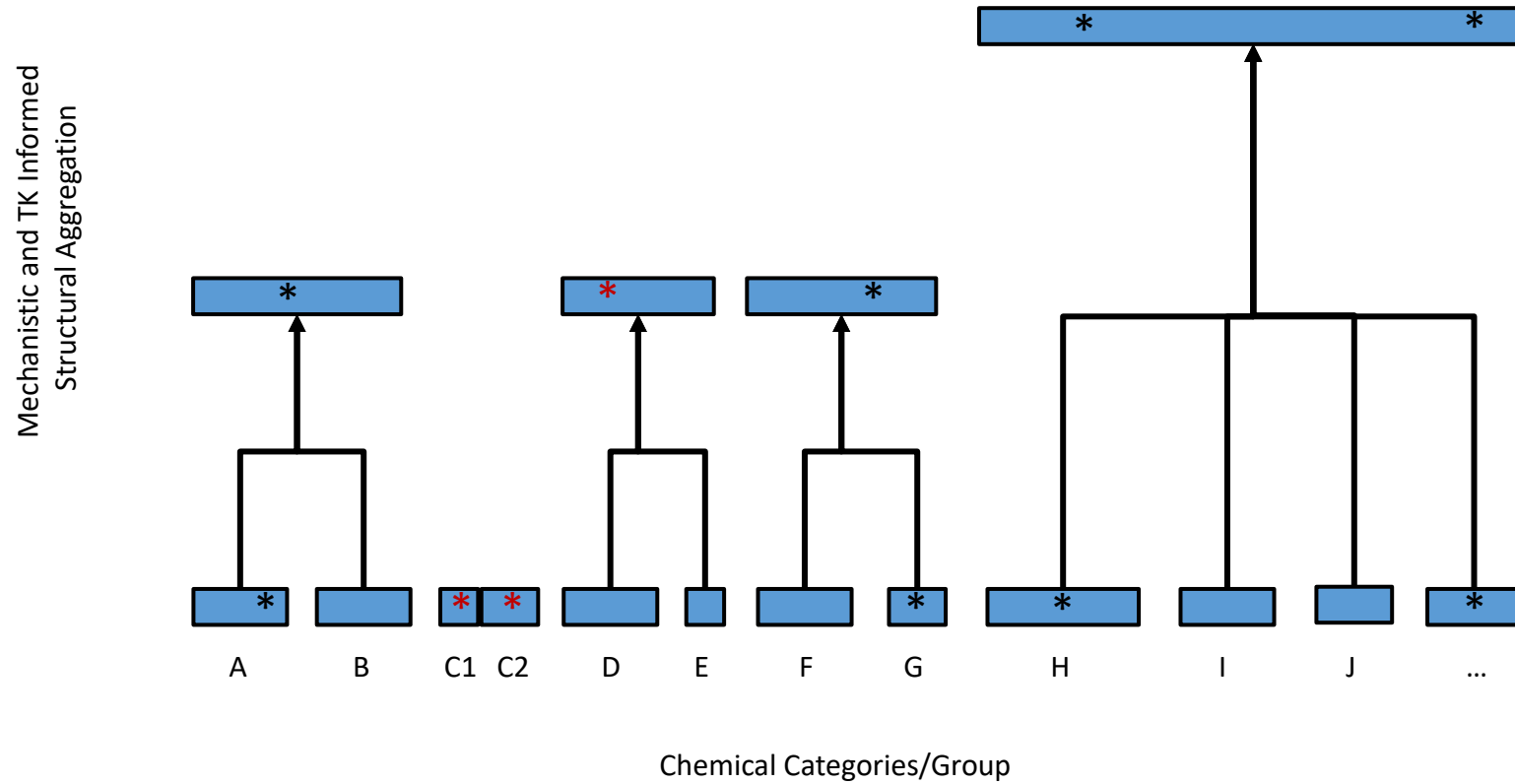
* Available source *in vivo* tox study

In Vitro Toxicity and Toxicokinetic Testing

Toxicological Response	Assay	Assay Endpoints	Purpose
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Plasma protein binding	Ultracentrifugation assay	Fraction of chemical not bound to plasma protein	Measure amount of free chemical in the blood

PFAS Category Aggregation that incorporates Structural, Mechanistic and Toxicokinetic Data



*Needed *in vivo* tox study

*Available source *in vivo* tox study

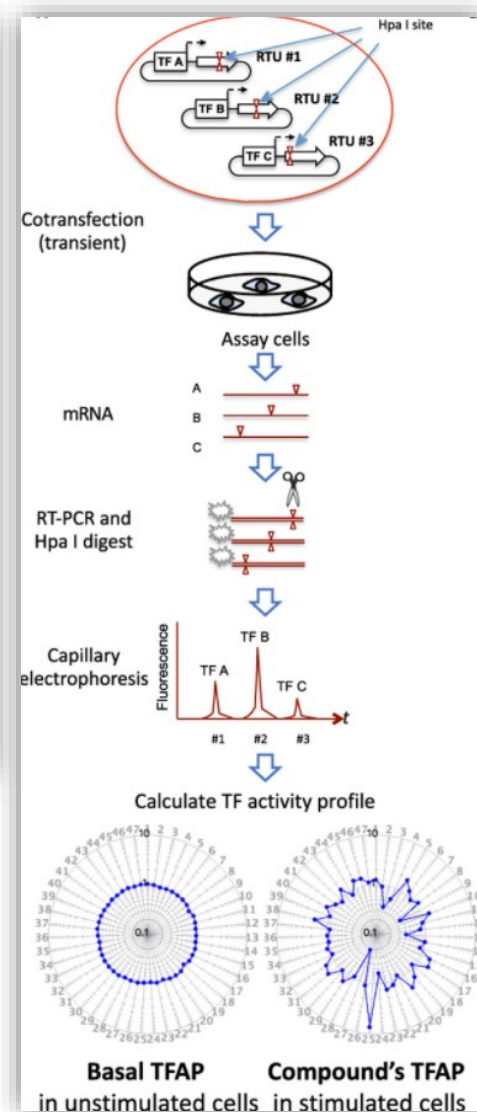
Targeted screening for nuclear receptor activation and cell stress



Gathering information on nuclear receptor and cell stress pathways via transcription factor activity profiling (TFAP)

>3800 ToxCast chemicals have been screened in concentration response in the Attagene transcription factor profiling system

- HepG2 HG19 subclone for elevated xenobiotic metabolic capacity
- "CIS" assays: endogenous transcription factors that regulated transfected reporters (nuclear receptor promoter elements, cell stress)
- "TRANS" assays: exogenous receptor-reporter system is transfected in (xenobiotic nuclear receptors)
- Used for environmental mixtures and single chemical screening



Number	Endpoint	Go Process	Number	Endpoint	Go Process
1	GAL4_TRANS	control	41	AP_1_CIS	response to stress
2	M_06_CIS		42	HIF1a_CIS	
3	M_06_TRANS		43	HSE_CIS	
4	M_19_CIS		44	MRE_CIS	
5	M_19_TRANS		45	NRF1_CIS	
6	M_32_CIS		46	NRF2_ARE_CIS	biosynthetic process
7	M_32_TRANS		47	Oat_MLP_CIS	
8	M_61_CIS		48	p53_CIS	
9	M_61_TRANS		49	Xbp1_CIS	
10	TA_CIS		50	CRE_CIS	
11	TAL_CIS		51	ERRa_TRANS	cell differentiation
12	CMV_CIS		52	ERRg_TRANS	
13	E_Box_CIS	cell proliferation	53	GR_TRANS	
14	E2F_CIS		54	GRE_CIS	
15	EGR_CIS		55	DR5_CIS	
16	Ets_CIS	reproduction	56	RARa_TRANS	anatomical structure development
17	Pax6_CIS		57	RARb_TRANS	
18	AR_TRANS		58	RARg_TRANS	
19	ERa_TRANS		59	RXRa_TRANS	
20	ERE_CIS		60	RXRb_TRANS	lipid metabolic process
21	THRa1_TRANS	immune system process	61	NURR1_TRANS	
22	VDR_TRANS		62	RORb_TRANS	
23	VDRE_CIS		63	RORg_TRANS	
24	ISRE_CIS		64	RORE_CIS	
25	NF_kB_CIS	lipid metabolic process	65	Sox_CIS	xenobiotic metabolic process
26	IR1_CIS		66	AP_2_CIS	
27	FXR_TRANS		67	BRE_CIS	
28	DR4_LXR_CIS		68	C_EBP_CIS	
29	LXRa_TRANS		69	FoxA2_CIS	xenobiotic metabolic process
30	LXRb_TRANS	xenobiotic metabolic process	70	FoxO_CIS	
31	PPARa_TRANS		71	GATA_CIS	
32	PPARd_TRANS		72	GLI_CIS	
33	PPARg_TRANS		73	HNF4a_TRANS	
34	PPRE_CIS	xenobiotic metabolic process	74	HNF6_CIS	xenobiotic metabolic process
35	SREBP_CIS		75	Myb_CIS	
36	Ahr_CIS		76	Myc_CIS	
37	CAR_TRANS		77	NFI_CIS	
38	PBRFEM_CIS		78	Sp1_CIS	
39	PXR_TRANS	xenobiotic metabolic process	79	STAT3_CIS	xenobiotic metabolic process
40	PXRE_CIS		80	TCF_b_cat_CIS	
			81	TGFb_CIS	

There are differences in assay sensitivity by mode and receptor, based on expression and design differences.

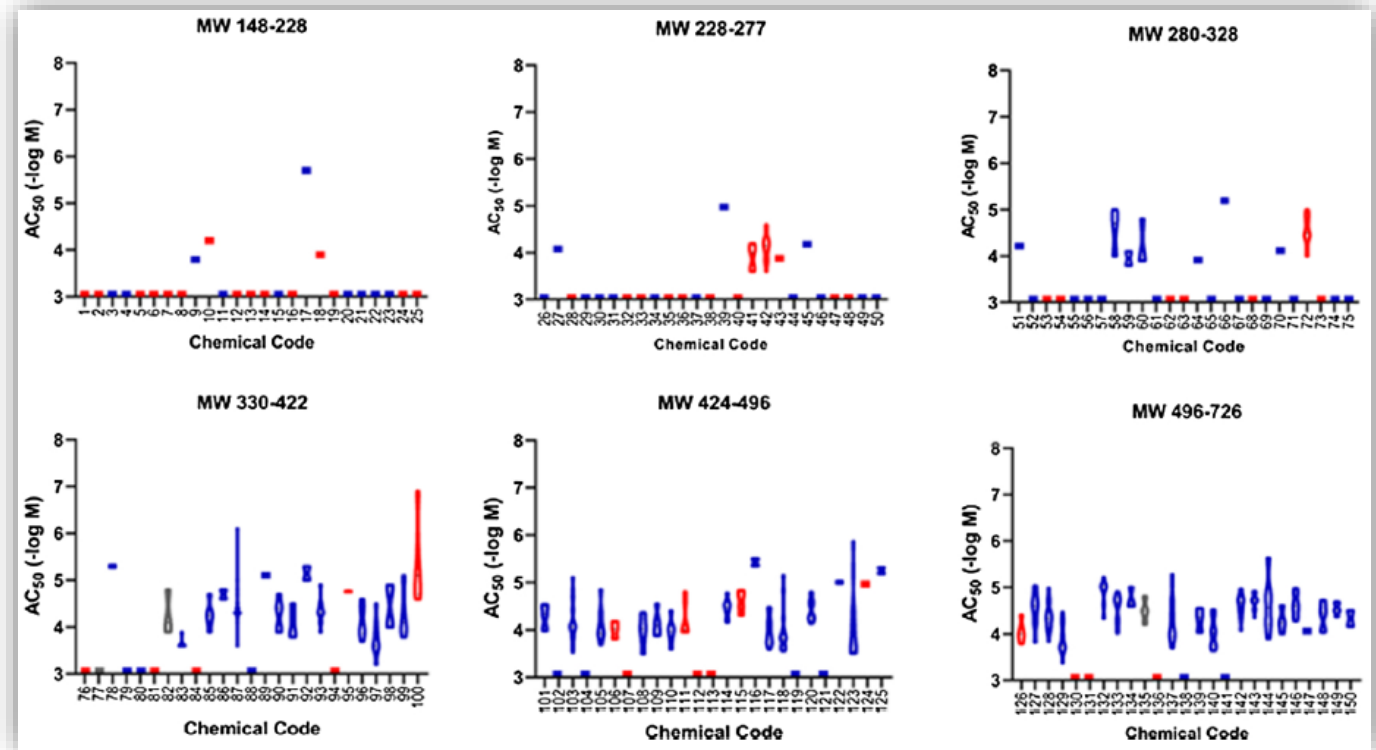
Table 1
Nuclear receptors included in FACTORIAL-TRANS assay.

#	Abbreviation	Receptor Name	Nomenclature	Reference Agonist (Fold-Increase)	cis-Factorial Assay (Fold-Increase)	Receptor Expression in HepG2 ¹
1	FXR	Farnesoid X receptor	NR1H4	Lithocholic acid (3.5)	IR1 (1.9)	Moderate
2	AR	Androgen receptor	NR3C4	Testosterone propionate (44.1)	NA	Very low
3	RAR γ	Retinoic acid receptor- γ	NR1B3	All-trans retinoic acid (3.9)	DR5 (20.2)	Moderate (RAR subfamily) ²
4	GAL4	Yeast GAL4, negative control	GAL4	NA	NA	NA
5	RXR α	Retinoid X receptor- α	NR2B1	Bexarotene (18.5)	DR5 (8.3)	Moderate (RXR subfamily) ²
6	GR	Glucocorticoid receptor	NR3C1	Betamethasone (29.1)	GRE (4.6)	Moderate
7	RAR β	Retinoic acid receptor- β	NR1B2	All-trans retinoic acid (1.6)	DR5 (20.2)	Moderate (RAR subfamily) ²
8	RAR α	Retinoic acid receptor- α	NR1B1	All-trans retinoic acid (5.5)	DR5 (20.2)	Moderate (RAR subfamily) ²
9	PPAR γ	Peroxisome proliferator-activated receptor- γ	NR1C2	Rosiglitazone maleate (44.8)	PPRE (3.8)	High
10	ERR γ	Estrogen-related receptor- γ	NR3B3	4-Nonylphenol, branched (2.7)	NA	NA
11	ROR β	RAR-related orphan receptor- β	NR1F1	SSR69071 (7.8)	RORE (5.9)	NA
12	ER α	Estrogen receptor- α	NR3A1	17 β -Estradiol (22.6)	ERE (19.1)	Very low; full-length human ER α co-expressed in FACTORIAL-CIS
13	LXR α	Liver X receptor- α	NR1H3	Lynestrenol (13.9)	DR4 (2.3)	High (LXR subfamily) ²
14	ERR α	Estrogen-related receptor- α	NR3B1	4-Nonylphenol, branched (2.7)	NA	NA
15	PXR	Pregnane X receptor	NR1I2	Rifampicin (3.8)	PXRE (9.1)	Moderate; full-length human PXR co-expressed in FACTORIAL-CIS
16	TR α	Thyroid hormone receptor- α	NR1A1	3,5,3'-Triiodothyronine (33.0)	NA	High
17	LXR β	Liver X receptor- β	NR1H2	Lynestrenol (8.7)	DR4 (2.3)	High (LXR subfamily) ²
18	CAR	Constitutive androstane receptor	NR1I3	p,p'-DDT (3.5)	PBREM (1.0)	Very low
19	PPAR α	Peroxisome proliferator-activated receptor- α	NR1C1	Pirixinic acid (14.1)	PPRE (2.4)	Moderate
20	ROR γ	RAR-related orphan receptor- γ	NR1F3	SSR69071 (14.2)	RORE (5.9)	NA
21	RXR β	Retinoid X receptor- β	NR2B2	Bexarotene (15.2)	DR5 (8.3)	Moderate (RXR subfamily) ²
22	HNF4 α	Hepatocyte nuclear factor-4 α	NR2A1	NA	NA	High
23	NURR1	Nuclear receptor related 1	NR4A2	Bexarotene (24.6)	NA	NA
24	VDR	Vitamin D receptor	NR1I1	Ergocalciferol (32.6)	VDRE (1.2)	Very low
25	PPAR δ	Peroxisome proliferator-activated receptor- δ	NR1C3	12-Hydroxyoctadecanoic acid (9.3)	PPRE (2.9)	NA

- Low- to negligible-expression in HepG2 cells of ER α and PXR was overcome by cotransfection of full-length receptors in the TRANS assay
- CAR and VDR have very low sensitivity to ligands due to reliance only on endogenous receptor expression in the host cell.

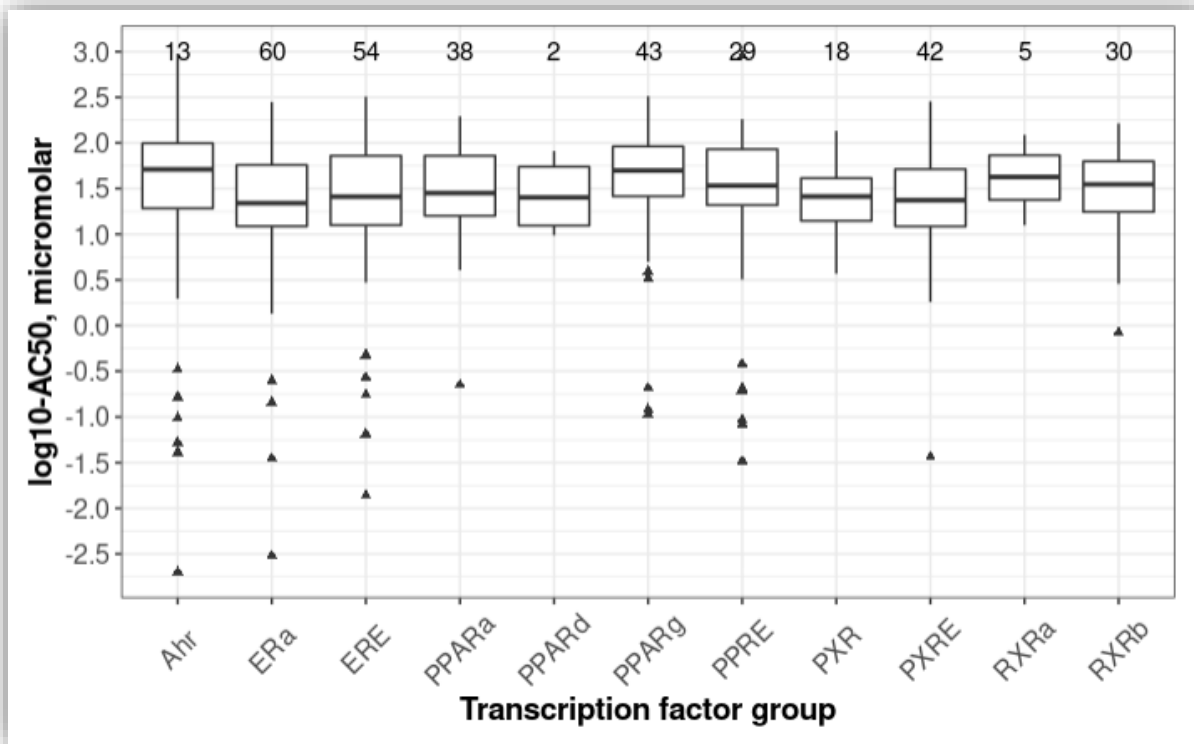
As with other assay platforms screened, lower MW often corresponded to more limited bioactivity, but there may be more than one reason.

- PFAS with molecular weight less than 330 g/mol appeared less likely to be active in the Attagene assays and more likely to “fail” analytical QC (defined as parent structure not detected).
- Activity was not detected for 76 distinct structures, of which 55 % failed analytical QC.
- 67% of the “failed” samples had predicted vapor pressures in excess of 100 mmHg, suggesting that chemical volatilisation may have played a role in limited bioactivity of some of these samples.
- The specific acid form of PFAS may also be important, as the free acid form of the chemical known as “GenX” (perfluoro-2-methyl-3-oxahexanoic acid (DTXSID70880215) did not have a high vapor pressure (was unlikely to have volatilised), but the ammonium salt form of this chemical (DTXSID40108559) showed activity as a PPAR α agonist when solubilised in water (rather than DMSO).



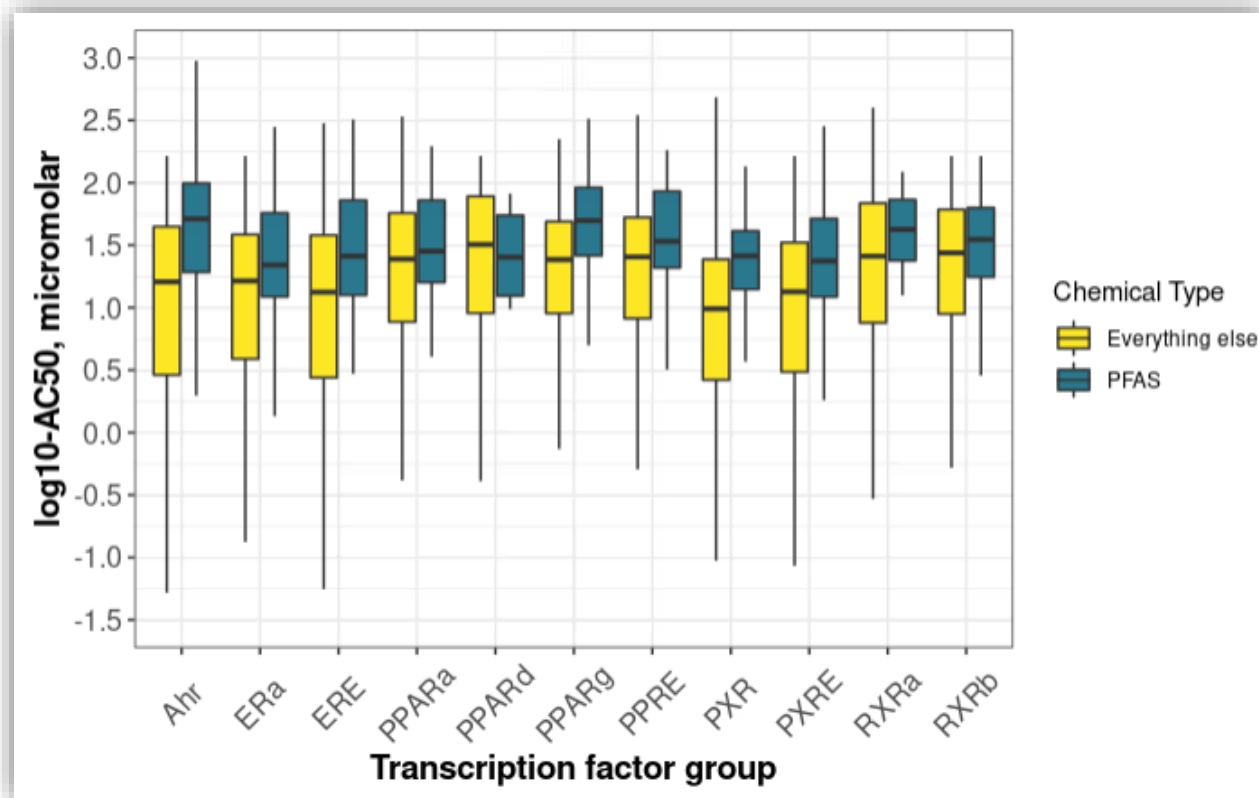
Houck et al. 2020, Fig1B.

Potency for the PFAS that were positive at key transcription factor targets tended to be somewhat left-shifted from the rest of the ToxCast library

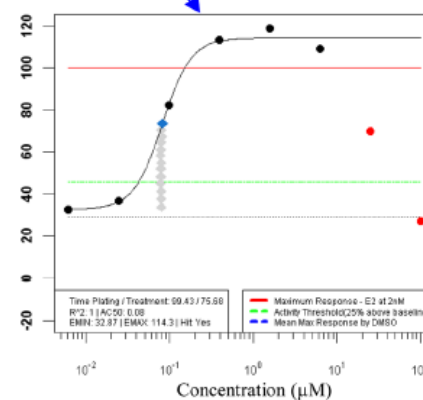
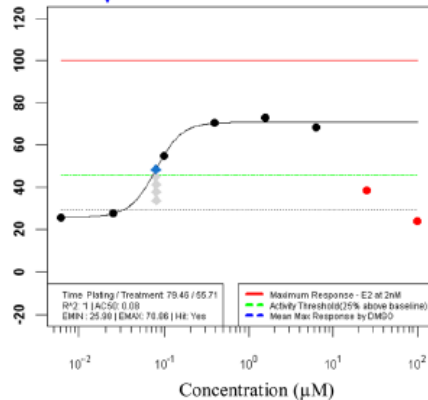
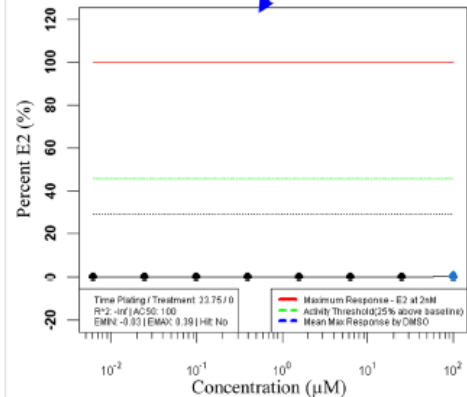
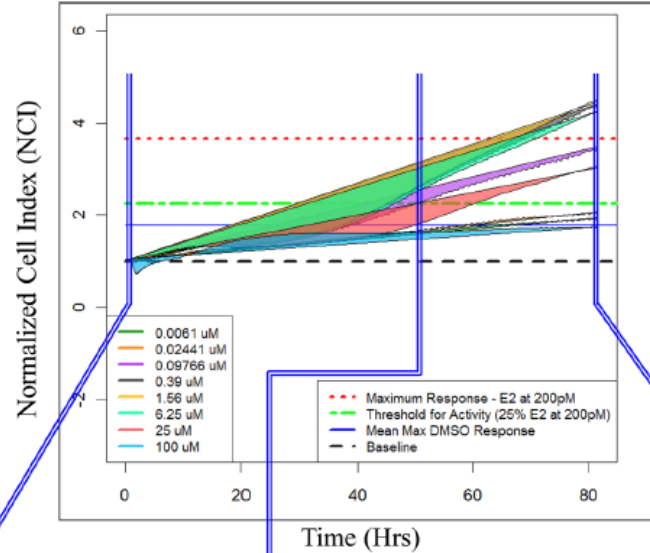


- Many PFAS were negative in the transcription factor activity screening
- Aryl hydrocarbon receptor (AhR), estrogen receptor alpha (ERa), PPAR alpha, delta, and gamma (PPARa,d,g), the pregnane X receptor (PXR), and RXR alpha and beta (RXRa,b) emerged as targets.

- The number of chemicals that simply hit one or more relevant assays for a particular transcription factor group can be examined in more depth for confirmation.



Estrogen receptor activity can be confirmed with orthogonal assays including ACEA: Real Time Cell Analysis Based on Electrical Impedance



Chemical Research in Toxicology

Article

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Real-Time Growth Kinetics Measuring Hormone Mimicry for ToxCast Chemicals in T-47D Human Ductal Carcinoma Cells

Daniel M. Rotroff,^{†,‡} David J. Dix,[‡] Keith A. Houck,[‡] Robert J. Kavlock,[‡] Thomas B. Knudsen,[‡] Matthew T. Martin,[‡] David M. Reif,[‡] Ann M. Richard,[‡] Nisha S. Sipes,[‡] Yama A. Abassi,[§] Can Jin,[§] Melinda Stampfl,[§] and Richard S. Judson^{*,‡}

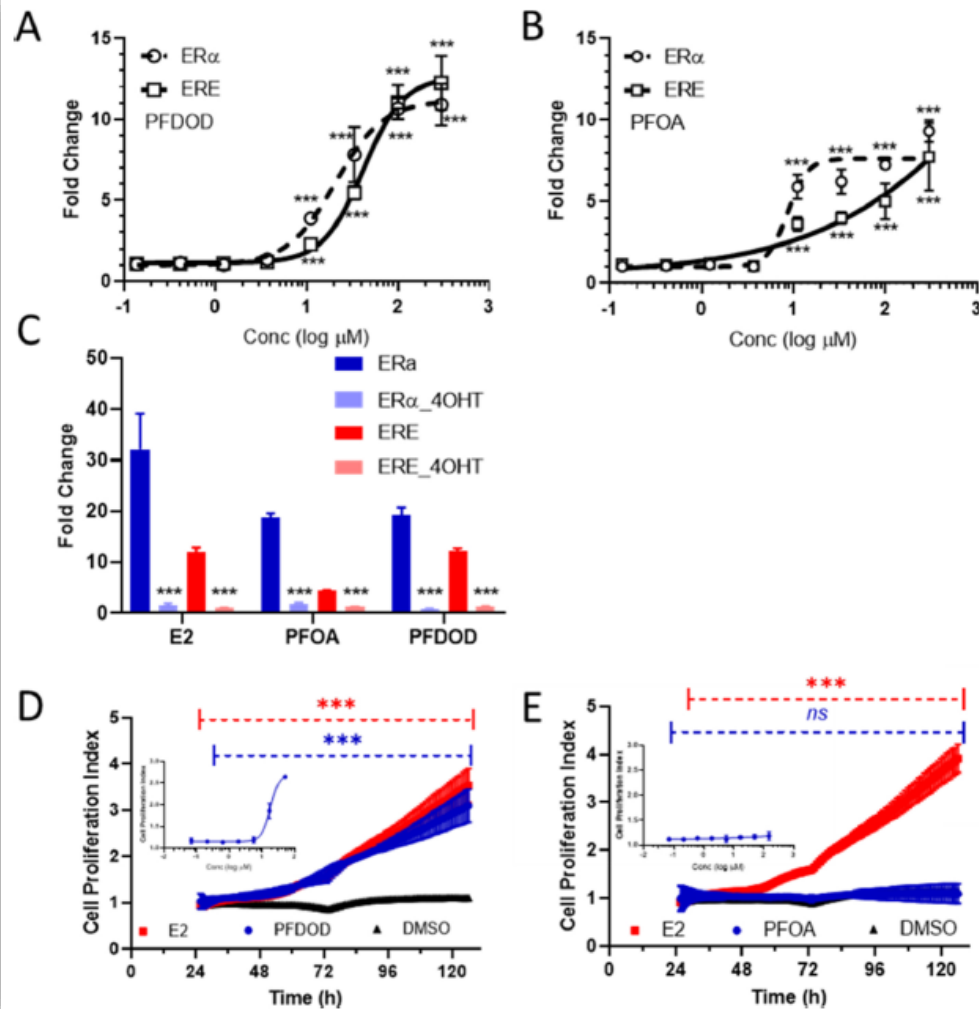
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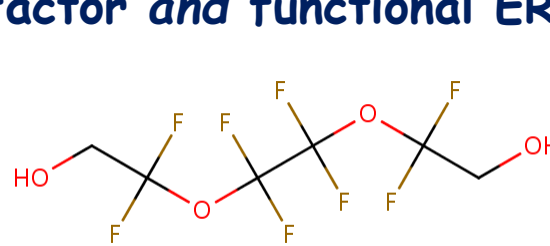
- Can measure cell proliferation or cytotoxicity depending on the direction
- Electrical impedance measured over 80 hr
- ACEA ER assay uses T-47D breast cancer cells

Confirmation of transcriptional responses with functional activity is an important strategy for ER bioactivity

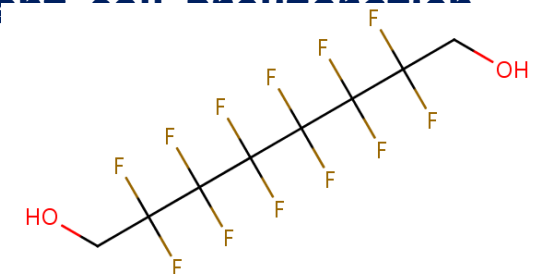


- 40-60 PFAS demonstrated some activity in the ATG $ER\alpha$ TRANS or ERE CIS assays; viewing these assays as orthogonal reduces the set to <10.
 - All of these were less potent than 17β -estradiol.
 - Acrylates and N-alkyl perfluoroalkyl (linear) sulfonamide structural categories were significantly associated with ER activity.

- Adding in ACEA as another orthogonal assay to confirm specificity leads indicates few PFAS with transcription factor and functional ER-dependent cell proliferation



1H,1H,8H,8H-Perfluoro-3,6-dioxaoctane-1,8-diol



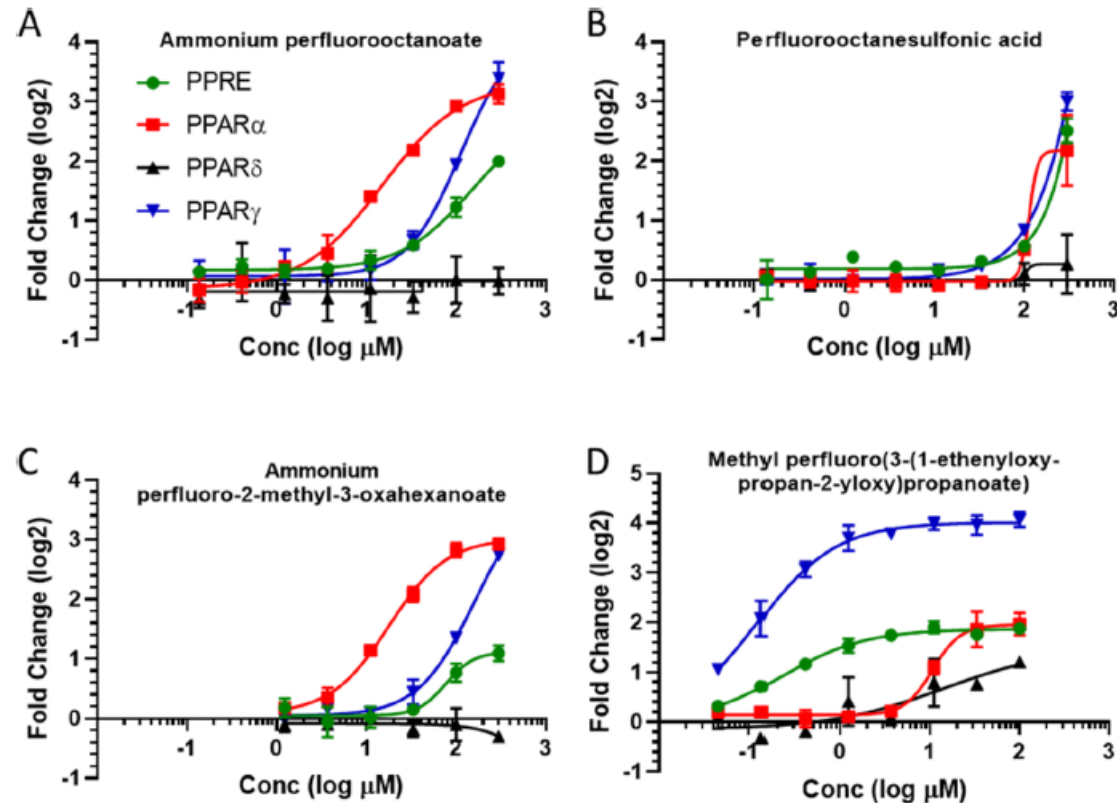
1H,1H,8H,8H-Perfluorooctane-1,8-diol

- PFOA activated ATG $ER\alpha$ TRANS and ERE CIS but failed to produce functional ER-dependent cell proliferation in ACEA.

As expected PPAR activity was observed for a subset of PFAS.

- The TRANS assay contained endpoints for all three human PPARs (α, δ, γ) whereas the CIS assay contained a reporter gene controlled by a PPAR-response element that responds to all three PPARs endogenously expressed in the HepG2 host cells.
- Functional groups enriched within the actives were mostly carboxylates along with sulfonates, sulfonamides and a thenoylketone, which all have a negative ionic charge at physiological pH, consistent with known critical components for ligand-binding.
- Not much activity at PPAR δ (smaller binding pocket?).

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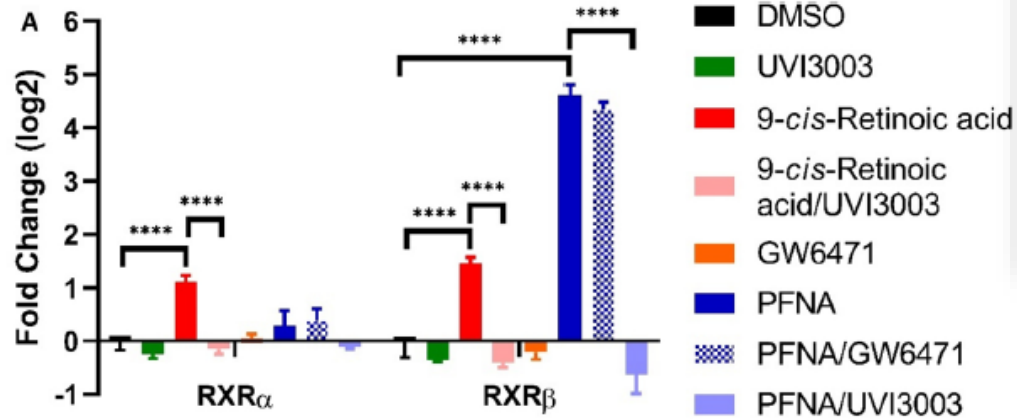
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Fig. 6. Transactivation of the peroxisome proliferator-activated receptors (PPARs) by example PFASs. Concentration-response data for PPAR- α , - δ , and - γ in the FACTORIAL-TRANS assays and the PPAR response element (PPRE) in the FACTORIAL-CIS assay following treatment for 20-24 h with increasing concentrations of ammonium perfluorooctanoate (A), perfluorooctanesulfonic acid (B), ammonium perfluoro-2-methyl-3-oxahexanoate (C), and methyl perfluoro(3-(1-ethenyloxypropan-2-yloxy)propanoate) (D). Values are the mean reporter gene activity expressed as fold-change (log₂) normalized by solvent control (dimethyl sulfoxide) values.

Houck et al. 2020, Fig6.

~17 PFAS activated RXR β , with two of these active at RXR α

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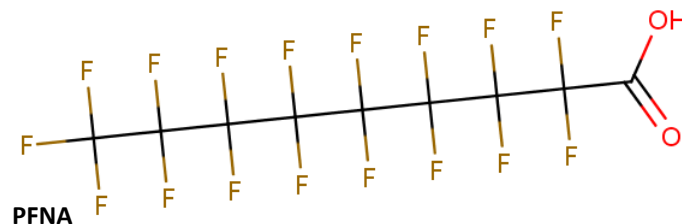


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Fig. 8. PFAS activity for retinoid X receptors (RXR). A) Responses of RXR α and RXR β to perfluorononanoic acid (PFNA) and effects of pharmacological agents UVI3003 (5 μ M), a pan-RXR antagonist; 9-cis retinoic acid (0.02 μ M), a pan-RXR agonist; and GW6471 (5 μ M), a PPAR α -selective antagonist; in the presence and absence of PFNA (66 μ M). No significant activation of RXR α by PFNA was observed. Significance was established with an ordinary one-way ANOVA and Tukey's multiple comparisons test. (**** = $P < .0001$). B) Radioligand

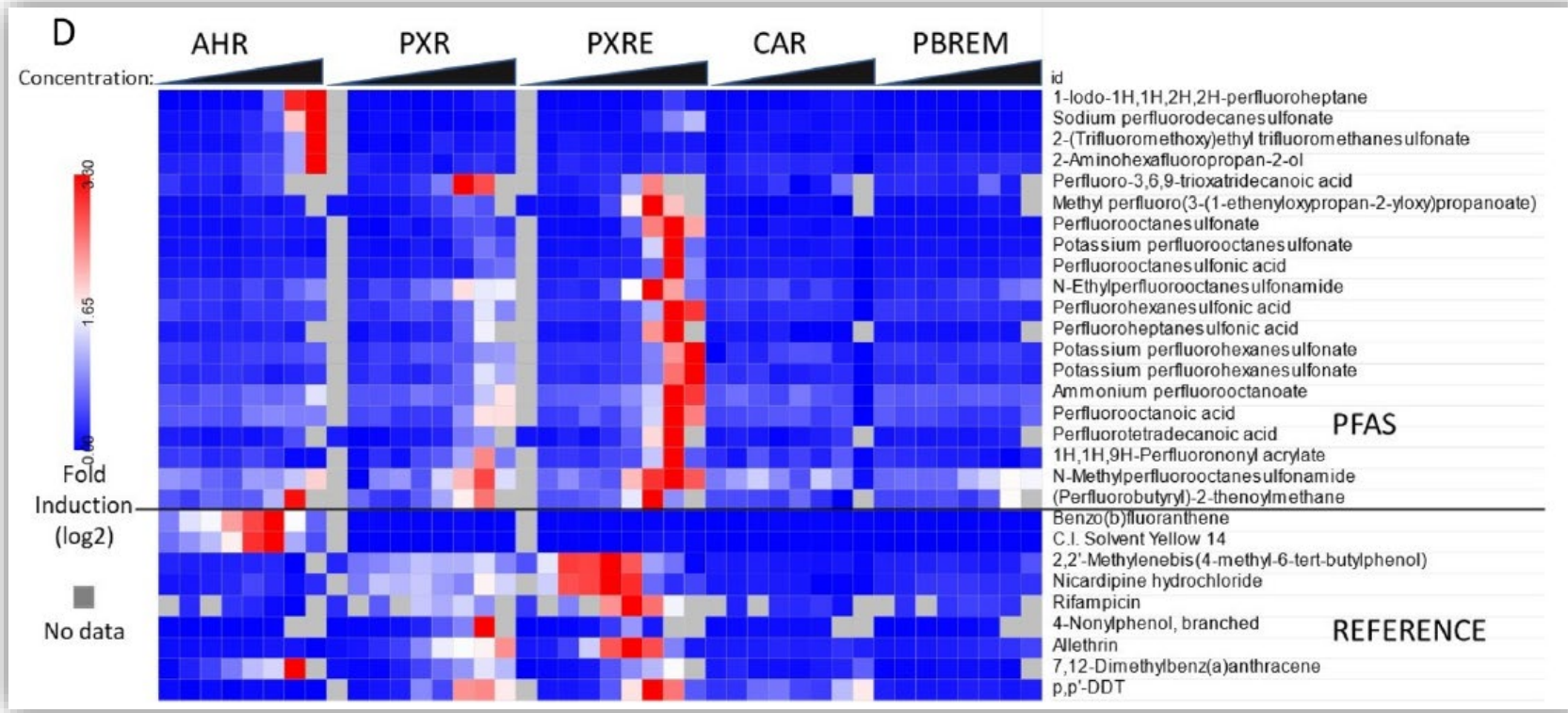
Houck et al. 2020, Fig8A.

PFNA appears to work through RXR specifically: an RXR-selective antagonist, UVI3003 (DTXSID501024375), completely blocked PFNA activation of RXR, whereas the PPAR α antagonist GW6471 was ineffective.



- Seventeen of the PFAS, mostly linear, fluorinated carboxylic acids, showed a novel finding of activation of RXR β .
- Most also activated PPAR α , PPAR γ and NRF2, with varying levels of selectivity. Only two activated RXR α ; however, NURR1 was activated, presumably through agonist effects on RXR β .
- All are structurally related perfluorinated carboxylic acids and meet defined ligand structural requirements for RXR.

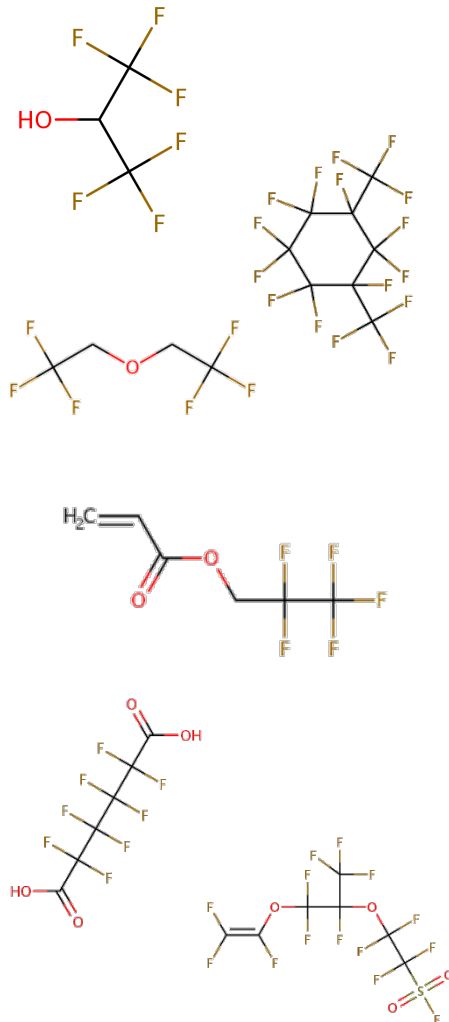
Xenobiotic nuclear receptor responses associated with hepatic metabolism may also be important targets to screen for PFAS bioactivity.



Houck et al. 2020, Fig3B.

- Many of the PFAS modulated the xenobiotic response, particularly PXR.
- Responses were generally modest with respect to potency and efficacy relative to prototypical PXR inducers.
- None of the PFAS were determined to be CAR activators, recognizing limitations in the FACTORIAL-CIS assay for CAR, likely due to negligible expression of CAR in HepG2 cells.
- Several PFAS structures activated the AhR, somewhat surprising in that all were linear fluoroalkyl molecules while the prototypical activator is a polycyclic aromatic hydrocarbon. Except for sodium perfluorodecanesulfonate and 1-Iodo-1H,1H,2H,2H-perfluoroheptane, the responses were very weak with unknown *in vivo* relevance.

Take Home Messages...



- Chemical curation efforts are important to harmonise structure, naming, and identifiers across the PFAS space
- A chemical library of 430 PFAS was assembled for chemical screening, analytical method development, and other research needs
- A subset of 150 PFAS selected for *in vitro* toxicity and toxicokinetic testing to refine/support read across categories
- *In vitro* toxicity and toxicokinetic testing and the ongoing analysis demonstrate the diverse biological activities and toxicokinetic properties of PFAS
- More information at <https://www.epa.gov/chemical-research/pfas-chemical-lists-and-tiered-testing-methods-descriptions>

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