

High-Throughput Transcriptomics (HTTr) Screening as a Component of NAMs-based Tiered Hazard Evaluation

Joshua A. Harrill

USEPA Center for Computational Toxicology and Exposure (CCTE)



Disclaimer

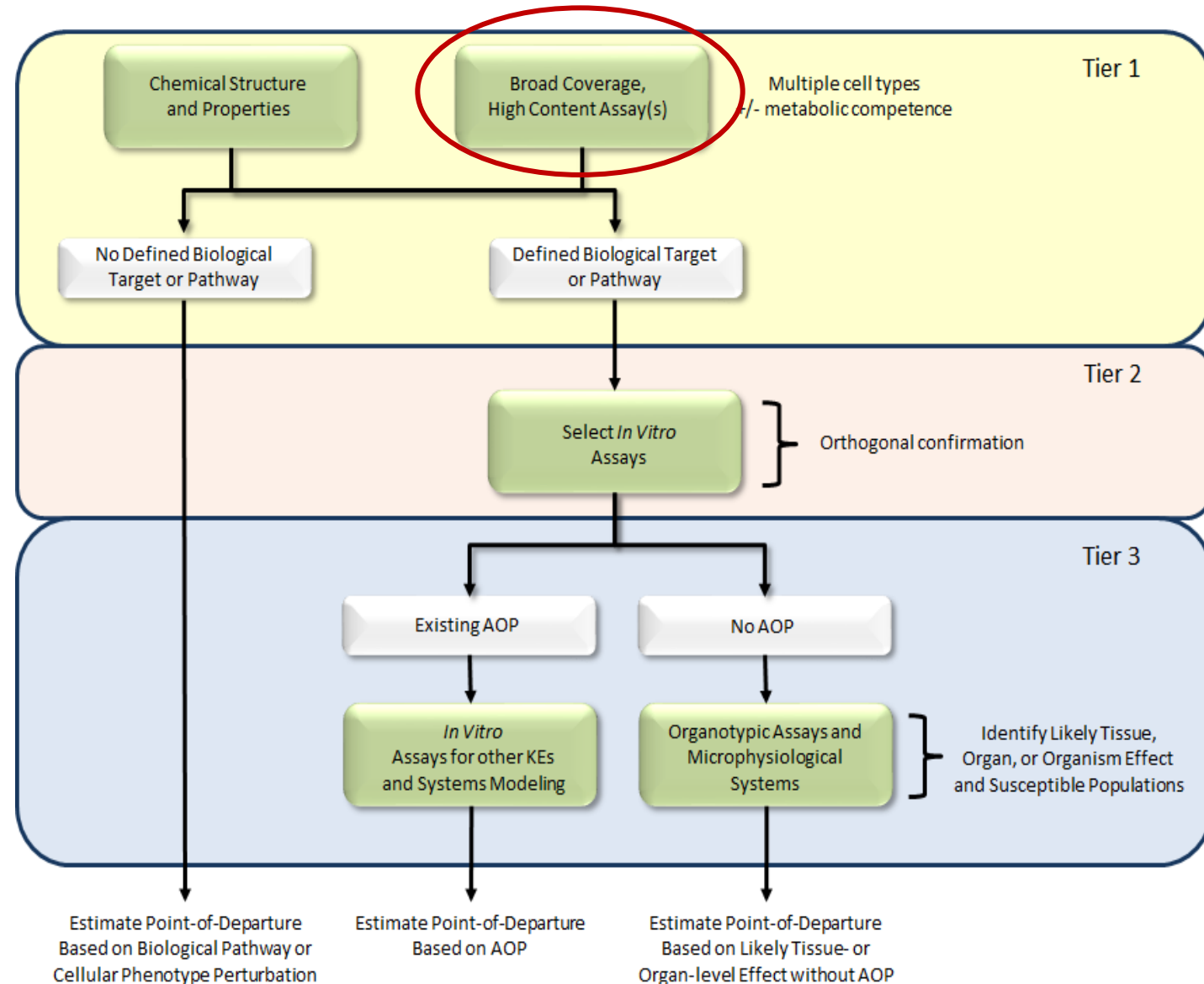
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Outline

- **NAMs-Based Tiered Hazard Evaluation Approach**
- **High-Throughput Transcriptomics (HTTr)**
 - Assay Concept
 - Data Analysis Pipeline
 - Concentration Response Modeling of Transcriptomic Signatures
- **Results**
 - Comparison to ToxCast
 - Comparison to ER Model
 - Bioactivity / *In Vivo* Effect Value Ratio Analysis
- **Current and Future Directions**

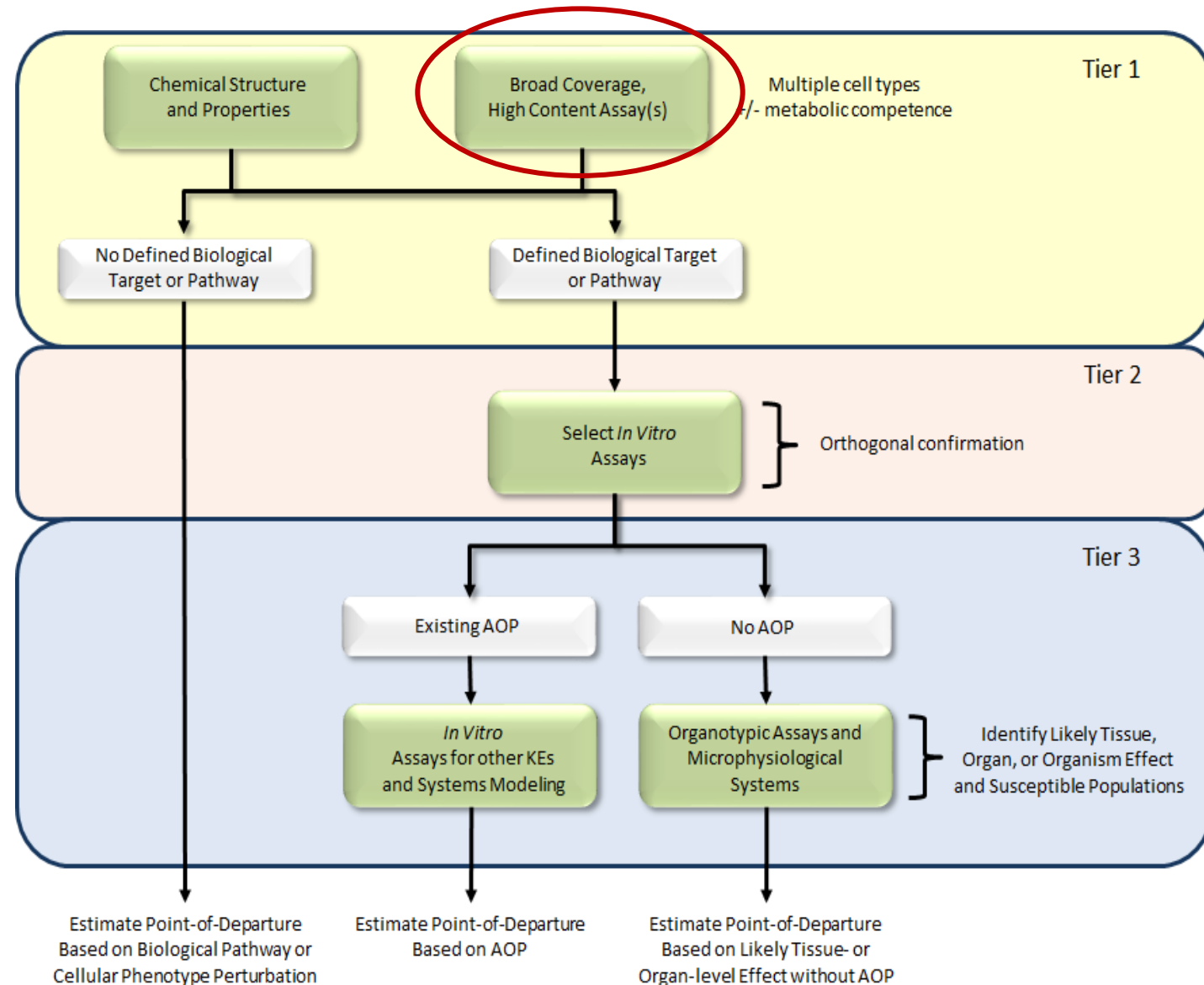
Tiered Hazard Evaluation Approach (1)

- New Approach Methodologies (NAMs)** are any technology, methodology, approach or combination thereof that can be used to provide information on chemical hazard and risk that avoids the use of intact animals.
- NAMs are a potential means to **reduce** the use of animals in toxicity testing and **accelerate** the pace of chemical risk assessment.
- US EPA CompTox Blueprint advocates the use of **high throughput profiling (HTP) assays** as the first tier in a NAMs-based hazard evaluation approach.
- HTP assay criteria:**
 - Yield bioactivity profiles that can be used for **potency estimation, mechanistic prediction** and evaluation of **chemical similarity**.
 - Compatible with multiple human-derived culture models.
 - Concentration-response screening mode.
 - Cost-effective.



Tiered Hazard Evaluation Approach (2)

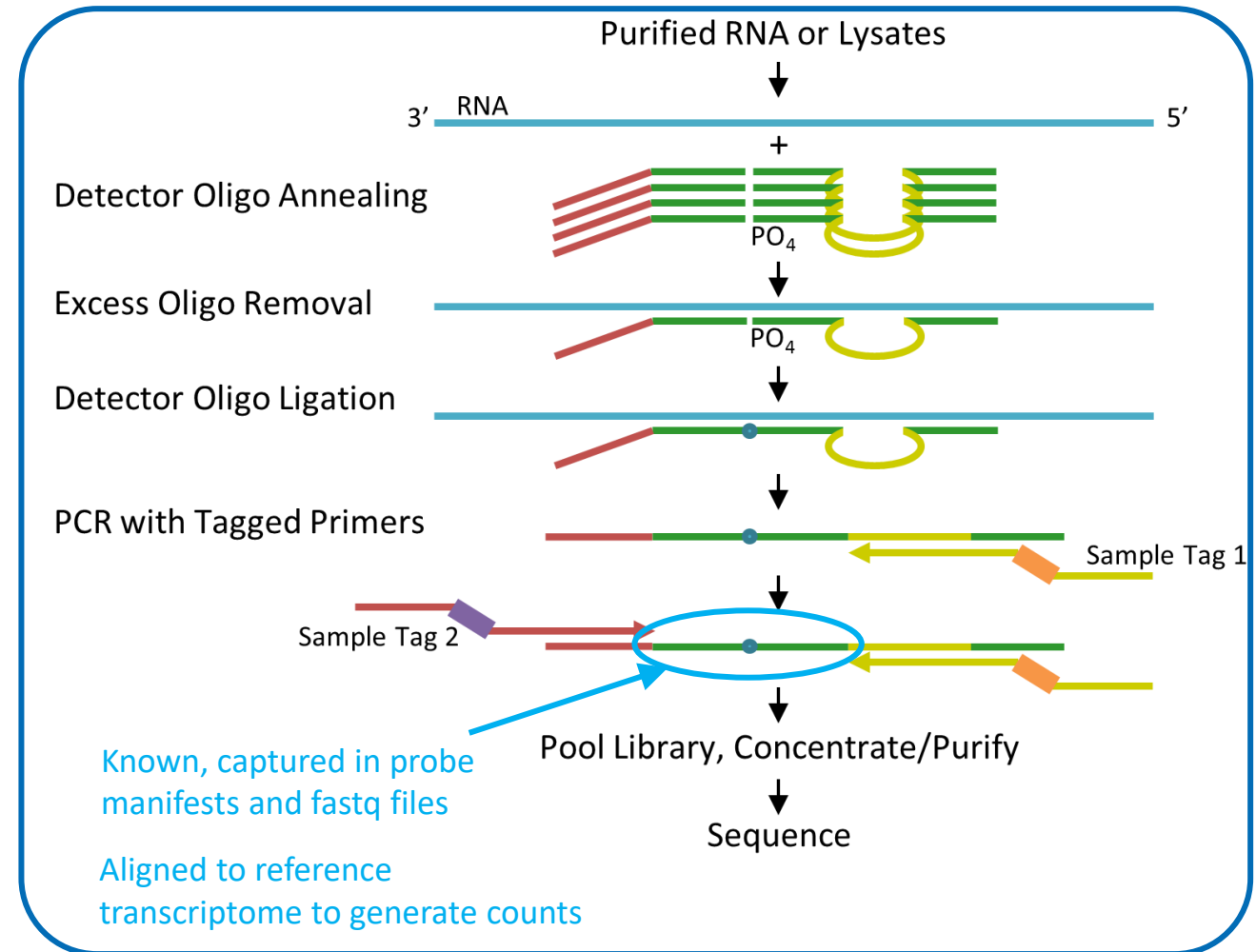
- To date, EPA has identified and implemented two HTP assays that meet this criteria.
- High-Throughput Transcriptomics [HTTr]**
 - Whole Transcriptome TempO-Seq
- High-Throughput Phenotypic Profiling [HTPP]**
 - Cell Painting
- Both methods are **complementary** to each other and can be used in many different human-derived cell types.
- EPA has established scalable laboratory and bioinformatics workflows for each assay.



Templated Oligo with Sequencing Readout (TempO-Seq)

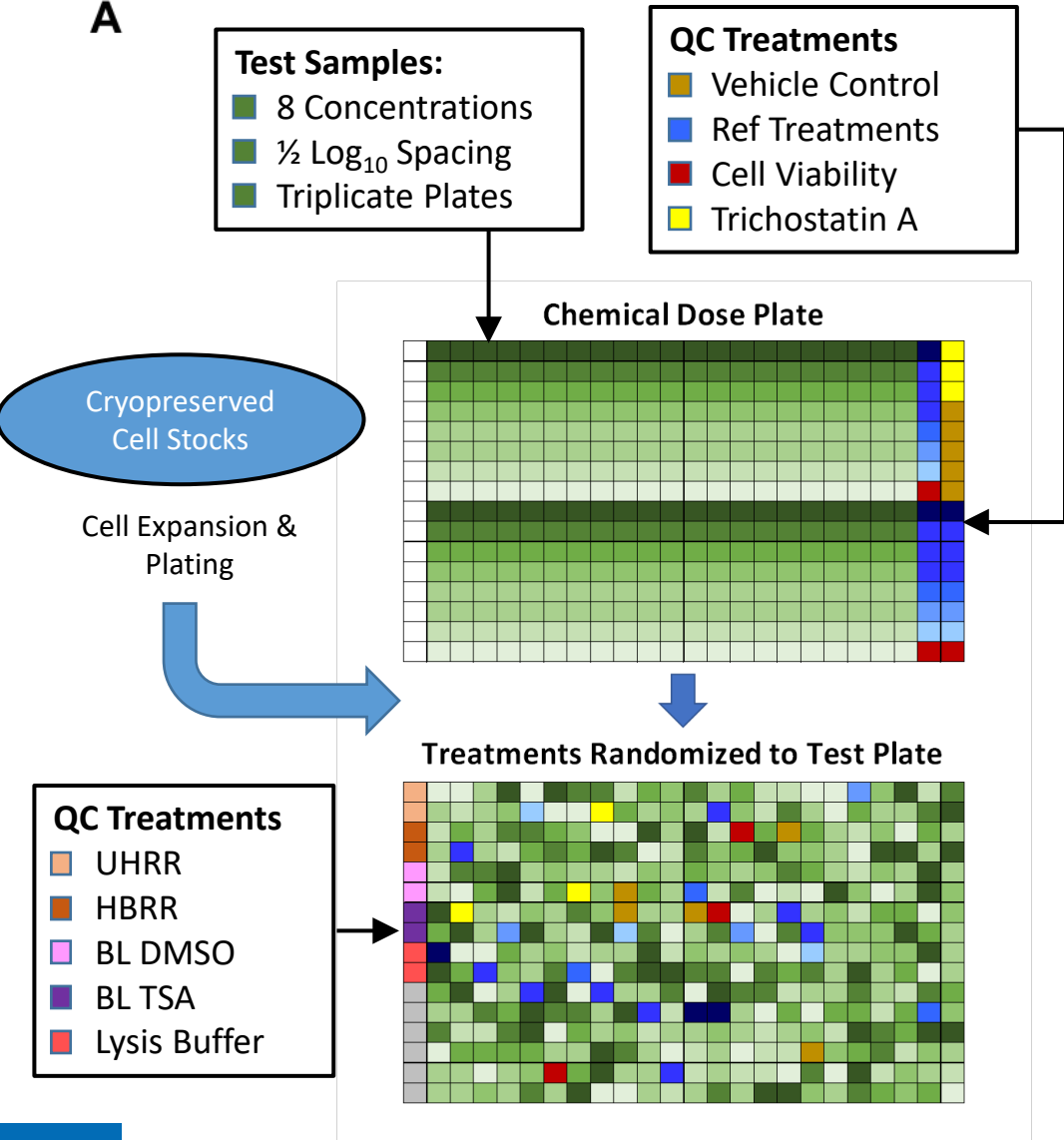
- The **TempO-Seq** human whole transcriptome assay measures the expression of greater than 20,000 transcripts.
- Requires only picogram amounts of total RNA per sample.
- Compatible with purified RNA samples or **cell lysates**.
- Lysates are barcoded according to sample identity and combined in a single library for sequencing using industry standard instrumentation.
- Scalable, targeted assay:
 - 1) specifically measures transcripts of interest
 - 2) ~50-bp reads for all genes
 - 3) requires less flow cell capacity than RNA-Seq

TempO-Seq Assay Illustration

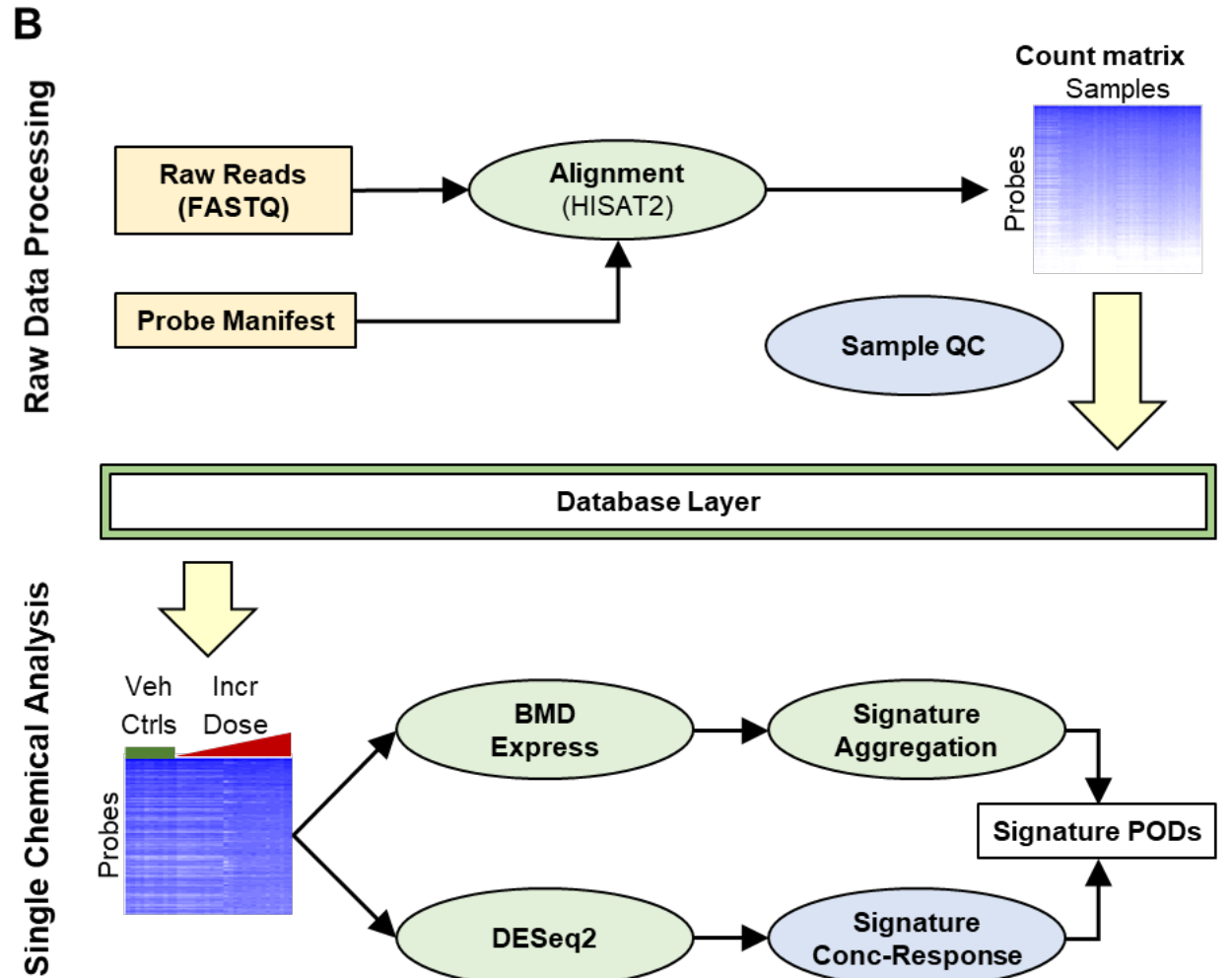


HTTr Experimental Design and Bioinformatics Workflow

A



B



Concentration-Response Modeling of Gene Signatures (1)

- Understanding the results of change in gene expression for **10,000 – 20,000 genes** is difficult.
- Grouping genes into gene signatures aids in **data interpretation**.
- **Examples of signature types:**
 - Genes that are perturbed in diseased tissue vs. healthy tissue
 - Genes perturbed in individuals with congenital diseases vs. those without
 - Genes perturbed by gene knockdowns / knockouts
 - Genes perturbed by drugs or other chemicals with known (or unknown) mechanisms
- **Example use:**
 - If an unknown *chemical X* perturbs genes that are also perturbed by a well-characterized chemical with a specific mechanism of action, then one can infer the *chemical X* may affect the same molecular target(s).
- **CCTE signature collection:**
 - Compiled from many public sources (MSigDB¹, BioPlanet², CMAP³, DisGeNET⁴) → ~10,000 signatures
 - Each signature has been manually-assigned a “super target” class to aid in interpretation → ~1000 super targets
 - Disease groups (Immune, Cancer, etc.)
 - Biological organization groups (molecule, pathway, cell, tissue, organ, etc.)

¹ Liberzon et al., *Bioinformatics*. 2011 Jun 15;27(12):1739-40

² Huang et al., *Front Pharmacol*. 2019 Apr 26;10:445

³ Subramanian et al., *Science*. 2006 Sep 29;313(5795):1929-35.

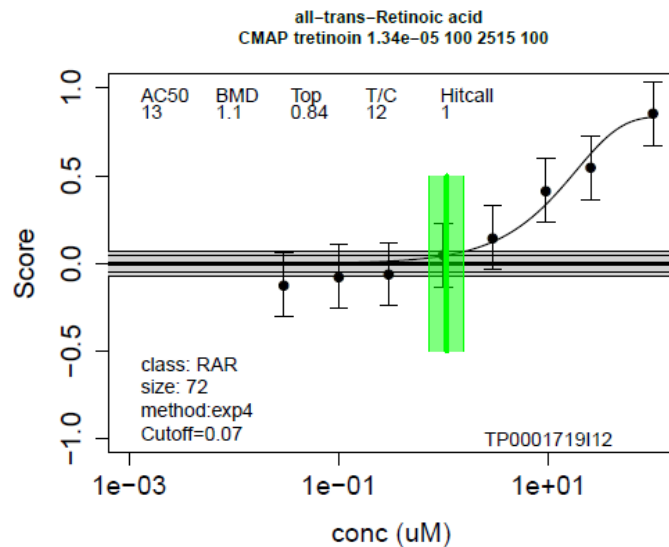
⁴ Pinero et al., *Database (Oxford)*. 2015 Apr 15;2015:bav028

Concentration-Response Modeling of Gene Signatures (2)

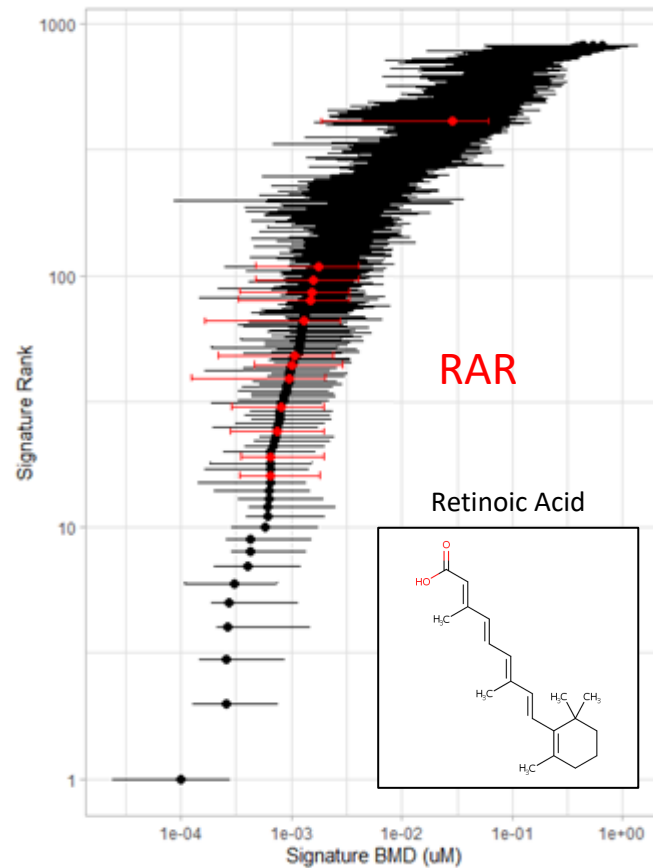
Concentration-Response Modeling (*tcp/fit2*)

Signature-Level:

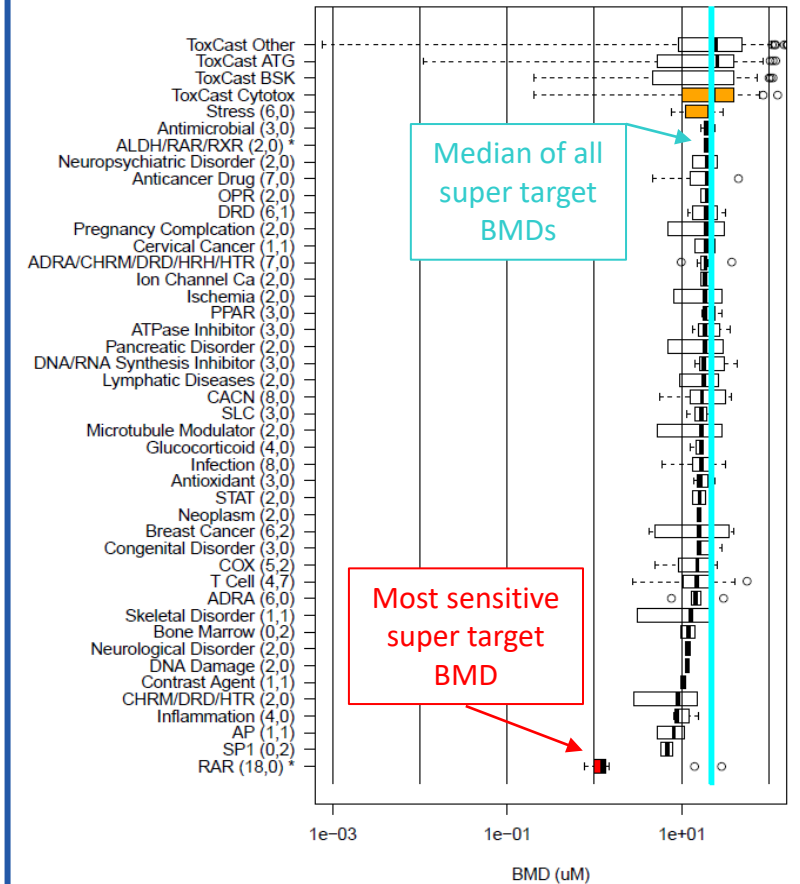
- Benchmark Dose (BMD)
- Confidence Interval on BMD
- Hit Call Probability



Ranking of Signatures

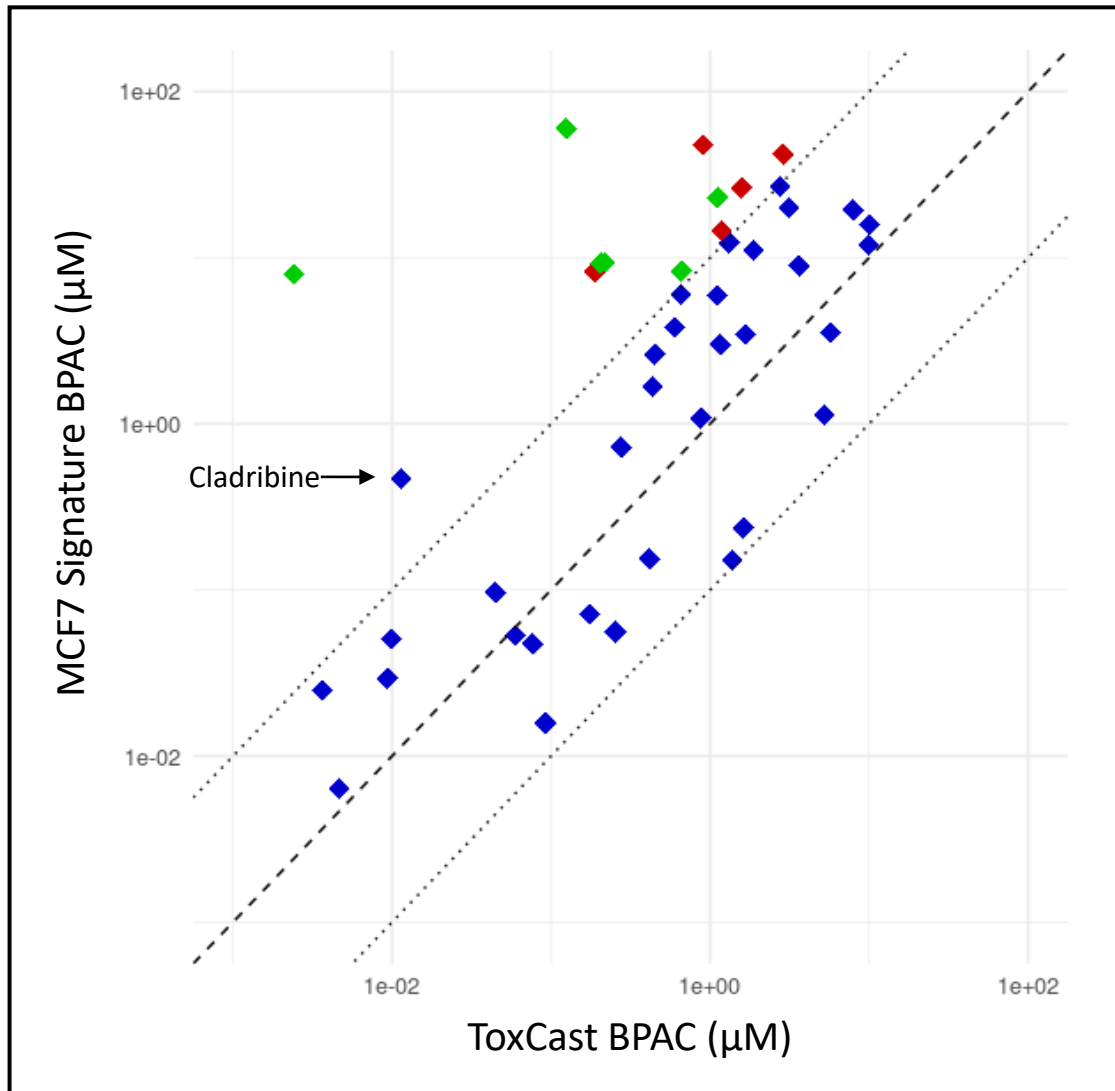


Signature Aggregation



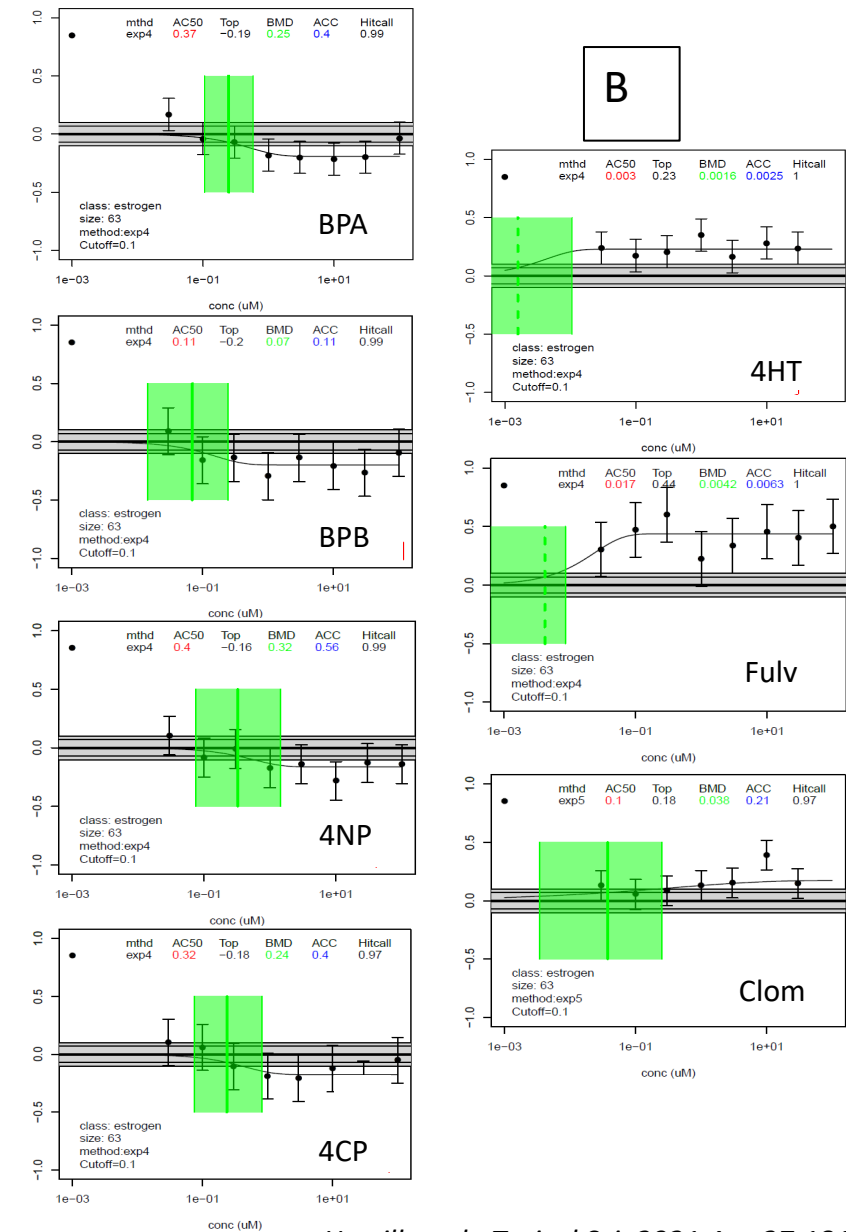
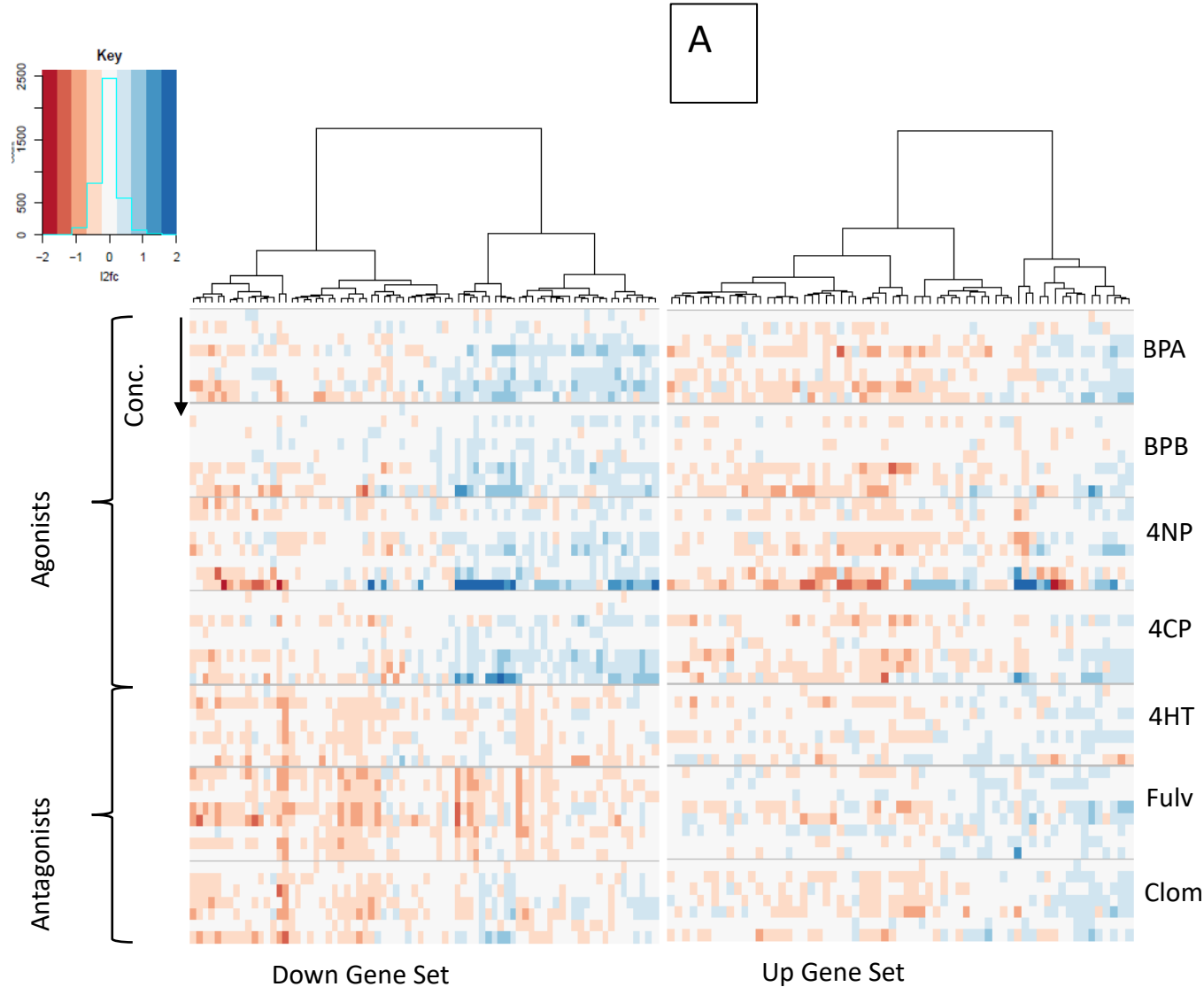
- Molecular PODs based on **biological pathway altering concentrations (BPACs)** may be derived in several ways.
- Most sensitive signature **OR** statistic based on distribution of active signatures (5th %ile) **OR** by target class.

Comparison of Transcriptional BPACs to ToxCast BPACs



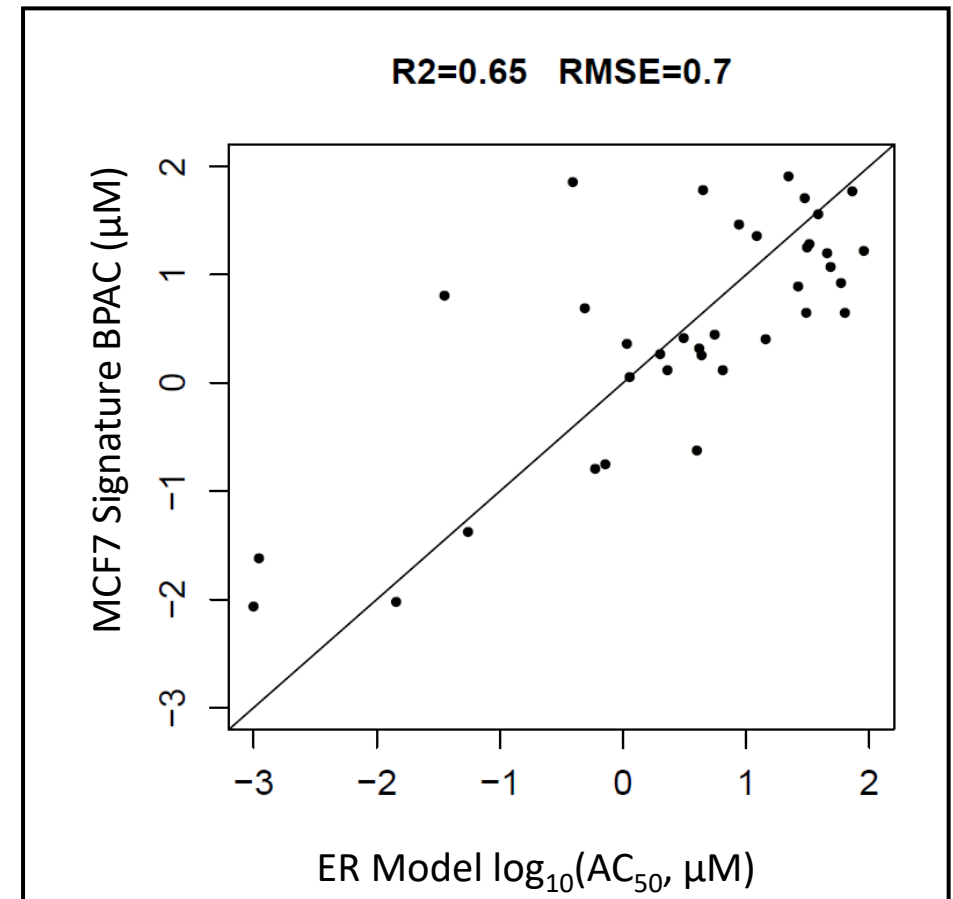
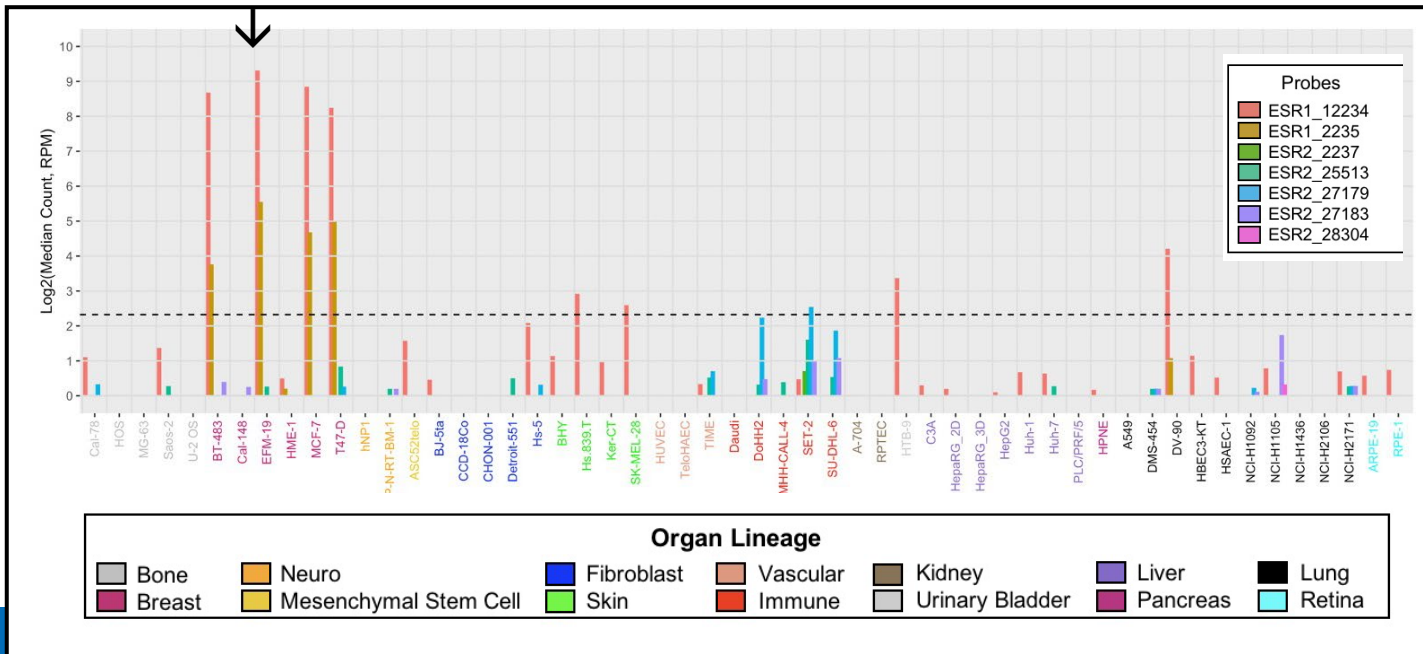
- Pilot study of 44 well-characterized ToxCast chemicals in MCF-7 cells (*Harrill et al., Toxicol Sci. 2021 Apr 27;181(1):68-89*).
- Compare HTTr-derived PODs from MCF-7 cells to previous ToxCast HTS assay results (*Paul-Friedman et al., Toxicol Sci. 2020 Jan 1;173(1):202-225*).
- Signature-based BPACs are highly concordant with ToxCast results for a majority of test chemicals in the pilot study.
 - 6 chemicals with targets that low/absent expression in MCF-7 cells.
 - 5 chemicals with off-target hit as most potent assay in ToxCast
 - Cladribine is a non-specific DNA synthesis inhibitor.

Signature Modeling of Estrogenic Chemicals

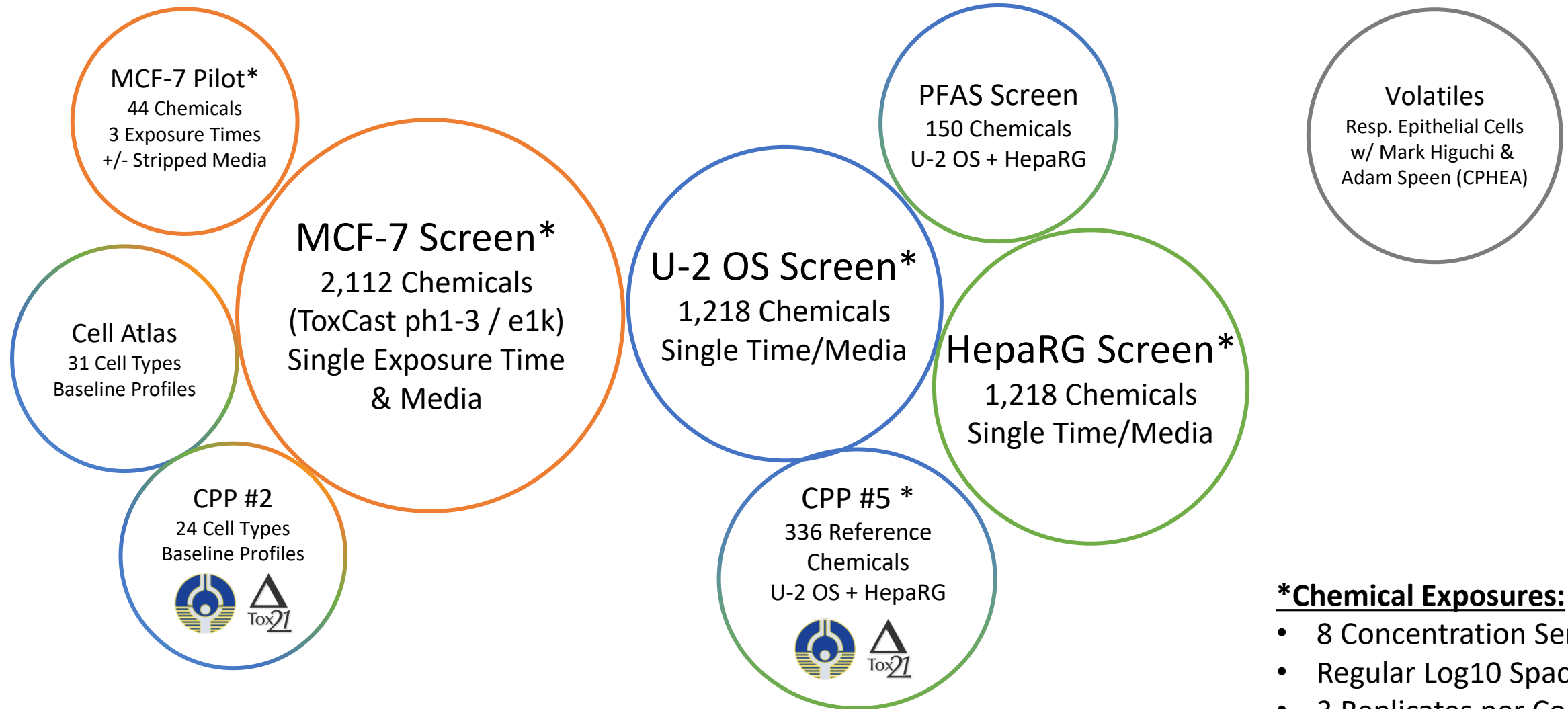


Comparison of Transcriptional BPACs to ER Model

- US EPA has developed a battery of 18 ToxCast assays to predict activity at the estrogen receptor (*Browne et al., Environ Sci Technol. 2015 Jul 21;49(14):8804-14*)
- $\log_{10} AC_{50}$ values from the ToxCast ER model assays were compared to transcriptomic signature BPACs in MCF-7 cells for a collection of 37 estrogenic chemicals.
- Signature-based BPACs are concordant with ER model predictions. →
- Estrogen receptor is also abundantly expressed in MCF-7 cells (and other breast-derived cell lines)



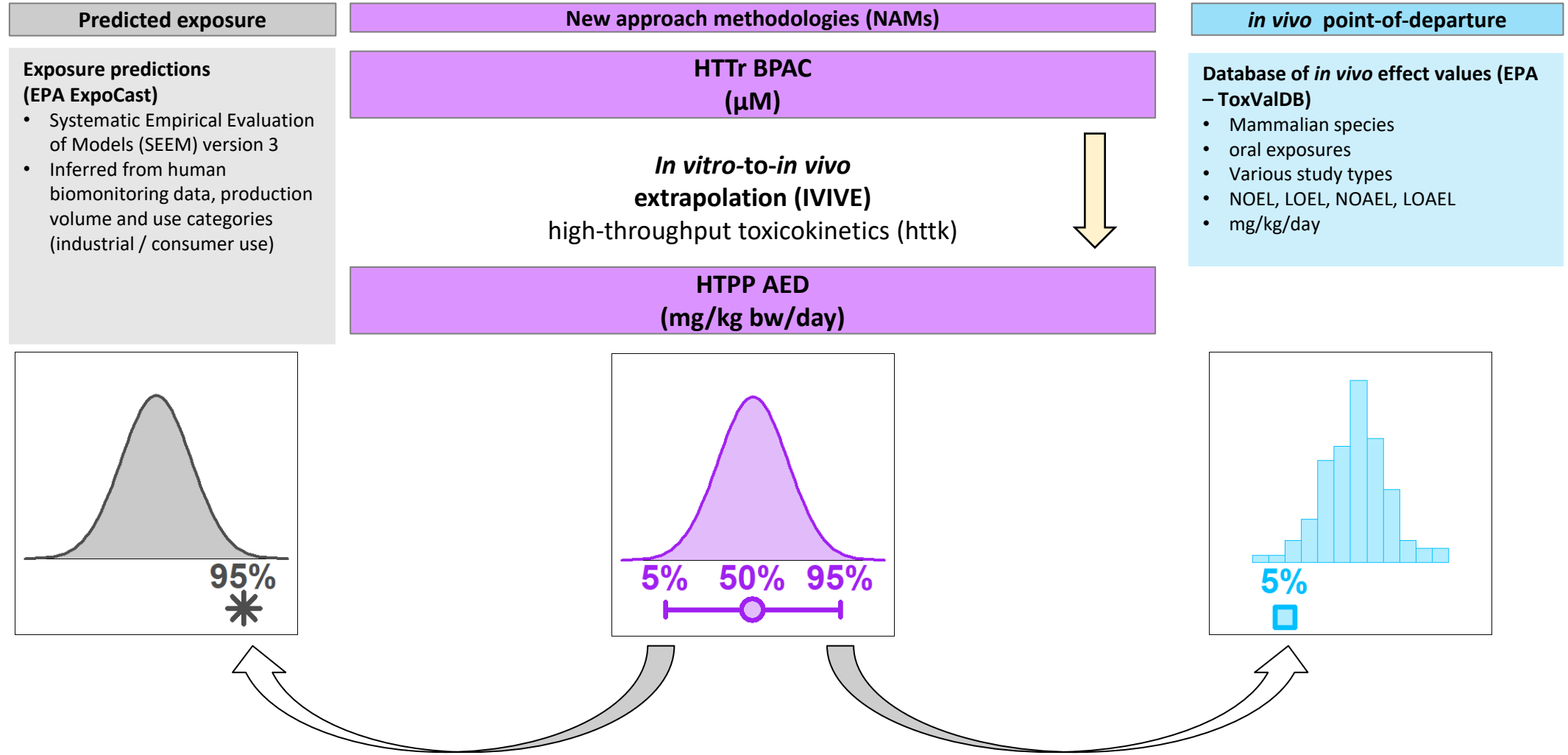
High Throughput Transcriptomics (HTTr) Data Landscape



*Chemical Exposures:

- 8 Concentration Series
- Regular Log10 Spacing
- 3 Replicates per Conc
- Randomized Plate Layout

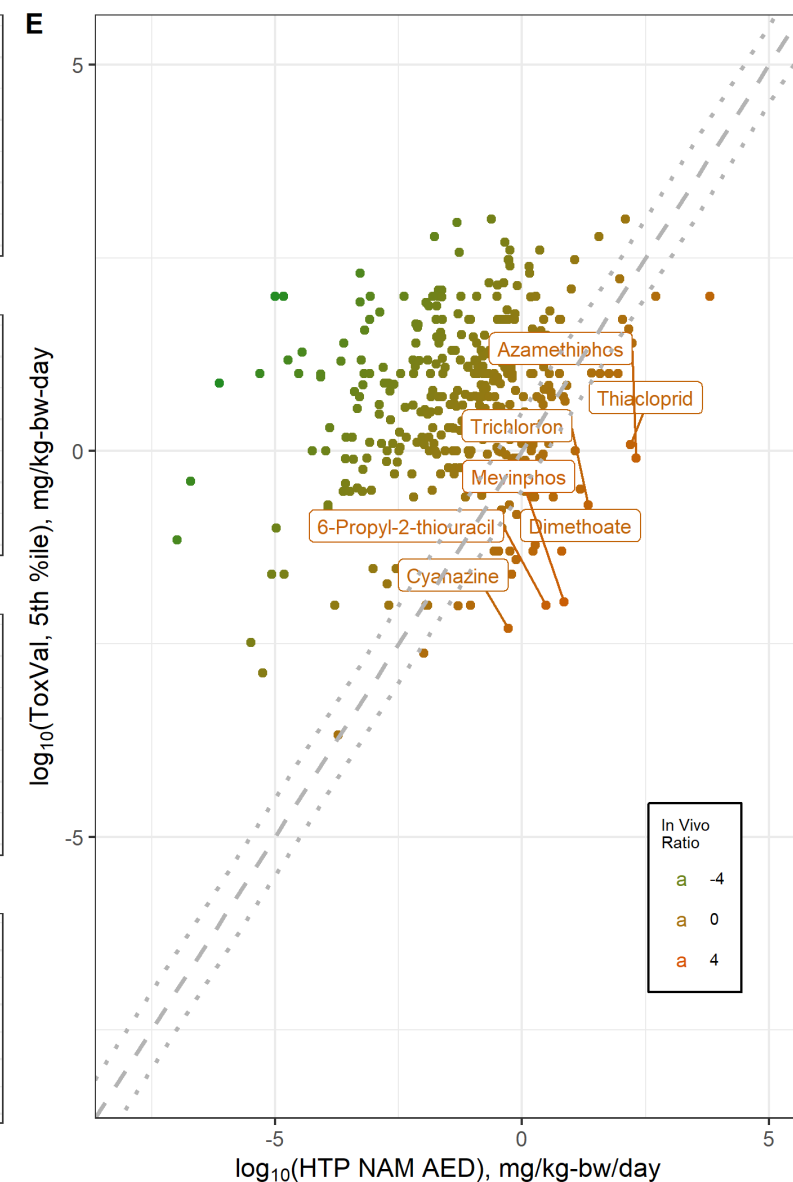
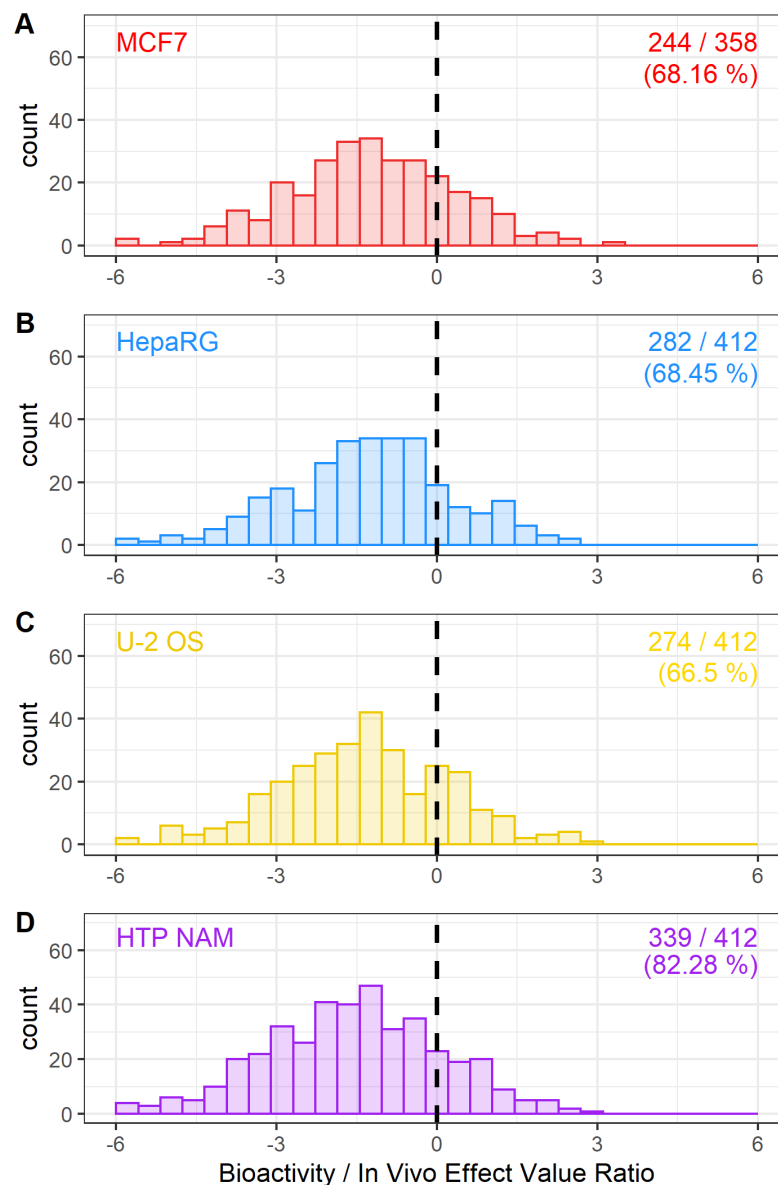
In Vitro to *In Vivo* Extrapolation (IVIVE) Using High-Throughput Toxicokinetic (httk) Modeling



Bioactivity / *In Vivo* Effect Value Ratio Analysis

- Negative ratios** indicate that AEDs derived from HTP NAMs molecular PODs are **conservative** surrogates for traditional *in vivo* PODs.
- When cell lines are considered individually, **~66-68%** of chemicals had negative ratios.
- When considered in combination, the number and percentage of chemicals with negative ratios **increased (82.3 %)**.
- Paul-Friedman et al. (2020):
 - Using ToxCast, **89 %** of APCRA chemicals had negative ratios.
- Positive ratios observed for several organophosphate and carbamate pesticides.

APCRA Chemicals



Future Directions

- **Expand chemical space**
 - Screen additional chemicals in accordance with programmatic needs / goals
- **Expand biological space**
 - Continue screening a subset of chemicals through many biologically diverse cell lines
- **Refine signature concentration-response modeling approach**
 - Reduce redundancy in signature collection
 - Continued curation of target annotation
- **Refine methods for putative target prediction & confirmation**
 - Integration with other NAM's data streams
 - Machine learning approaches
 - Bioactivity confirmation within tiered hazard evaluation framework

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