

Computational Toxicology



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The views expressed in this presentation are those of the author and do not necessarily reflect the views or policies of the U.S. EPA



Dr. Nisha S. Sipes has over 10 years of experience developing and using bioinformatics and computational toxicology approaches to translate high-throughput screening (HTS) data and other new approach methodologies (NAMs) to better estimate *in vivo* likelihood of chemical-biological interaction. She currently serves as the Assistant Center Director for Research Translations & Program/Regulatory Support for the US EPA Center for Computational Toxicology and Exposure (CCTE), where she facilitates the translation of CCTE research for use in decisions and provides scientific and technical expertise to internal and outside stakeholder groups.

Learning Objectives

- Why Alternatives are Needed
- Understand Computational Toxicology Concepts, Tools, and Approaches
 - New Approach Methodologies (NAMs)
 - Read-across
 - High-throughput *in vitro* assays
 - *In vitro* to *in vivo* extrapolation
 - Rapid exposure predictions
- Computational Toxicology in Practice and Potential Use

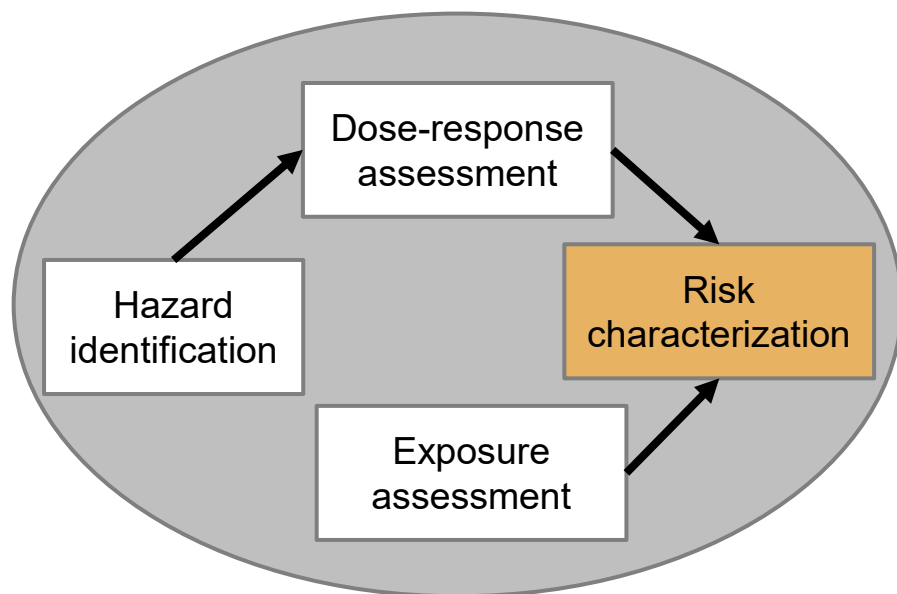
What Do You Need for Traditional Chemical Risk Characterization?

Data needs

- Chemical characterization
- Hazard; human health and ecological data, *in vivo* data, biological targets (effect), dose-response characterization (dose)
- Toxicokinetics
- Exposure; exposure scenarios, exposure levels

How to obtain

- Animal models + exposure sampling



Why Is There a Need for Alternative Approaches?

- Too many chemicals to test with standard animal-based methods
 - >40,000 active substances on US EPA Toxic Substances Control Act (TSCA) inventory
<https://www.epa.gov/assessing-and-managing-chemicals-under-tsca>
 - We do not have detailed exposure information
- Traditional toxicity testing is costly and time consuming
 - Natural or industrial disasters (e.g., Gulf of Mexico oil spill)
- Traditional animal-based testing has issues related to ethics and relevance
 - Mechanistic understanding
 - Physiology comparisons (e.g., respiratory physiology in rats)
- Looking into new ways to address these problems

Computational Toxicology

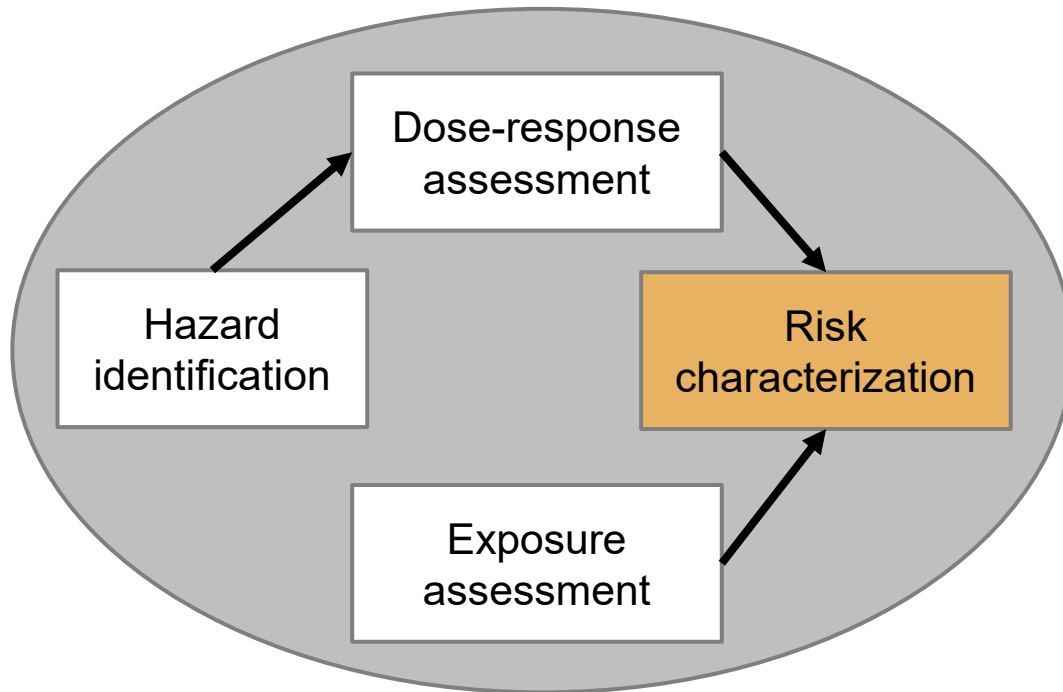
- Gathering, integrating, and evaluating data and information using mathematical and computer-based approaches to better understand chemical hazards and risks to human health and the environment
- Typically refers to non—*in vivo* toxicological tools and approaches
 - New Approach Methodologies (NAMs)—*in silico*, *in chemico*, *in vitro*, hazard + exposure
- Some tools and approaches are already used in hazard and risk assessments
 - E.g., Quantitative Structure-Activity Relationships (QSARs), *in vitro* assays used in lieu of *in vivo* assays

New Approach Methodologies (NAMs)

National Academies of Sciences, Engineering, and Medicine 2017
<https://doi.org/10.17226/24635>.

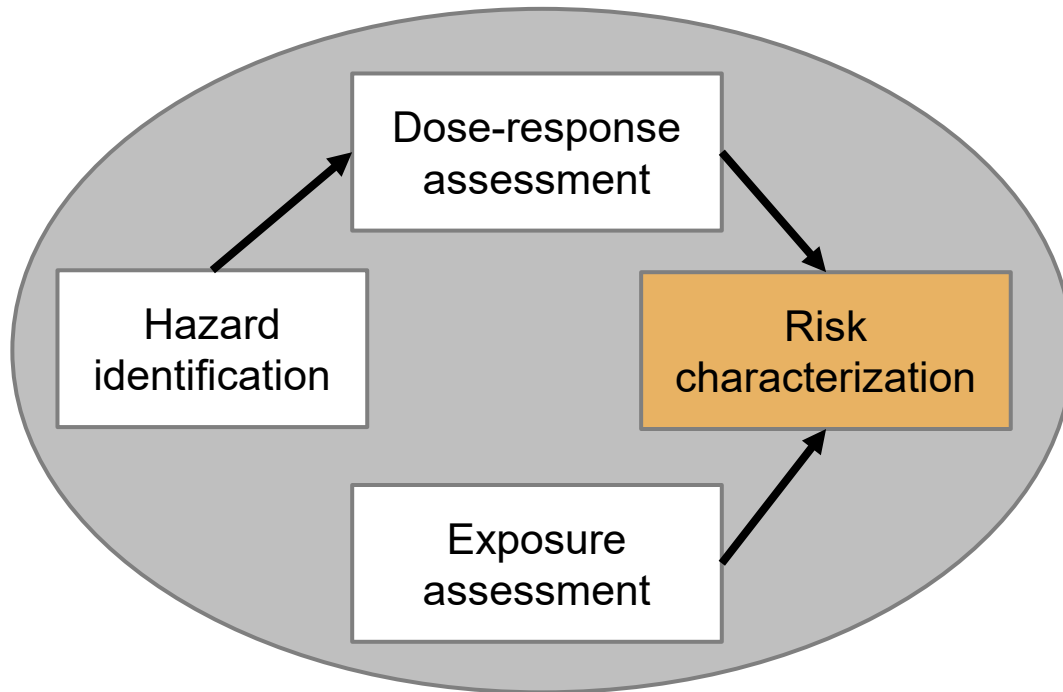
- Adverse outcome pathways (AOPs)
Pathway identification and knowledge integration
- *In silico* (e.g., QSAR and read-across)
Estimate effects and doses
- *In vitro* assays
 - *Broad / screening (transcriptomics, cell painting)*
 - *Targeted (receptors, enzymes)*
 - *In vitro PODs, modes/mechanisms of action*
- *In vitro* toxicokinetics
Allow conversion of an in vitro POD to in vivo (IVIVE)
- Computer models
Integrate multiple in silico and in vitro data streams
- Databases of existing traditional toxicology data
Enables training and validation of NAM models

How Can We Use Alternative Approaches in Risk Characterization?



1. *In silico* read-across (data gap analysis)
2. Hazard assessment (ID and dose-response)
 - a. *In vitro* assays
 - b. Making sense of *in vitro* potencies using *in vitro* to *in vivo* extrapolation (IVIVE)
3. High throughput exposure assessment
4. Risk characterization



How Can We Use Alternative Approaches in Risk Characterization?



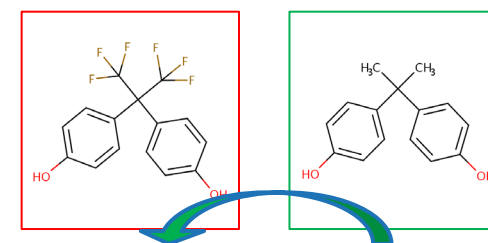
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In Silico Read-Across (Data Gap Analysis)

- Read-across describes the method of filling a data gap whereby a chemical with existing data (source analogue) is used to make a prediction for a “similar” chemical (target chemical)
- Need curated chemical structures, physical-chemical properties
- Several freely available tools
 - Including GenRA in the EPA CompTox Chemicals Dashboard

	Target chemical	Source analogue
Property		
	Missing data	Reliable data

Analogue identification



Acute toxicity?

Predicted

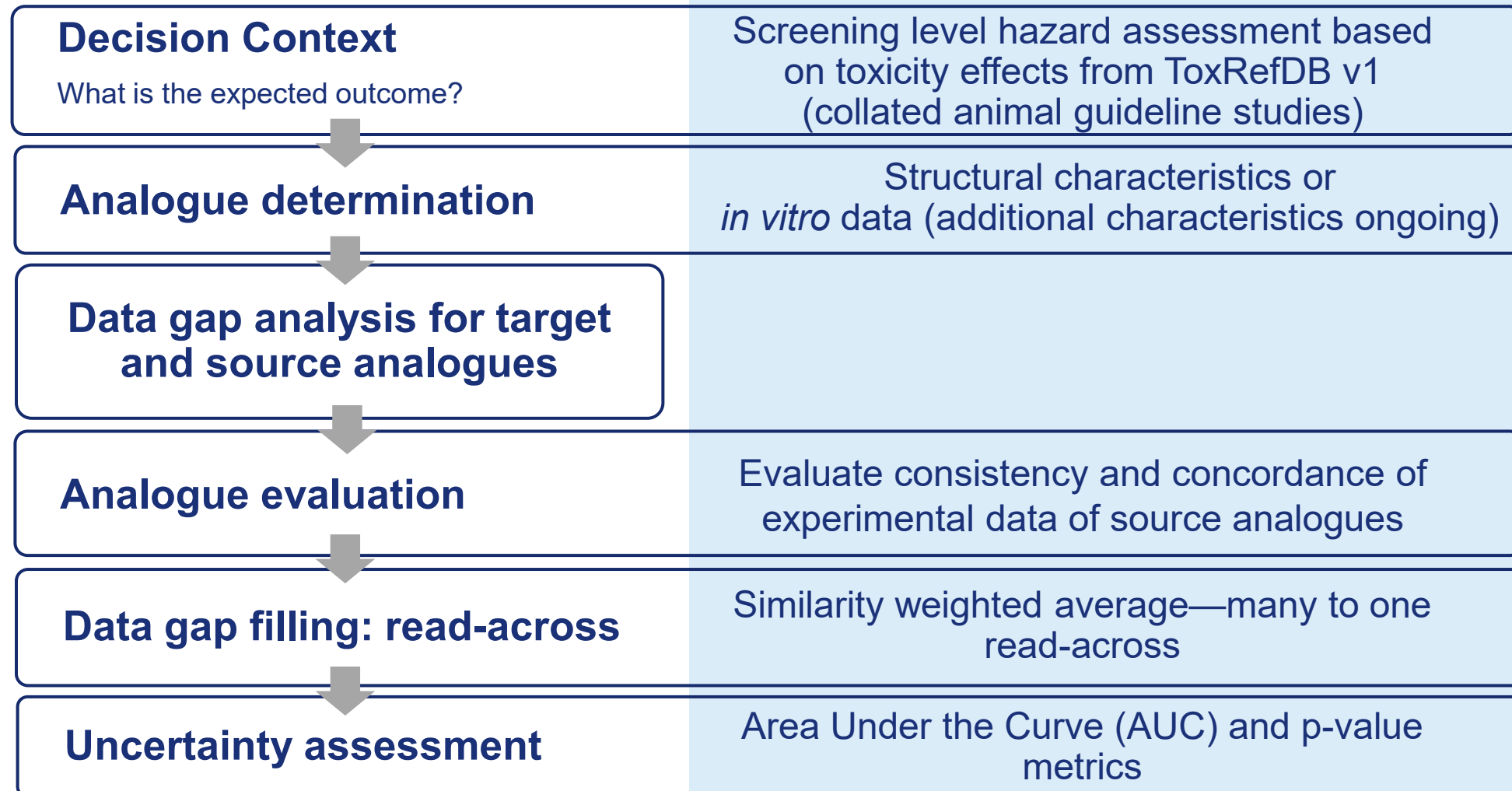
Known

G Patlewicz, et. al., Comput Toxicol. 2017;3:1-18. doi:10.1016/j.comtox.2017.05.003
Shah I, et al., Regul Toxicol Pharmacol. 2016 Aug;79:12-24. doi: 10.1016/j.yrtph.2016.05.008

Read-Across Workflow

Specific for US EPA GenRA v1.0

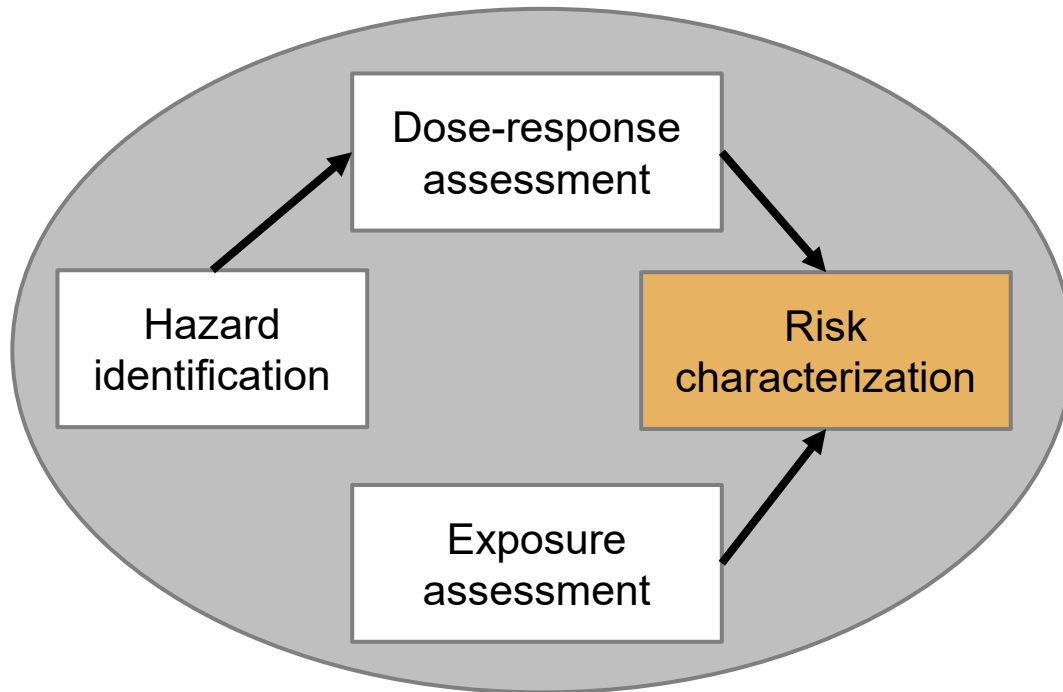
Generalized read-across (GenRA)



Read-Across in Practice

- Valuable for chemical safety assessment
 - Read-across acceptance for regulatory purposes remains an issue
 - Difficulties addressing residual uncertainties
 - Subjective, expert-driven assessment
 - Need more experience using these tools
- GenRA is an attempt to move toward an objective read-across approach where uncertainties and performance can be quantified
- Organisation for Economic Co-operation and Development (OECD)
 - Guidance on Grouping of Chemicals (No.194, 2014)
[http://www.oecd.org/officialdocuments/displaydocument/?cote=env/jm/mono\(2014\)4&doclanguage=en](http://www.oecd.org/officialdocuments/displaydocument/?cote=env/jm/mono(2014)4&doclanguage=en)
 - Integrated approaches to testing and assessment (IATA) case studies

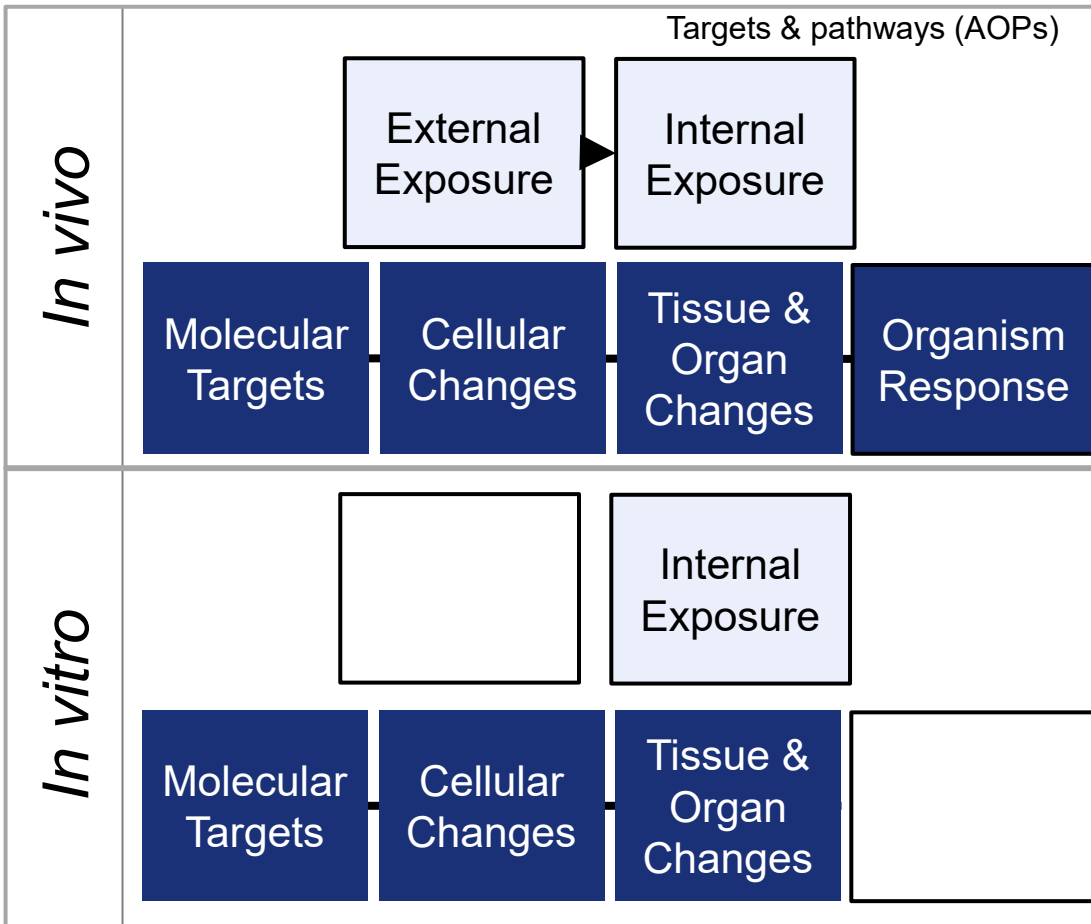
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In Vitro Assay Hazard Identification and Dose-Response

Hazard identification

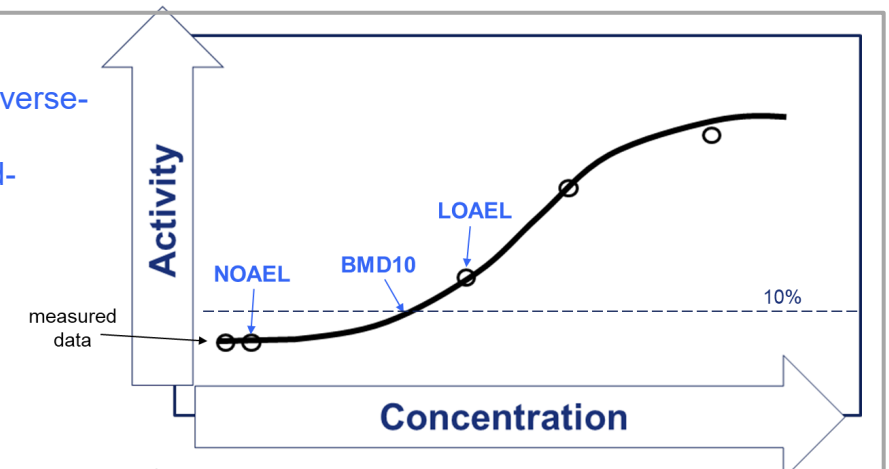


Dose-response

Point of Departure (POD) corresponds to an estimated lowest effect level

POD

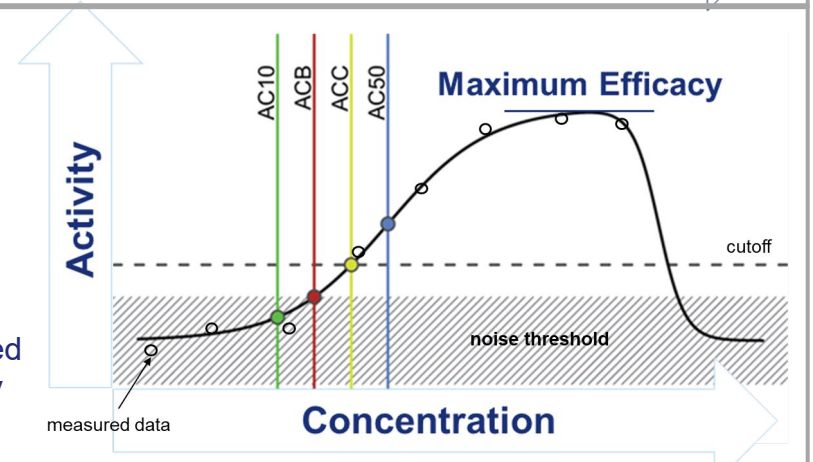
- **NOAEL** no-observed-adverse-effect level
- **LOAEL** lowest-observed-adverse-effect level
- **BMD** benchmark dose
 - *Predetermined threshold*



POD

- **AC10** activity concentration at 10% of maximal activity
- **ACB** activity concentration at baseline (noise)
- **ACC** activity concentration at cutoff (significance)

Across multiple assays—determined for the most sensitive *in vitro* assay



Summary of *In Vitro* Assay Types for Hazard Evaluation

- High-Throughput Screening assays (HTS)
- High-Throughput Transcriptomics (HTTr)
- High-Throughput Phenotypic Profiling (HTPP)
 - High Content Screening (HCS)
- Organotypic Models
- Microphysiological Systems (MPS)
- Small Model Organisms

High-Throughput Screening (HTS) Assays

in vitro assay

- Easily screens hundreds to thousands of chemicals
- Typically targeted to measure specific chemical-target interactions (e.g., individual receptor, enzyme reporter assays)
- Takes a coordinated (preferably automated) data analysis pipeline

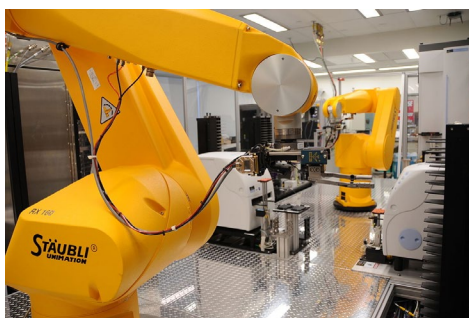
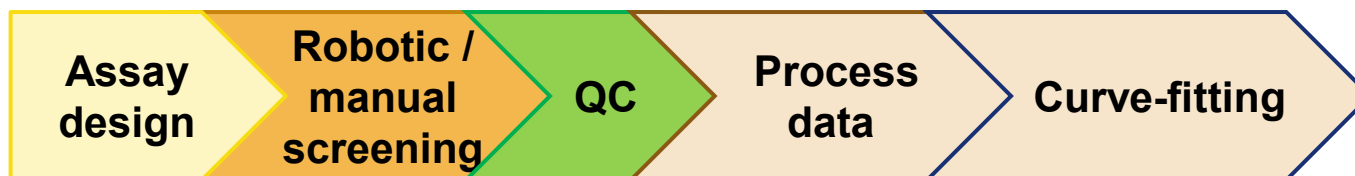


Image:
<https://ncats.nih.gov/news/releases/2018/tox21-strategic-plan>

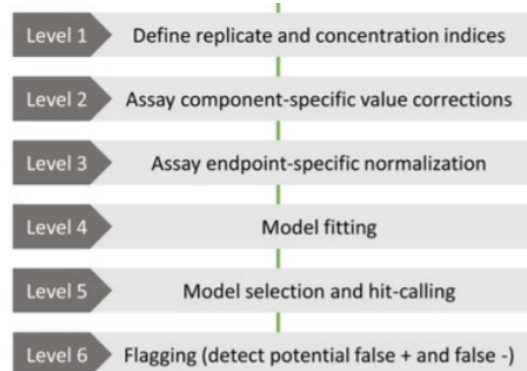
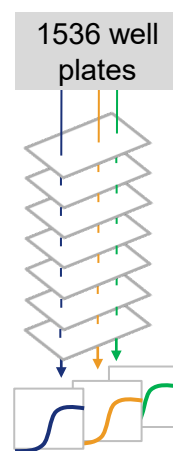
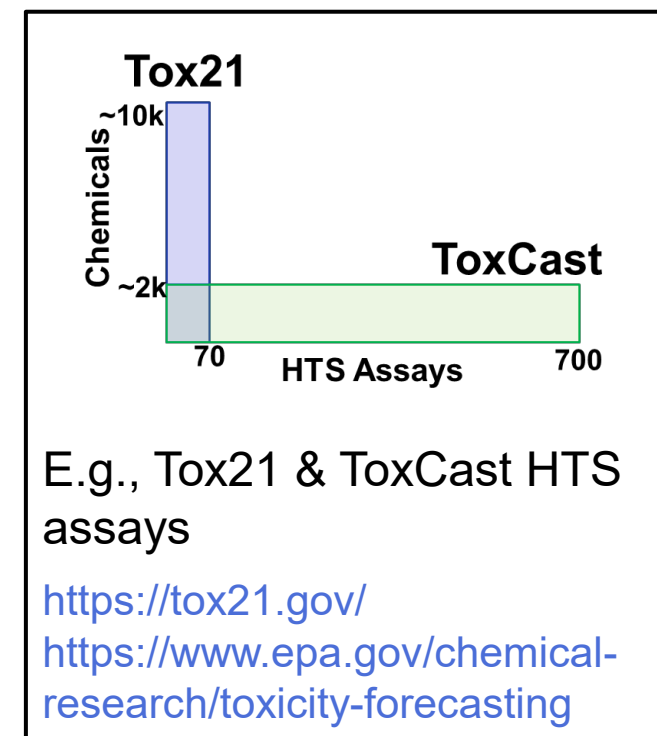
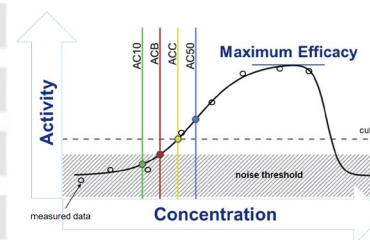


Image:
<https://www.epa.gov/sites/production/files/2018-04/documents/toxcastownermanual4252018.pdf>



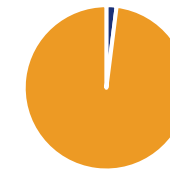
Limitations: need *a priori* knowledge of molecular targets; incomplete coverage of important pathways (i.e., biological space)

High-Throughput Transcriptomics (HTTr)

in vitro assay

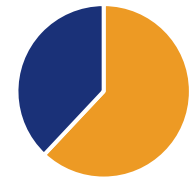
- Whole transcriptome assay measures expression of ~20,000 transcripts at once
- Increasing biological coverage over single reporter assays
- Low cost, uses purified RNA samples or cell lysates
- Scalable, targeted assay: measure transcript of interest, greater throughput than RNA-Seq, attenuate highly expressed genes

Gene Coverage

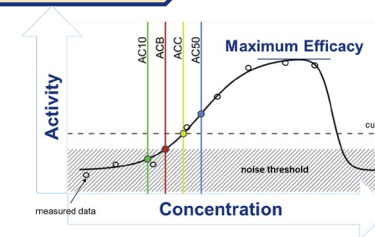
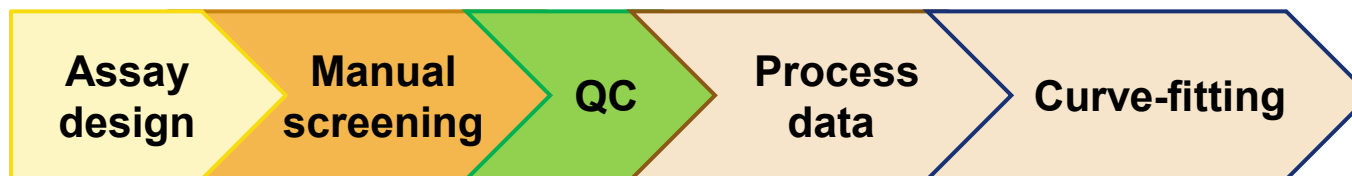


■ ToxCast
■ Not in ToxCast

Pathway Coverage*



* At least one gene from pathway represented

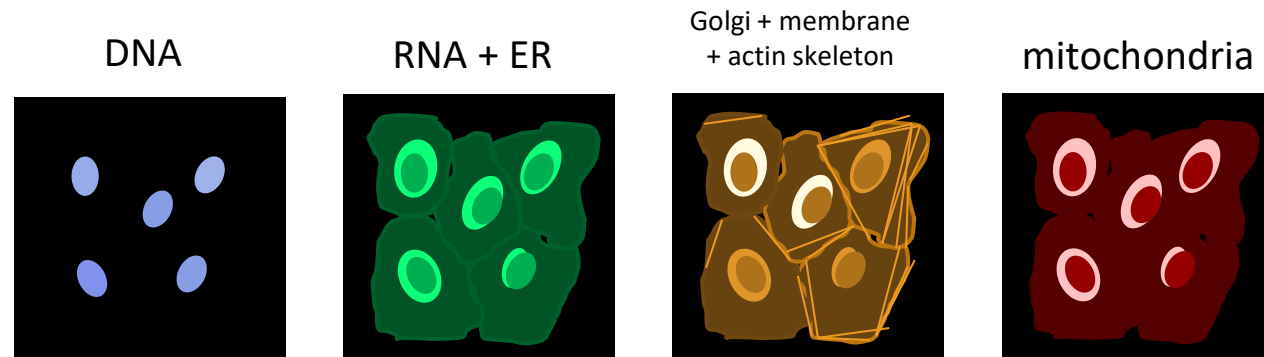


Limitation: only reveals transcript expression changes

High-Throughput Phenotypic Profiling (HTPP)

in vitro assay

- Image-based phenotypic profiling measures a large variety of morphological features of individual cells in *in vitro* cultures
- Successfully used for functional genomic studies and in the pharmaceutical industry for compound efficacy and toxicity screening



Flourescent labels
DNA: H-33342
RNA: SYTO14
ER: Concanavalin A-488
Actin: Phalloidin-568
Golgi + Membrane: wheat germ agglutinin (WGA) -555
Mitochondria: MitoTracker

shape

intensity



localization

texture

1300 features

Computational processing



← **profile**

for each chemical x concentration

High Content Screening
Cell Painting

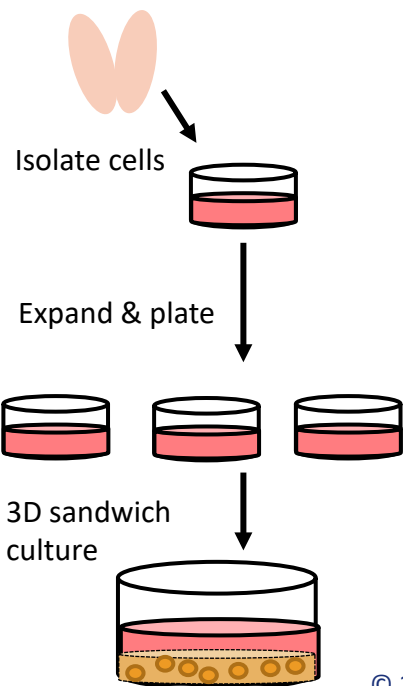
Limitation: does not identify
tissue/organ effects

Organotypic Models and Microphysiological Systems (MPS) *in vitro* assay

Three-dimensional cellular *in vitro* models

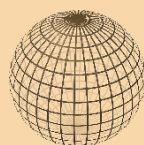
E.g., Organotypic model

Normal human thyroid gland



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commons.wikimedia.org/wiki/File:Sphere_wireframe_10deg_6r.svg

spherical



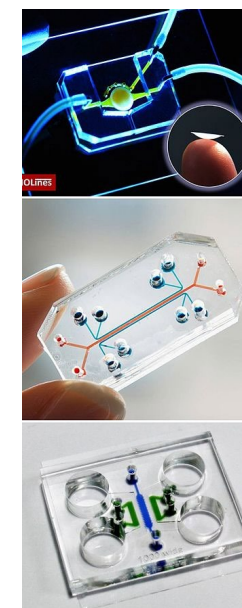
Hollow on cross section



Limitation: low throughput;
more difficult to develop; not
yet an integrated system

Interconnected *in vitro* models in
microphysiologically relevant “chips”

E.g., MPS or Organ-on-a-chip



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https://commons.wikimedia.org/wiki/File:Organ_on_a_chip.jpg

Small Model Organisms

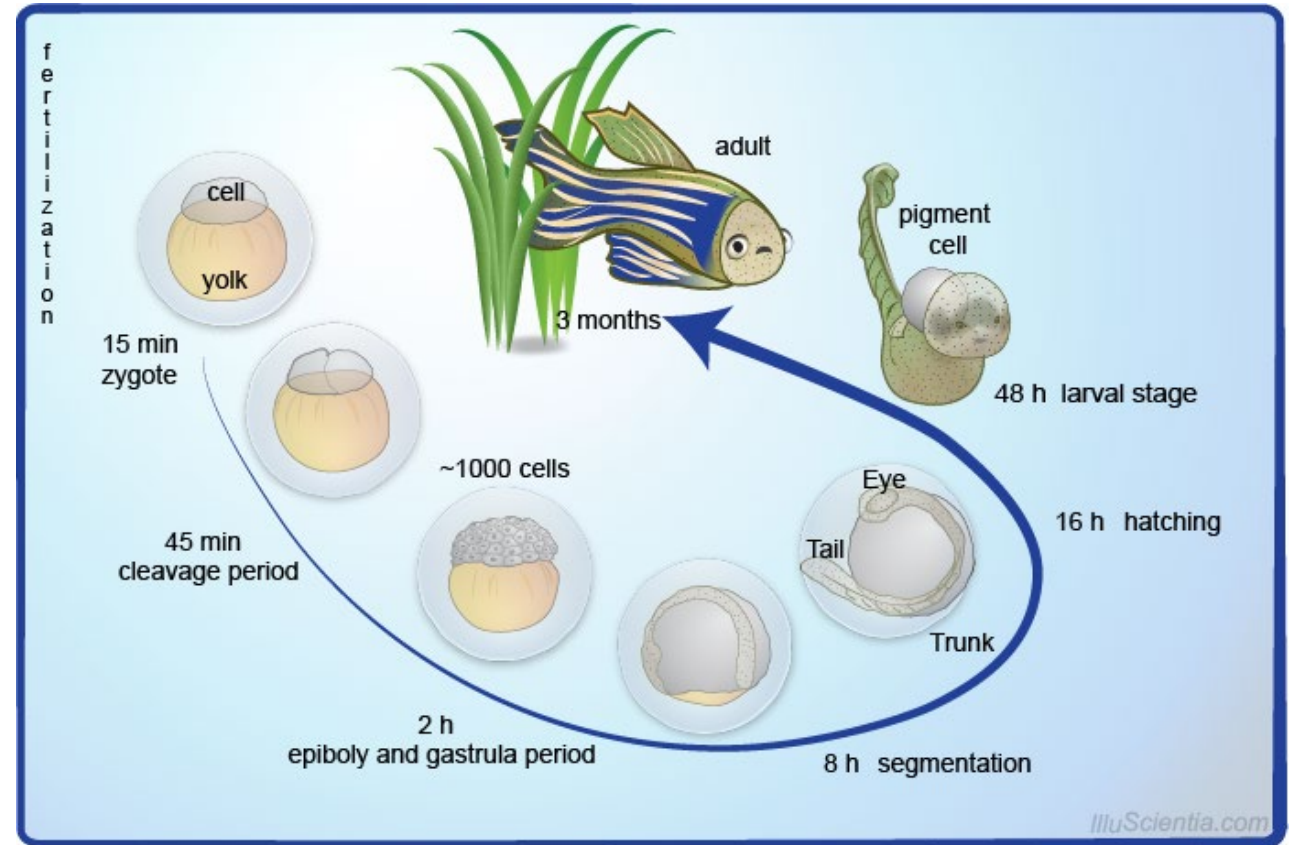
Danio rerio (zebrafish)

Drosophila melanogaster (fruit fly)

Daphnia (water flea)

C. elegans (round worm)

- Integrated model
- Ease of genetic manipulation
- Drug screening
- Reproductive and Developmental Toxicity

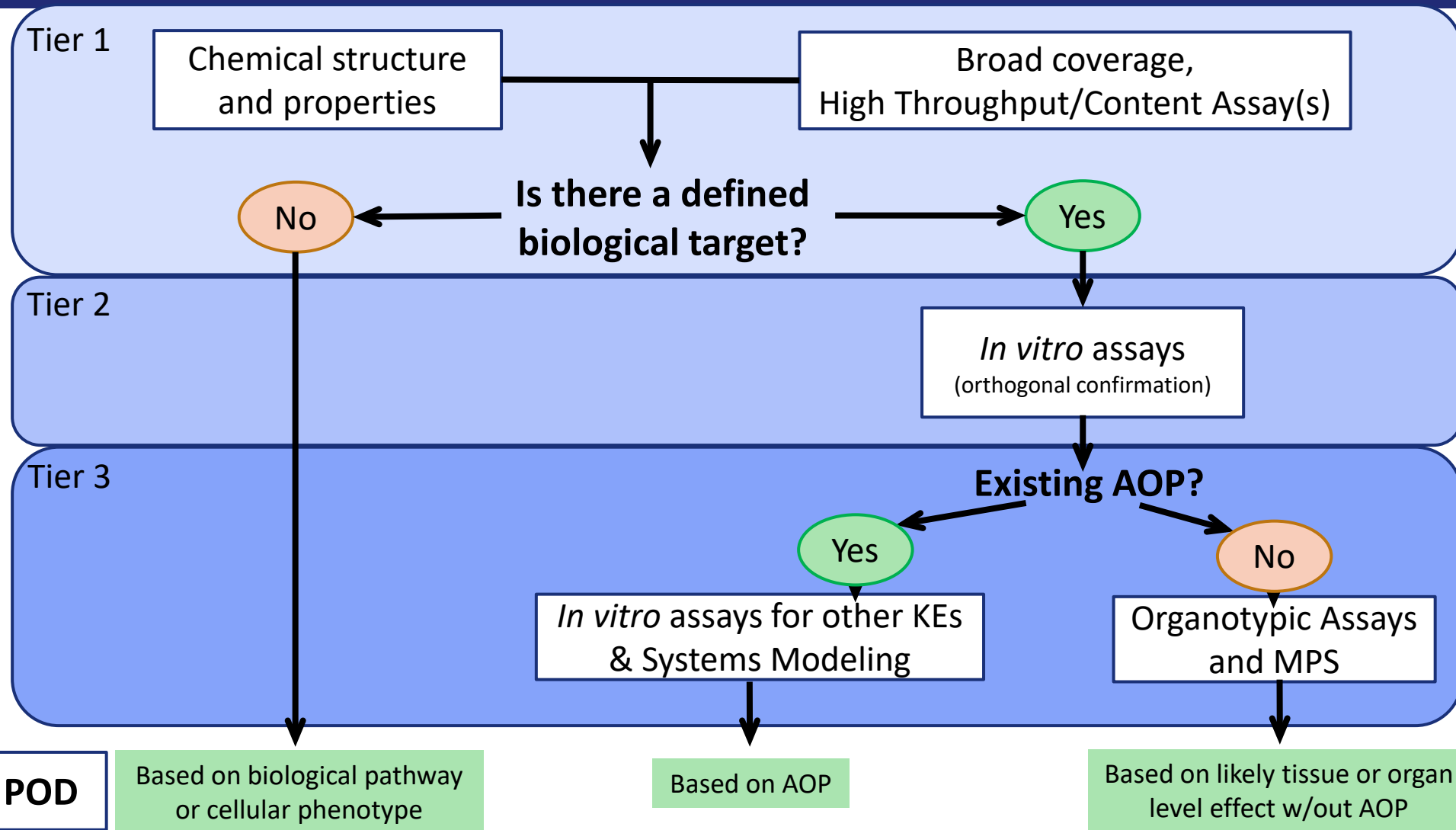


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<https://creativecommons.org/licenses/by/3.0/legalcode> ; https://commons.wikimedia.org/wiki/File:Zebrafish_Cycle.png

In Vitro Assays in Practice

- Valuable for chemical safety and risk assessment
 - Routinely used by industries and regulatory authorities
 - OECD
 - Skin Irritation
 - Serious Eye Damage/Eye Irritation
 - Sensitization
 - Genotoxicity
- } Can replace some traditional *in vivo* animal tests
- Potential replacement for uncertainty factors
 - Limitations
 - Metabolism *in vitro*
 - Non-specific binding to plastics in *in vitro* system
 - Uncertainty analysis*
 - Need analysis pipeline and integration approaches
 - Need international involvement and case studies
- *same with animal studies

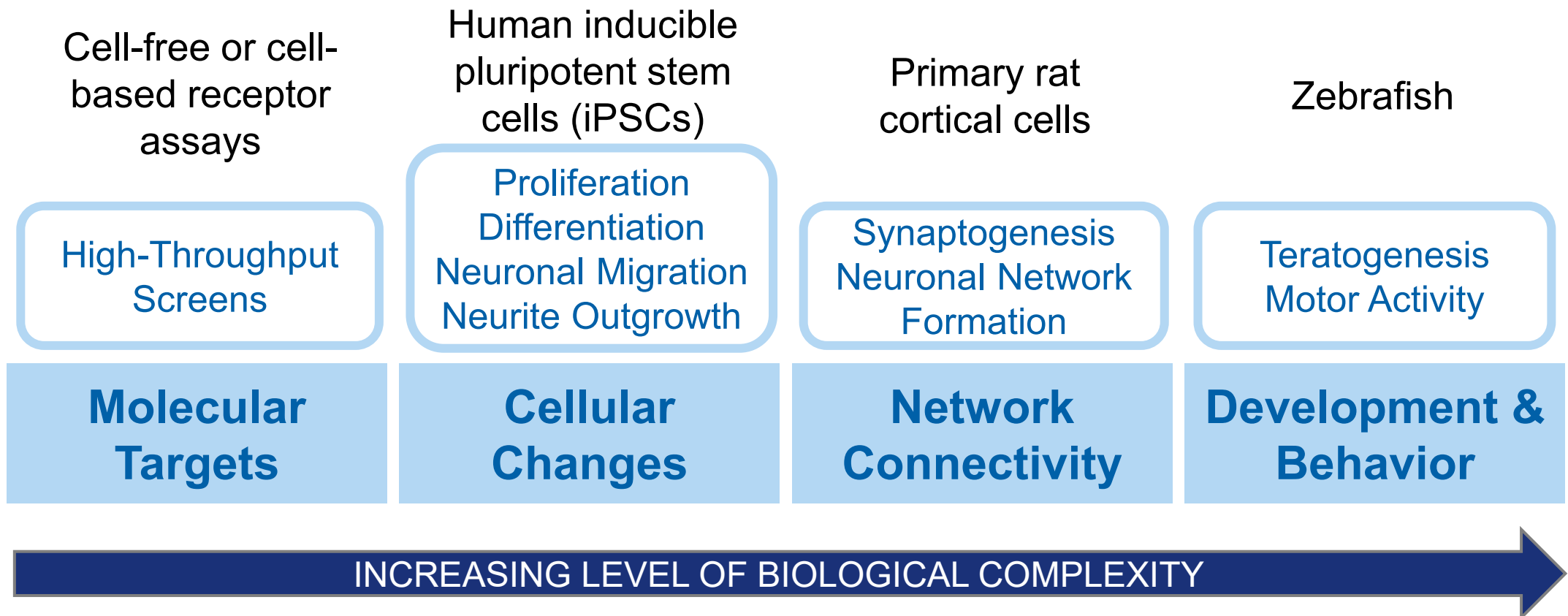
Example of a Tiered Hazard Evaluation Approach





Can You Imagine a Way to Integrate *In Vitro* Assays into Your Own Assessments? **What about for Developmental Neurotoxicity (DNT)?**

- No standard regulatory data requirements for DNT
- Resource intensive, difficult to interpret
- ~100–150 chemicals with DNT toxicological hazard information
- We know pathways for neurodevelopment



International Effort to Improve DNT Testing

- International Collaboration (e.g., EFSA, OECD, US EPA) EFSA—European Food Safety Authority
 - OECD DNT Expert Group OECD—The Organisation for Economic Co-operation and Development
 - Improve DNT testing
 - Incorporate mechanistic knowledge
 - Provide regulatory relevant examples through case studies
 - Accelerate regulatory uptake of the DNT *in vitro* battery
1. Adverse outcome pathways (AOPs)
 2. *In vitro* battery
 3. OECD Integrated Approaches to Testing and Assessment (IATA)

International Discussion to Increase NAM Use

OECD Integrating Approaches to Testing and Assessment (IATA)

“IATA are pragmatic, science-based approaches for chemical hazard characterisation that rely on an integrated analysis of existing information coupled with the generation of new information using testing strategies.”

<http://www.oecd.org/chemicalsafety/risk-assessment/iata-integrated-approaches-to-testing-and-assessment.htm>

Case studies are critical to acceptance

Problem formulation

- Define the regulatory need
- Identify relevant information

Gather and evaluate existing data

- *In vivo, in vitro, in silico*

Weight of Evidence

- Characterize uncertainty

...ce, consider additional info

Endocrine-Disrupting
Chemicals (EDC)

...as

...al methods

Adapted from OECD

[http://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=ENV/JM/MONO\(2020\)25&docLanguage=en](http://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=ENV/JM/MONO(2020)25&docLanguage=en) ;

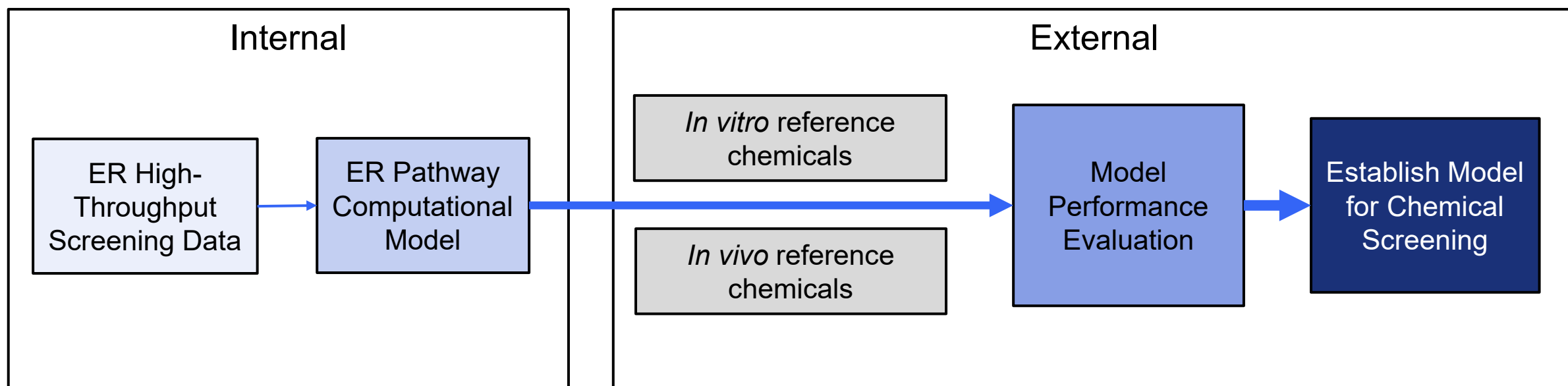
<http://www.oecd.org/chemicalsafety/risk-assessment/iata-integrated-approaches-to-testing-and-assessment.htm>

Endocrine-Disrupting Chemicals (EDCs)

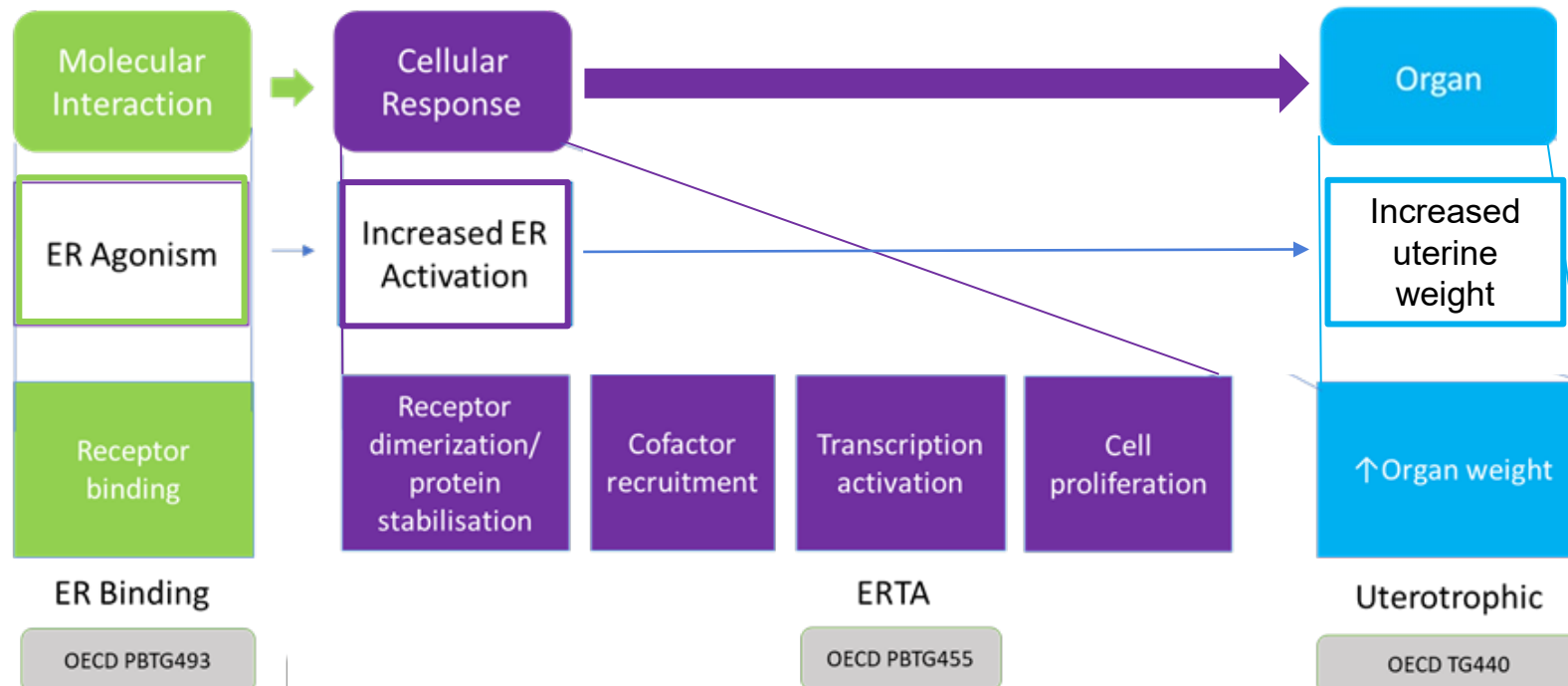
- EDCs are a diverse set of substances that have the potential to interfere with normal endocrine function and lead to an adverse outcome
- Regulatory agencies in many countries evaluate endocrine activity of environmental chemicals for specific regulatory endpoints
- US EPA Endocrine Disruptor Screening Program (EDSP) uses a two-tiered testing battery approach
 - Tier 1 screens for potential to interact with estrogen, androgen, or thyroid hormone
 - Running Tier 1 battery costs ~\$1 million / chemical
 - Tier 1 tests on 52 chemicals over 6 years, ~10,000 chemicals on EDSP Universe list
 - Tier 2 tests to verify the interaction and quantify dose-response relationship
- IATA EDC Case Study
 - Identification of endocrine disruption via estrogen receptor agonism by a substance

} Need for alternative approaches

Overall Approach



Adverse Outcome Pathway (AOP) for EDCs



Adverse Outcome Pathway (AOP) for EDCs

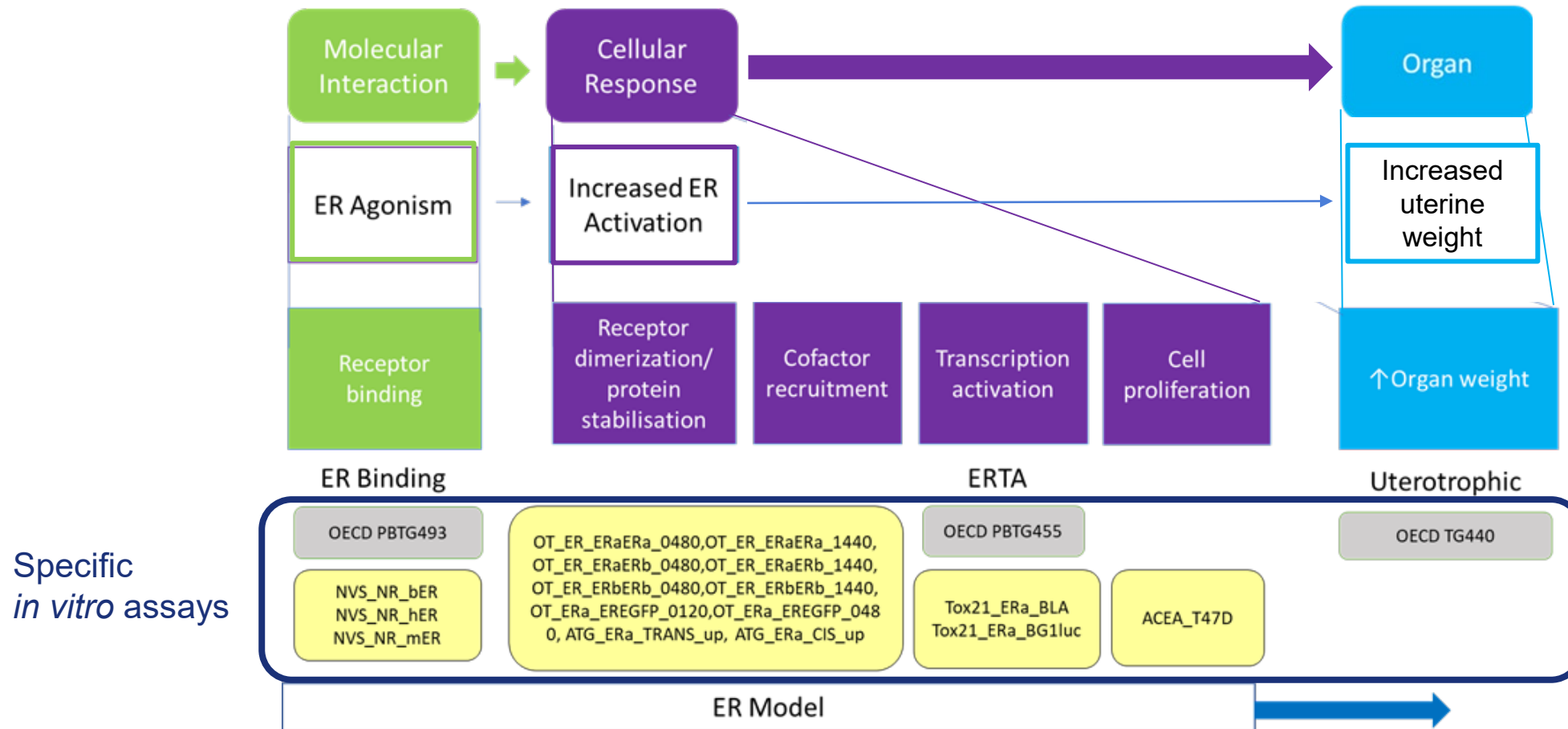


Image used with author permission and adapted from Browne P, Noyes PD, Casey WM, Dix DJ.
Environ Health Perspect. 2017 Sep 1;125(9):096001. doi: 10.1289/EHP1304.

ER Model from *In Silico* Aggregation of HTS Assays

- Orthogonal assays on pathway
 - Different technologies
 - Different points in pathway
- No assay is perfect
 - Assay interference
 - Noise
- Use computational model to integrate assays
- Model creates a composite dose-response curve for each chemical to summarize results from all assays
- Evaluate model against reference chemicals

Evaluation Using *In Vitro* & *In Vivo* Reference Chemicals

In vitro

- OECD Test Guideline 457 BG1 ER Transactivation Guidance document: <https://doi.org/10.1787/9789264185395-en>

Model Performance

40 chemicals

Accuracy: 93% (95%)

Sensitivity: 93% (93%)

Specificity: 92% (100%)

*Values in parentheses exclude chemicals w/inconclusive model scores

In vivo

- Comprehensive literature search identified 103 chemicals; however...
 - **Uncertainty in *in vivo* guideline data**
 - 26% of chemicals tested multiple times in the uterotrophic assay gave discrepant results

Model Performance

43 chemicals

Accuracy: 86% (95%)

Sensitivity: 97% (97%)

Specificity: 67% (89%)

*Values in parentheses exclude chemicals w/inconclusive model scores

Outcome: Risk Assessment Guidance

EPA notice: “The approach incorporates validated high throughput assays and a computational model and, based on current research, can serve as an alternative for some of the current assays in the Endocrine Disruptor Screening Program (EDSP) Tier 1 battery.”

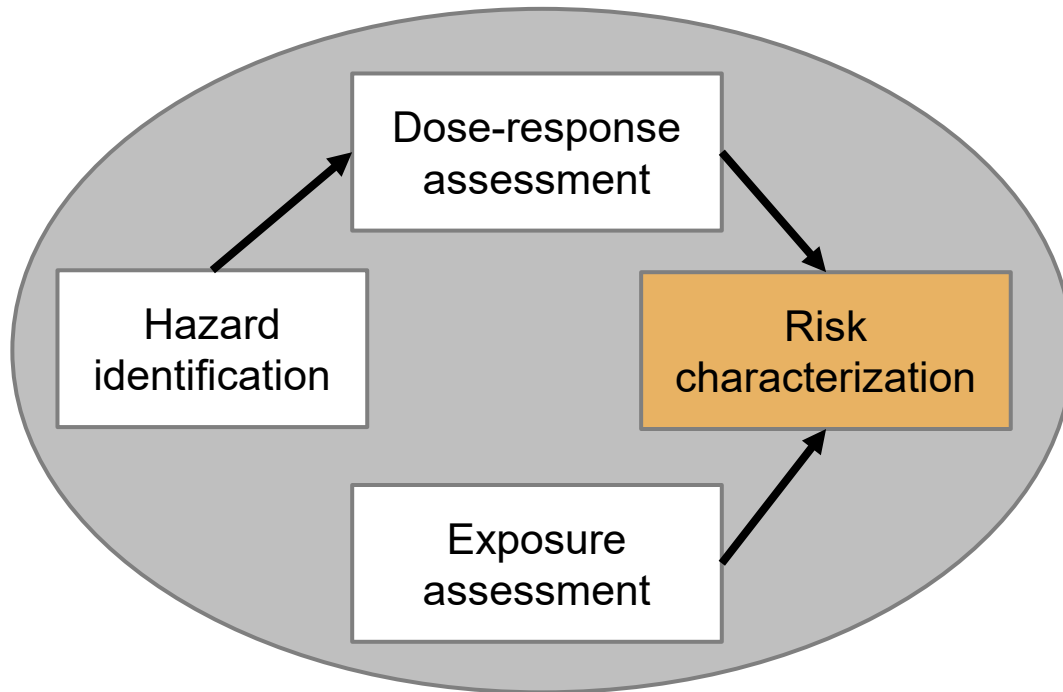
US Environmental Protection Agency (2015) National Archives Federal Register PA-HQ-OPPT-2015-0305

OECD:

- Integrated Approach for Testing Assessment (IATA) for the ER assays was reviewed and published
- Assays incorporated in an annex of Test No. 455, covering all ER transcriptional assays
 - Test Guidelines give more specific details on how to run each assay and combine the results and will take multiple years for full guideline to be made

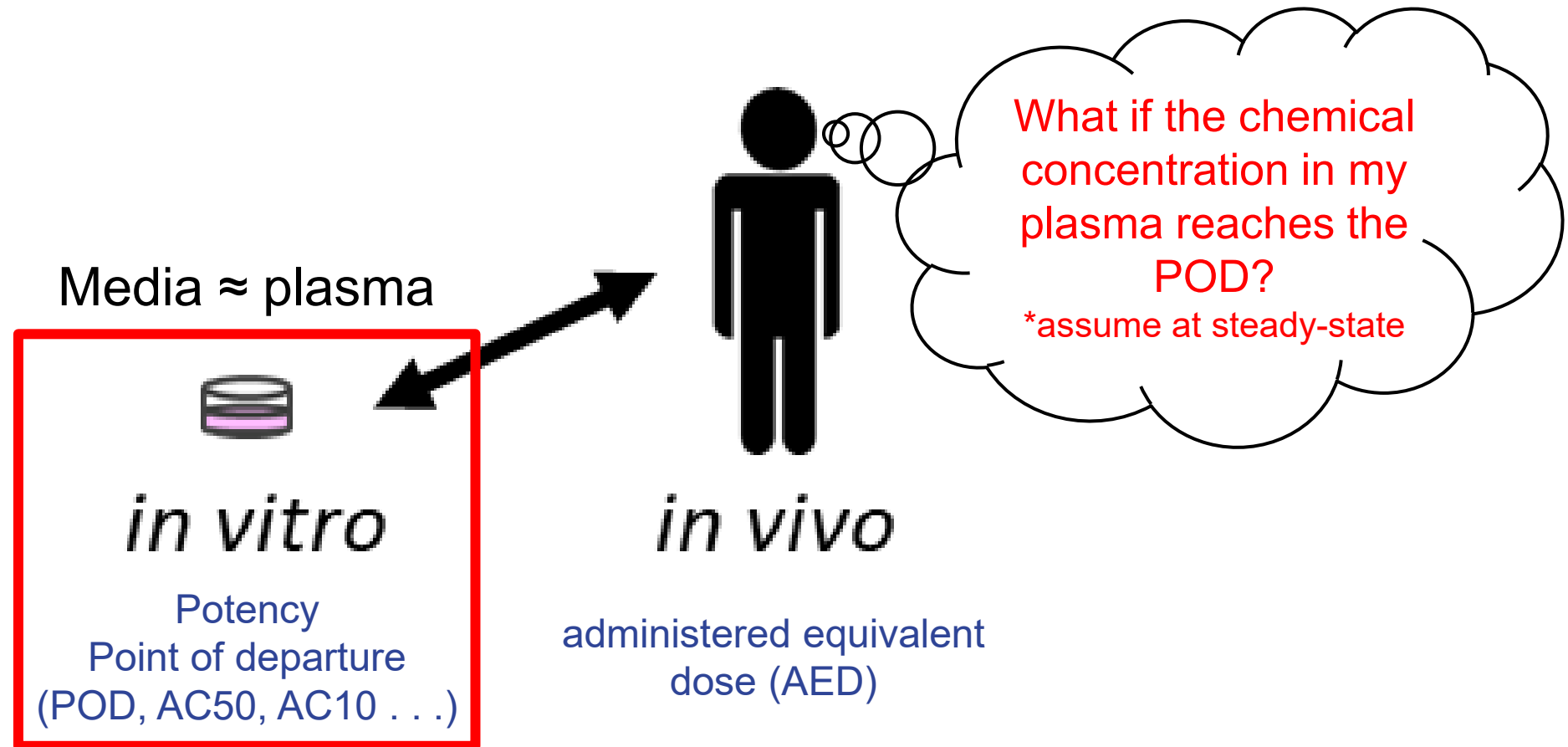
Webster F, et. al. (2019) Regul Toxicol Pharmacol ; OECD (2015) Test No. 455 <https://doi.org/10.1787/9789264243040-en>

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Making Sense of *In Vitro* Potencies



In Vitro-In Vivo Extrapolation (IVIVE)


aka reverse dosimetry, reverse toxicokinetics

Toxicokinetics + Toxicodynamics

Definition: utilization of *in vitro* experimental data to predict phenomena *in vivo*

Use of IVIVE tools to incorporate dosimetry has enabled a shift from a hazard-based to a risk-based interpretation of *in vitro* data

input

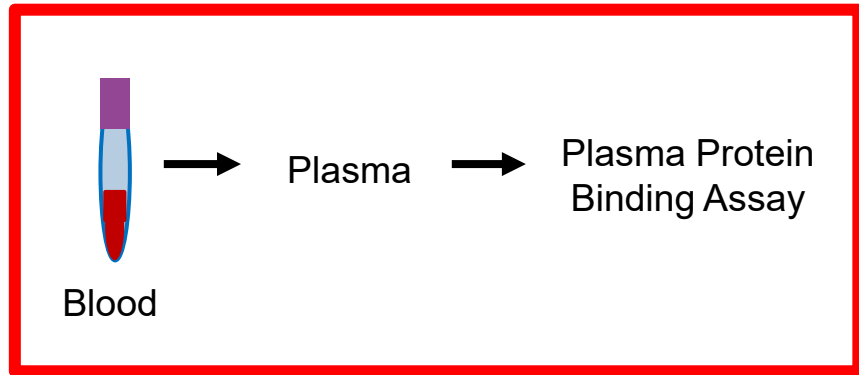

in vitro
potency

$$\approx C_{ss} = \frac{\text{oral dose rate}}{\underbrace{(GFR * F_{ub})}_{\text{kidney}} + \underbrace{\left(Q_l * F_{ub} * \frac{Cl_{int}}{Q_l + F_{ub} * Cl_{int}} \right)}_{\text{liver}}}$$

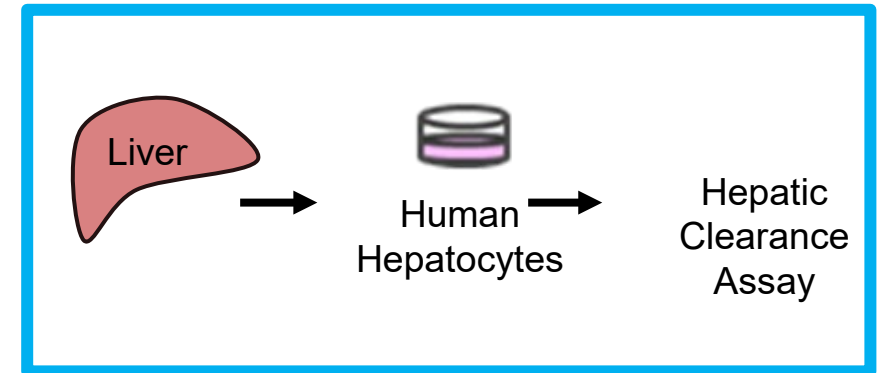
100% oral bioavailability assumed; kinetics are assumed to be linear

Estimating Clearance Using *In Vitro* Measurements of F_{ub} & Cl_{int}

Fraction of the compound unbound in plasma



Intrinsic metabolic clearance



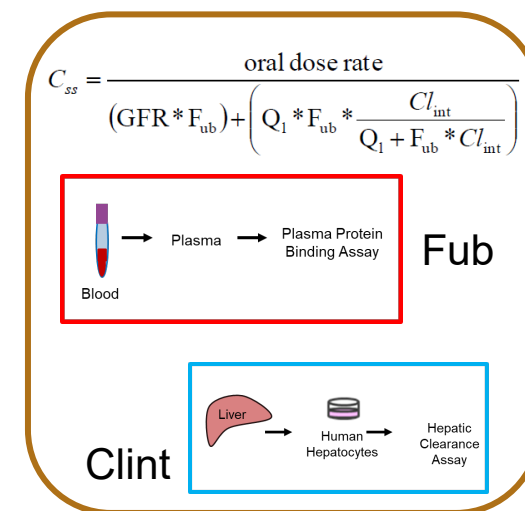
$$C_{ss} = \frac{\text{oral dose rate}}{\underbrace{(GFR * F_{ub})}_{\text{kidney}} + \underbrace{\left(Q_1 * F_{ub} * \frac{Cl_{int}}{Q_1 + F_{ub} * Cl_{int}} \right)}_{\text{liver}}}$$

Further efforts have focused on predicting F_{ub} & Cl_{int} using *in silico* approaches

High-Throughput Toxicokinetics (HTTK)

Meanings of “HTTK”

- Any component of evaluating toxicokinetics in a high-throughput manner
 - E.g., *in vitro* or *in silico* Clint and/or Fup
 - E.g., C_{ss}
- HTTK R-package
 - Generic pharmacologically based toxicokinetic (PBTk) model
 - Developed at the US EPA, free and publicly available
 - High throughput with appropriate input data
 - Forward dosimetry
 - Reverse dosimetry/reverse toxicokinetics (IVIVE)

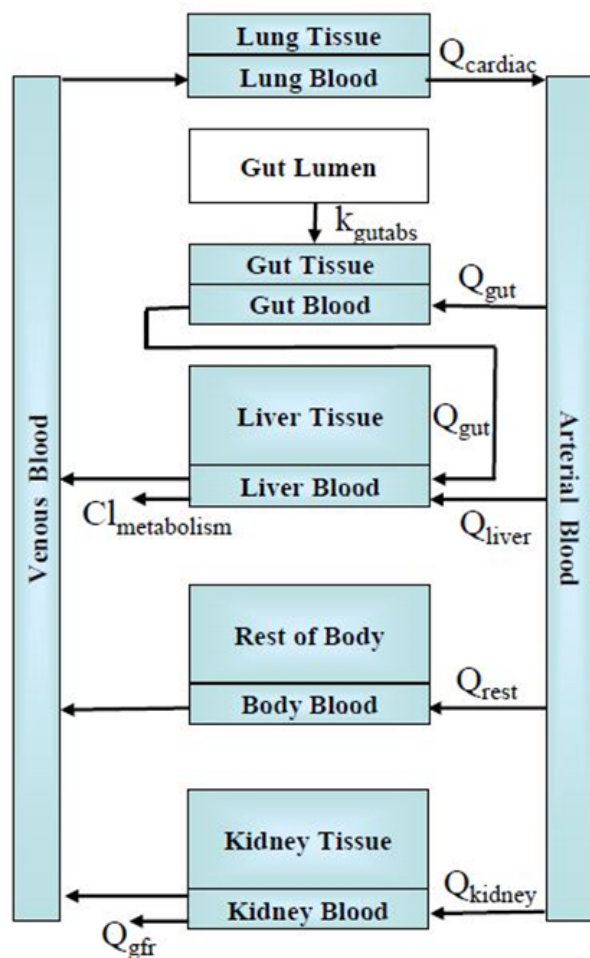


Commercial PBTk Software

- Simcyp: <https://www.certara.com/>
- ADMET Predictor / GastroPlus: <https://www.simulations-plus.com/>
- MEGen: <http://cefic-lri.org/toolbox/pbpkmegen/>
- IndusChemFate: <http://cefic-lri.org/toolbox/induschemfate/>

High-Throughput Toxicokinetics (HTTK) R-package

Generic PBTK model



- Simple models (1 and 3 compartments)
- Generic PBTK model
- Specialty: gas inhalation, aerosol inhalation, dermal, human gestational model

Body is represented by “compartments” and connected by “flows,” mass balance applies. Some compartments represent individual organs/tissues (e.g., liver); others are “lumped” (e.g., rest of body).

Parameterized using physicochemical properties (QSARs) + F_{up} & Cl_{int} data

- >1,000 and >8,000 chemicals with *in vitro* or *in silico* estimated parameter data, respectively

Various species (e.g., rat, rabbit, dog, human, monkey)

Assumptions

- Fast absorption rate (1/h)
- 100% bioavailability
- Chemical exits via metabolism or excretion by glomerular filtration

HTTK-Pop: Population Simulator for HTTK R-package

Not every adult is the same. Not every person is an “average” adult (e.g., children, elderly). *In vitro* and *in silico* tools to predict TK variability.

Similar approach to many commercial software

Sample NHANES quantities

Sex
Race/ethnicity
Age
Height
Weight
Serum creatinine



Regression equations
from literature
(+ residual marginal
variability)

Predict physiological
quantities

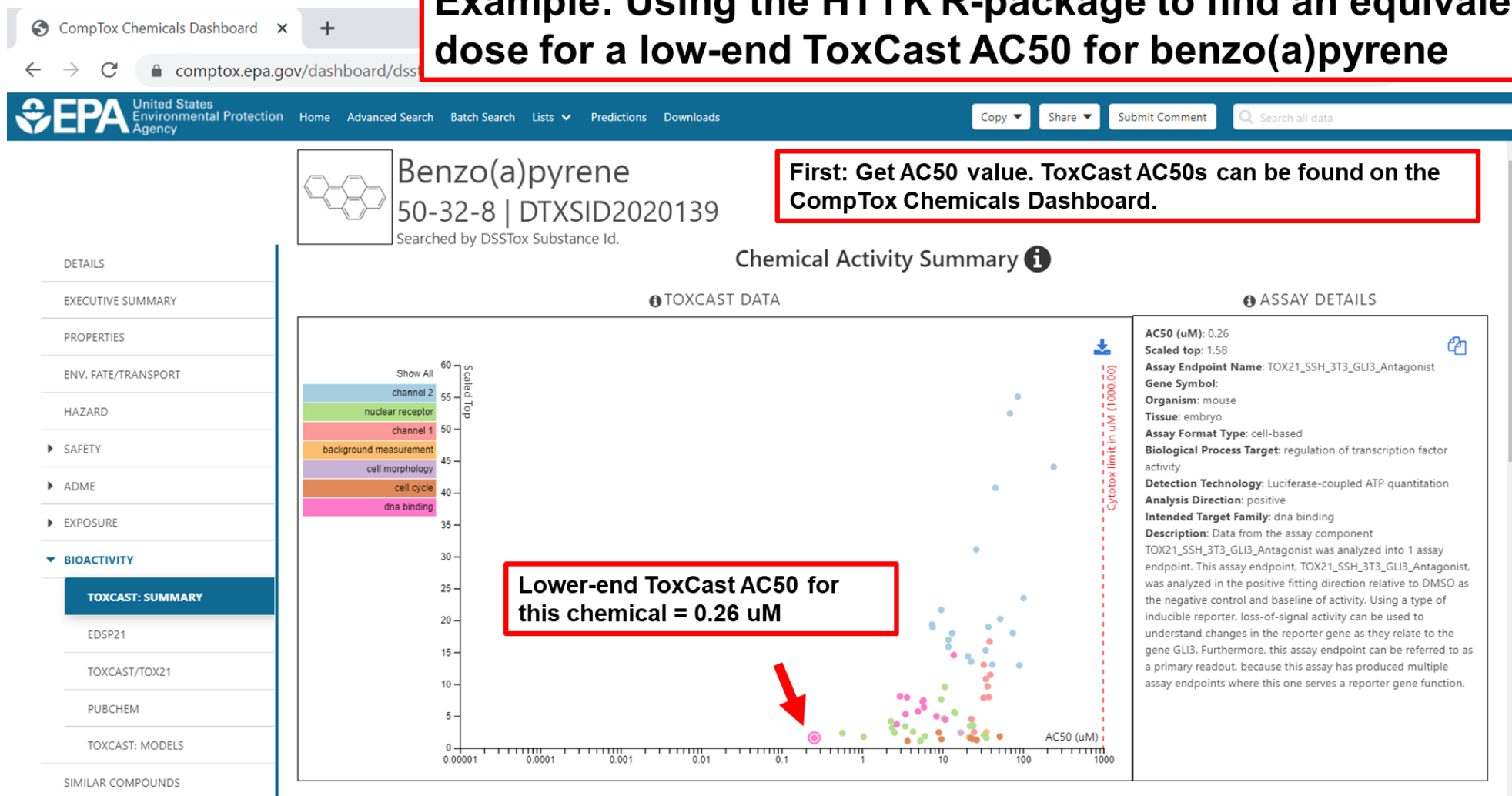
Tissue masses
Tissue blood flows
GFR (kidney function)
Hepatocellularity

Potentially eliminate the need for some uncertainty factors—human heterogeneity in vulnerability to exposures

Adapted from <https://ntp.niehs.nih.gov/iccvam/meetings/ivive-wksp-2016/wksp-ppts/1-2-ring-508.pdf>

How Can I Calculate Equivalent Dose from In Vitro Data?

Example: Using the HTTK R-package to find an equivalent dose for a low-end ToxCast AC50 for benzo(a)pyrene



Calculating equivalent dose is straight-forward

Use HHTK R-package Function `calc_mc_oral_equiv()`

```
> #Steady-state equivalent dose (mg/kg BW/day) to produce 0.26 uM in plasma:
```

```
> library(httk)
```

```
> set.seed(42)
```

```
> calc_mc_oral_equiv(conc=0.26,  
                     chem.name="benzo(a)pyrene",  
                     which.quantile = c(0.95, 0.5, 0.05),  
                     input.units = "uM",  
                     output.units = "mgpkgpday")
```

input

```
uM concentration converted to mgpkgpday dose for 0.95 0.5 0.05 quantile.
```

```
      95%      50%      5%
```

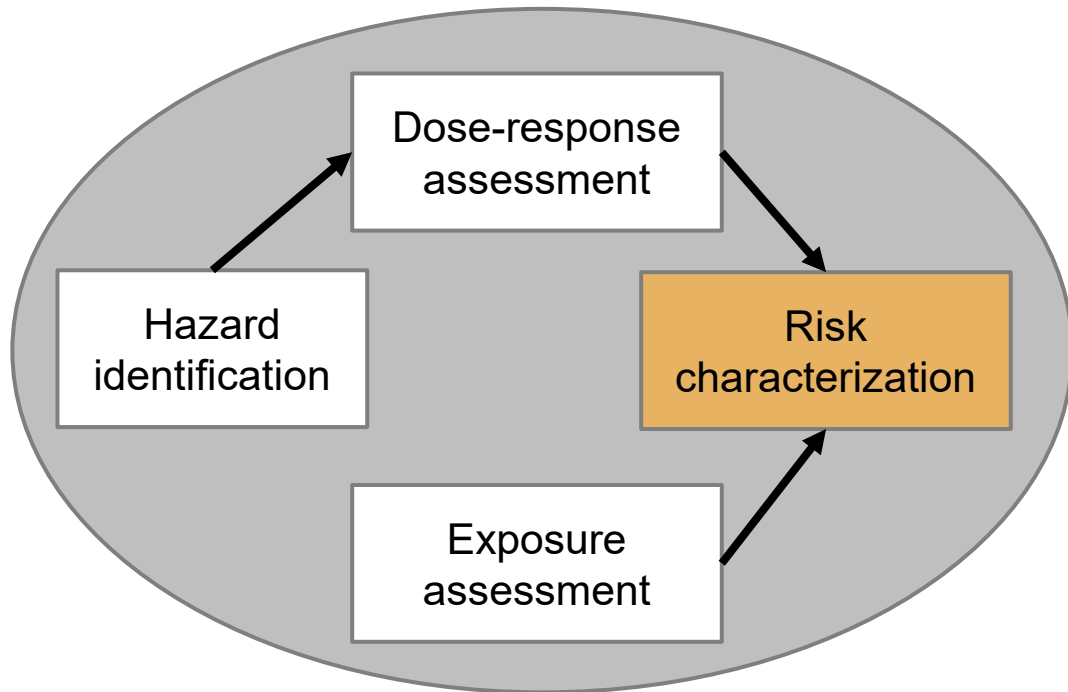
```
0.003821 0.019090 0.067080
```

output

In Vitro to *In Vivo* Extrapolation in Practice

- Provide context for *in vitro* data with respect to *in vivo* interaction likelihood
 - Conservative estimate (e.g., 100% assumed bioavailable)
 - Species differences
 - Population variability
 - Identifying sensitive population
 - Replace use of default safety factors in risk assessment
- Challenges
 - Chemical training sets with PK data
 - Phase II and III metabolism (transporters, glucuronidation)
 - Tissue distribution (blood versus target tissue)
 - C_{ss} versus C_{max}

How Can We Use Alternative Approaches in Risk Characterization?



1. *In silico* read-across (data gap analysis)
2. Hazard assessment (ID and dose-response)
 - a. *In vitro* assays
 - b. Making sense of *in vitro* potencies using *in vitro* to *in vivo* extrapolation (IVIVE)
3. High throughput exposure assessment
4. Risk characterization

Rapid Exposure Predictions (ExpoCast)

<https://www.epa.gov/chemical-research/rapid-chemical-exposure-and-dose-research>

- High-throughput, rapid exposure predictions for thousands of chemicals
- Environmental chemical focused
- Multiple routes of exposure

We can try to predict the exposure by describing the process leading to exposure

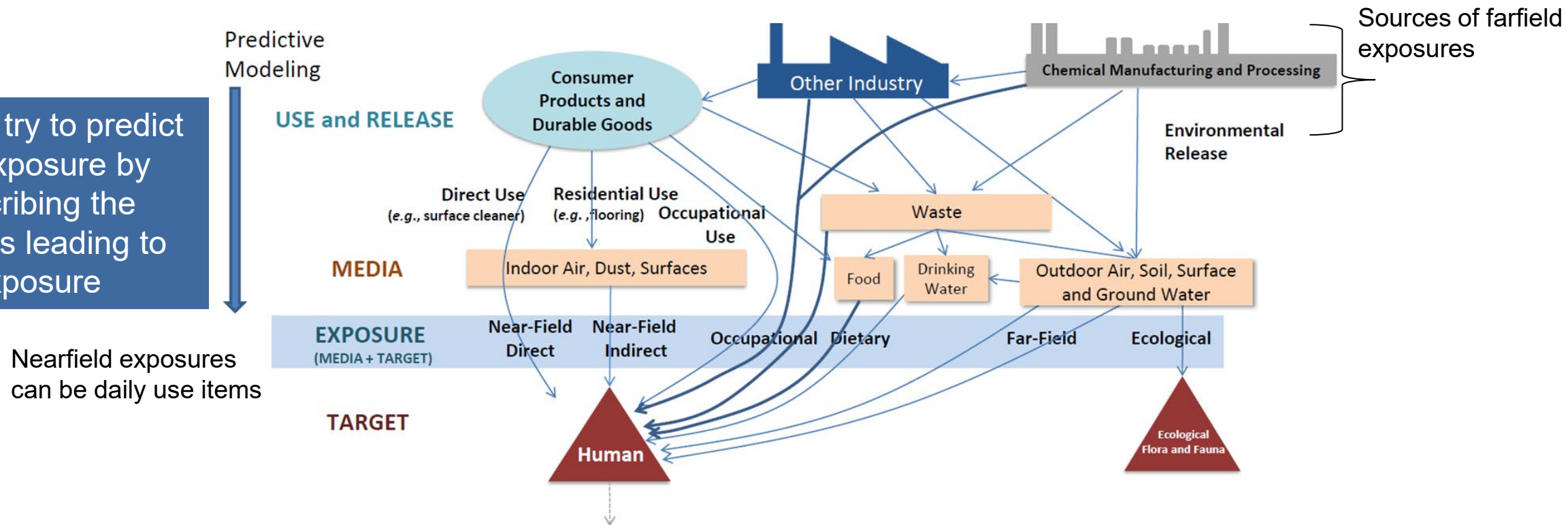


Image used with permission from John Wambaugh and Kristin Isaacs

Farfield Exposure Models

<https://www.epa.gov/chemical-research/rapid-chemical-exposure-and-dose-research>

- Predict exposure from chemicals that are released into the outdoor environment (air, water, soil) through industrial releases
- USETox
 - <https://usetox.org/>
 - Treat models like related assays and look for consensus while considering model appropriateness
 - Global scientific consensus fate, exposure, and effect model
- Risk Assessment IDentification and Ranking (RAIDAR) Model
 - <https://arnotresearch.com/raidar/>
 - Environmental fate and transport mass balance model linked with food web bioaccumulation models for representative ecological and agricultural targets
 - Applicable when little or no empirical data exist. Can “bin” chemicals into high or low risk potential.

Nearfield Exposure Models

<https://www.epa.gov/chemical-research/rapid-chemical-exposure-and-dose-research>

- Provide estimates of exposure (over various product types, scenarios, and routes) to chemicals used in consumer products and in-home products
- Stochastic Human Exposure and Dose Simulation Model (SHEDS)
 - Probabilistic models that can estimate everyday exposures
 - Detailed use patterns drive exposure
 - Multiple models: multimedia, dietary, residential, [high-throughput](#)
Isaacs KK, et al. Environ Sci Technol. 2014 Nov 4;48(21):12750-9.
- EPA Chemical and Products Database (CPDat)
 - >75,000 chemicals and 15,000 consumer products

Williams, P., B. Hubbell, E. Weber, C. Fehrenbacher, David Hrdý and V. Zartarian. "CHAPTER 3 An Overview of Exposure Assessment Models Used by the US Environmental Protection Agency." (2009).

Calibration and Evaluation of Models

<https://www.epa.gov/chemical-research/rapid-chemical-exposure-and-dose-research>

- Systematic Empirical Evaluation of Models (SEEM) framework
 - Calibration and evaluation of models toward consensus predictions
 - Compare with National Health and Nutrition Examination Study (NHANES)—blood and urine

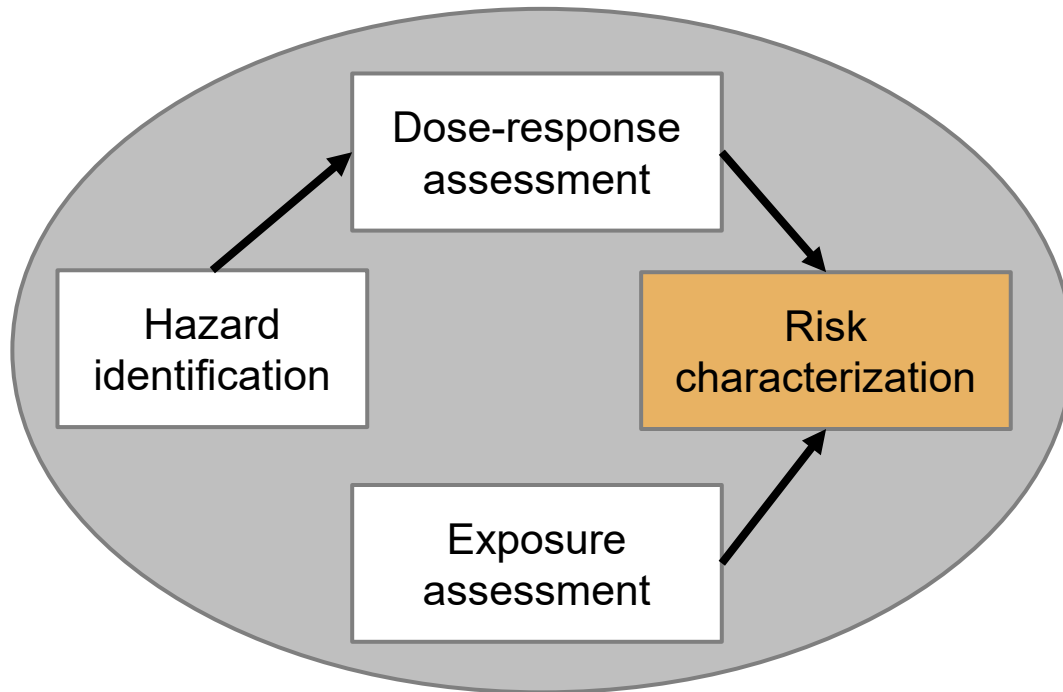
Wambaugh et al. “New Approach Methodologies for exposure science,” *Current Opinions in Toxicology*, 15, 76-92 (2019)

Non-targeted Chemical Screening

<https://www.epa.gov/chemical-research/rapid-chemical-exposure-and-dose-research>

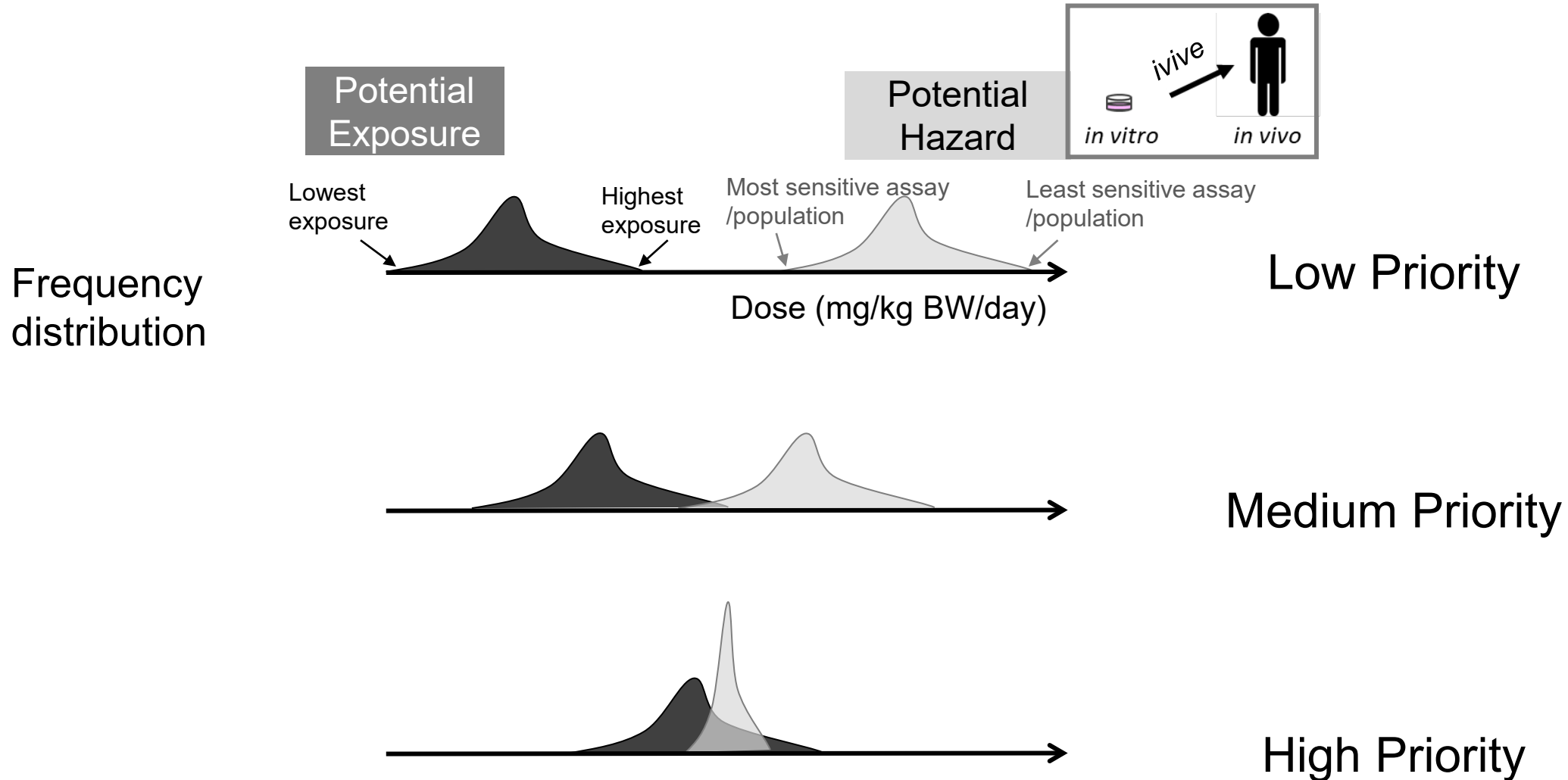
- Identify unknown chemicals in water, soil, and other types of samples, without having a preconceived idea of what chemicals are present
- Non-Targeted Analysis Collaborative Trial (ENTACT)
 - Evaluate ability of non-targeted methods to consistently and correctly identify unknown chemicals in a sample
 - 30 academic, government, and industry groups
- E.g., detection of GenX in the Cape Fear River, NC

How Can We Use Alternative Approaches in Risk Characterization?



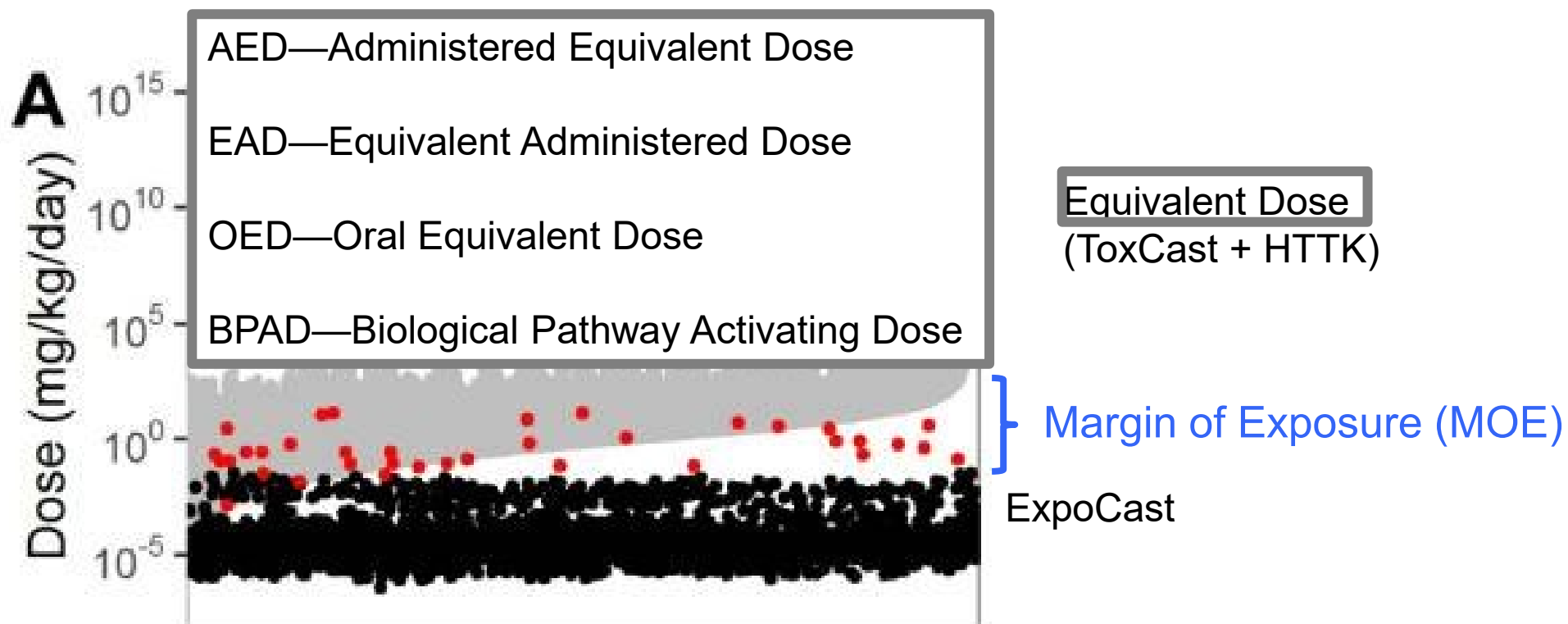
1. *In silico* read-across (data gap analysis)
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3. High throughput exposure assessment
4. Risk characterization

Bringing It All Together: High-Throughput Risk-Based Prioritization



Enabling Risk Based Prioritization

Margin of Exposure (MOE)

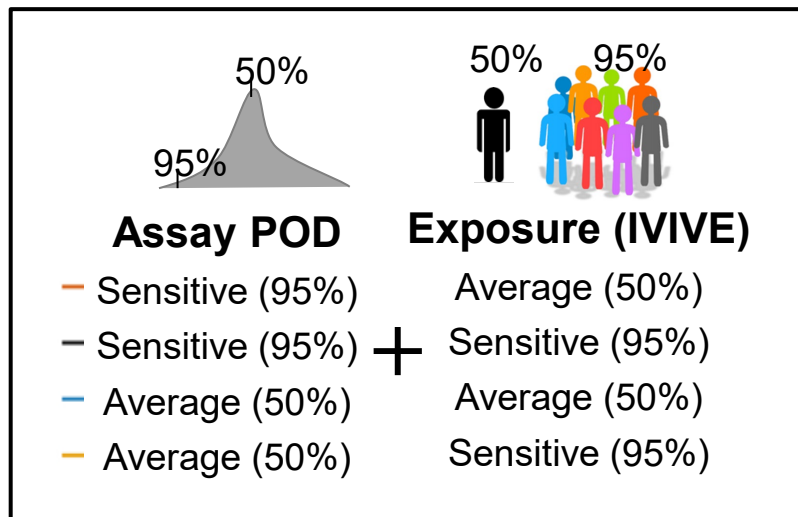


ToxCast chemicals (limited by availability of Fup & Clint data)

Enabling Risk Based Prioritization—Alternative View

BER: Bioactivity-Exposure Ratio (also known as AER: Activity-Exposure Ratio)

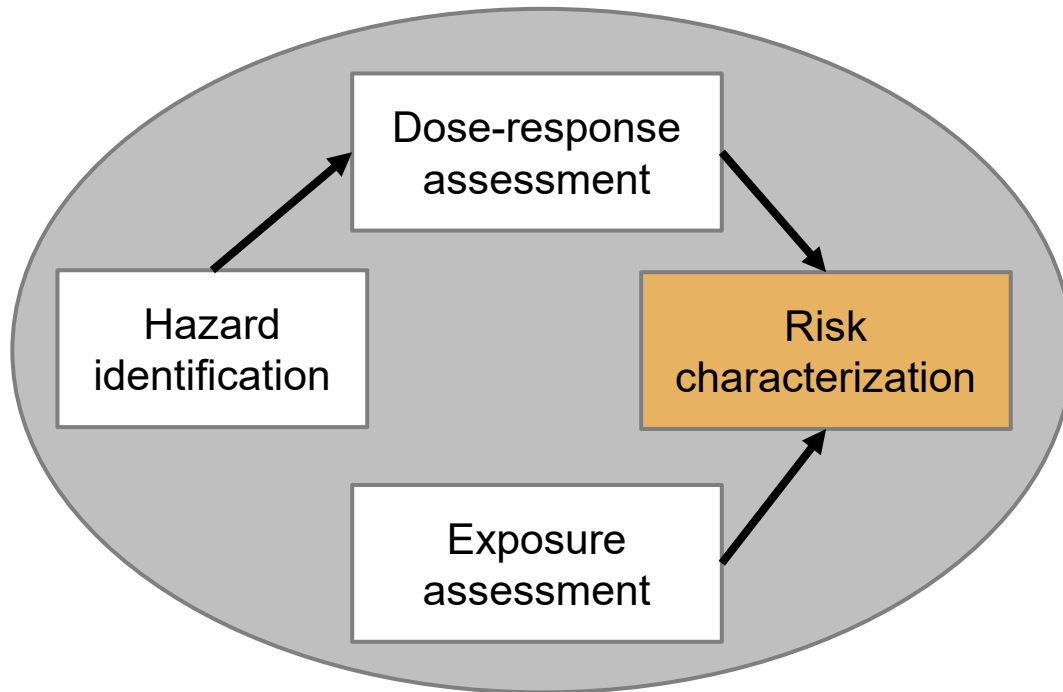
Administered Equivalent Dose (AED)



Make choices based on tolerable uncertainty (i.e., on use case)

Compare to Predicted Exposure (e.g., ExpoCast)

How Can We Use Alternative Approaches in Risk Characterization?



1. *In silico* read-across (data gap analysis)
2. Hazard assessment (ID and dose-response)
 - a. *In vitro* assays
 - b. Making sense of *in vitro* potencies using *in vitro* to *in vivo* extrapolation (IVIVE)
3. High throughput exposure assessment
4. Risk characterization

The basics were shown throughout this presentation

Covering All the Components of a 21st-Century Risk Assessment

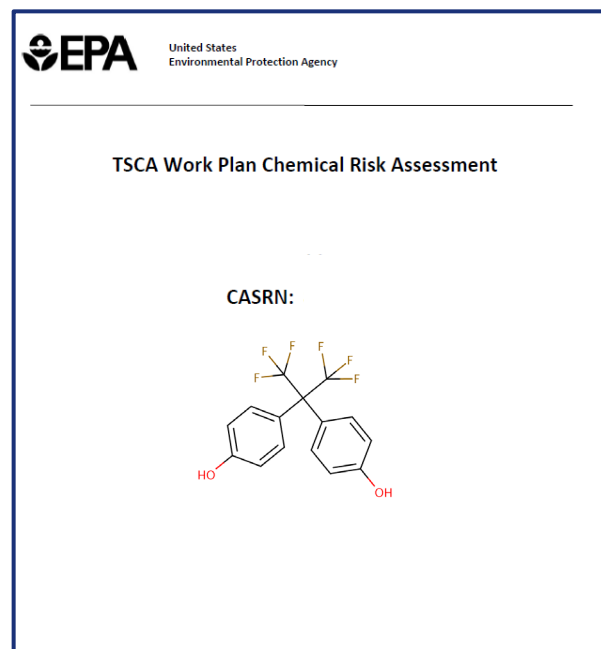


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Phys Chem

Exposure

Hazard

Dose Response, PK,
and PODs

Variability

Risk Summary

Uncertainty

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Closing

- Incorporating new technologies and innovations through computational toxicology can more rapidly and inexpensively screen chemicals for potential adverse biological effects.
- The field has made great advances in the development of NAMs and tools for filling information gaps for decision-making and integrating those tools and data streams into chemical risk assessment.
- International collaborations are leveraging resources and developing NAMs that can support different regulatory contexts.
- Building confidence in the use of NAMs for regulatory decision-making is key to the increased implementation of these methods.



What Is Needed to Expand Translation and Implementation of Computational Toxicology Approaches?

- Understand the need
- Integration of NAM data with traditional data
- Fit-for-purpose applications
- Transparency
- Stakeholder engagement, communication, and education
- Build confidence and understanding
 - Outreach and training
 - Hand's on use

EPA CompTox Chemicals Dashboard

Publicly available chemistry, toxicity, and exposure information for over 875,000 chemicals

The screenshot shows the EPA CompTox Chemicals Dashboard for Bisphenol A (DTXSID7020182). The interface includes a sidebar with navigation options like DETAILS, EXECUTIVE SUMMARY, PROPERTIES, ENV. FATE/TRANSPORT, HAZARD, SAFETY, ADME, EXPOSURE, BIOACTIVITY, SIMILAR COMPOUNDS, GENRA (BETA), RELATED SUBSTANCES, SYNONYMS, LITERATURE, LINKS, and COMMENTS. The main content area displays the chemical structure of Bisphenol A, its molecular formula (C₁₅H₁₆O₂), average mass (228.291 g/mol), and monoisotopic mass (228.11503 g/mol). It also includes sections for Wikipedia, Quality Control Notes, Intrinsic Properties, Structural Identifiers, Linked Substances, Presence in Lists, and Record Information. A text box at the bottom states: "Incorporates an extensive number of databases, tools, and publications".

Chemical characterization

- Physico-chemical properties (*in vitro* and/or *in silico*)
- Lists
- *In silico* read-across

Hazard—dose and effect

- *In vivo* animal legacy data
- *In vitro* assays
- Public literature
- Ecotoxicology (separate tools not on the dashboard)
 - Environmental toxicity data on aquatic life, terrestrial plants, and wildlife (ECOTOX knowledgebase)
 - Sequence alignment to predict across species susceptibility (SeqAPASS)

Toxicokinetics

- High-throughput *in vitro* and *in silico* parameters and model outputs from *in vitro* to *in vivo* extrapolation (IVIVE)

Exposure

- Exposure predictions, biomonitoring, production volume, use categories

Thank You!

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References

- Toxic Substances Control Act (TSCA): <https://www.epa.gov/assessing-and-managing-chemicals-under-tsca>
- National Academies of Sciences, Engineering, and Medicine 2017. Using 21st Century Science to Improve Risk-Related Evaluations. Washington, DC: The National Academies Press. <https://doi.org/10.17226/24635>.
- Kavlock RJ, et al. Computational Toxicology – A State of the Science Mini Review. Toxicol Sci. 2008;103(1):14-27.
- Kleinstreuer NC, Tetko IV, Tong W. Introduction to Special Issue: Computational Toxicology. Chem Res Toxicol. 2021;34(2):171-175.
- EPA Chemical Dashboard: <https://www.epa.gov/chemical-research/comptox-chemicals-dashboard>
- Read-across
 - Patlewicz G, et al. Navigating through the minefield of read-across tools: A review of in silico tools for grouping. Comput Toxicol. 2017;3:1-18. doi:10.1016/j.comtox.2017.05.003
 - Shah I, Liu J, Judson RS, Thomas RS, Patlewicz G. Systematically evaluating read-across prediction and performance using a local validity approach characterized by chemical structure and bioactivity information. Regul Toxicol Pharmacol. 2016 Aug;79:12-24. doi: 10.1016/j.yrtph.2016.05.008.
 - Rovida, C, et. al., NAM-supported read-across: From case studies to regulatory guidance in safety assessment, ALTEX - Alternatives to animal experimentation. 2021 38(1), pp. 140-150. doi: 10.14573/altex.2010062.
 - Chesnut, M., et al., (2018) “Regulatory acceptance of read-across”, ALTEX - Alternatives to animal experimentation, 35(3), pp. 413-419. doi: 10.14573/altex.1805081

References

- Filer DL, tcpl: the ToxCast pipeline for high-throughput screening data, *Bioinformatics*, 33(4), 15 Feb 2017, p618–620, <https://doi.org/10.1093/bioinformatics/btw680>
- Tox21 federal collaboration: <https://tox21.gov/>
- US EPA ToxCast Program: <https://www.epa.gov/chemical-research/toxicity-forecasting>
- Tox21 and ToxCast
 - Attene-Ramos MS, et al. The Tox21 robotic platform for the assessment of environmental chemicals--from vision to reality. *Drug Discov Today*. 2013 Aug;18(15-16):716-23. doi: 10.1016/j.drudis.2013.05.015. Epub 2013 May 31. PMID: 23732176; PMCID: PMC3771082.
 - Collins FS, Gray GM, Bucher JR. Transforming Environmental Health Protection. *Science*. 2008; 319(5865):906-907
 - Judson RS, Houck KA, Kavlock RJ, et al. In Vitro Screening of Environmental Chemicals for Targeted Testing Prioritization: The ToxCast Project. *Environ Health Perspect*. 2010;118(4):485-92
 - Kavlock R, Chandler KC, Houck K, et al. Update on EPA's ToxCast Program: Providing High Throughput Decision Support Tools for Chemical Risk Management. *Chem Res Toxicol*. 2012; 25(7):1287-302
 - Tice RR, Austin CP, Kavlock RJ, Bucher JR. Improving the Human Hazard Characterization of Chemicals: a Tox21 update. *Environ Health Perspect*. 2013;121(7):756-65
 - Thomas RS, Bahadori T, Buckley TJ, et al. The Next Generation Blueprint of Computational Toxicology at the U.S. Environmental Protection Agency. *Toxicol Sci*. 2019; 169(2): 317-332
 - Thomas RS, Paules RS, Simeonov A, et al. The US Federal Tox21 Program: A Strategic and Operational Plan for Continued Leadership. *ALTEX*. 2018; 35(2): 163-168

References

- High-Throughput Transcriptomics (HTTr)

- Harrill JA, et al. High-Throughput Transcriptomics Platform for Screening Environmental Chemicals. Toxicol Sci. 2021 Feb 4:kfab009. doi: 10.1093/toxsci/kfab009. Epub ahead of print.
- Harrill J, et al. Considerations for Strategic Use of High-Throughput Transcriptomics Chemical Screening Data in Regulatory Decisions. Curr Opin Toxicol. 2019;15:64-75. doi: 10.1016/j.cotox.2019.05.004.
- Phillips JR, et al. BMDExpress 2: Enhanced Transcriptomic Dose-Response Analysis Workflow. Bioinformatics. 2019; 35(10):1780-1782
- Ramaiahgari SC, et al. The Power of Resolution: Contextualized Understanding of Biological Responses to Liver Injury Chemicals Using High-throughput Transcriptomics and Benchmark Concentration Modeling. Toxicol Sci. 2019; 169(2): 553-566
- Yang L, Allen BC, Thomas RS. BMDExpress: a software tool for the benchmark dose analyses of genomic data. BMC Genomics. 2007; Oct25; 8:387
- Yeakley JM, et al.,. A trichostatin A expression signature identified by TempO-Seq targeted whole transcriptome profiling. PLoS One. 2017 May 25;12(5):e0178302. doi: 10.1371/journal.pone.0178302. PMID: 28542535; PMCID: PMC5444820.

- High-Throughput Phenotypic Profiling (HTPP)

- Bray MA, et al. Cell Painting, a high-content image-based assay for morphological profiling using multiplexed fluorescent dyes. Nat Protoc. 2016;11(9):1757-1774. doi:10.1038/nprot.2016.105
- Gustafsdottir SM, et al. Multiplex cytological profiling assay to measure diverse cellular states. PLoS One. 2013 Dec 2;8(12):e80999. doi: 10.1371/journal.pone.0080999.
- Nyffeler J, et al. Bioactivity screening of environmental chemicals using imaging-based high-throughput phenotypic profiling. Toxicol Appl Pharmacol. 2020 Jan 15;389:114876. doi: 10.1016/j.taap.2019.114876. Epub 2019 Dec 30.
- Willis C, Nyffeler J, Harrill J. Phenotypic Profiling of Reference Chemicals across Biologically Diverse Cell Types Using the Cell Painting Assay. SLAS Discov. 2020 Aug;25(7):755-769. doi: 10.1177/2472555220928004. Epub 2020 Jun 17.

References

- Organotypic Models and Microphysiological Systems (MPS)

- Deisenroth C, et al. Development of an In Vitro Human Thyroid Microtissue Model for Chemical Screening. Toxicol Sci. 2020 Mar 1;174(1):63-78. doi: 10.1093/toxsci/kfz238.
- LeCluyse EL, Witek RP, Andersen ME, Powers MJ. Organotypic liver culture models: meeting current challenges in toxicity testing. Crit Rev Toxicol. 2012;42(6):501-548. doi:10.3109/10408444.2012.682115
- Shen JX, et al. Organotypic and Microphysiological Models of Liver, Gut, and Kidney for Studies of Drug Metabolism, Pharmacokinetics, and Toxicity. Chemical Research in Toxicology 2020 33 (1), 38-60. DOI: 10.1021/acs.chemrestox.9b00245
- <https://www.epa.gov/research-grants/star-horizons-and-challenges-organotypic-culture-models-predictive-toxicology>
- <https://www.nist.gov/programs-projects/multi-organ-microphysiological-systems>

- Small Model Organisms

- Garcia GR, Noyes PD, Tanguay RL. Advancements in zebrafish applications for 21st century toxicology. Pharmacol Ther. 2016 May;161:11-21. doi: 10.1016/j.pharmthera.2016.03.009. Epub 2016 Mar 22. PMID: 27016469; PMCID: PMC4851906.
- Padilla S, Corum D, Padnos B, Hunter DL, Beam A, Houck KA, Sipes N, Kleinstreuer N, Knudsen T, Dix DJ, Reif DM. Zebrafish developmental screening of the ToxCast™ Phase I chemical library. Reprod Toxicol. 2012 Apr;33(2):174-87. doi: 10.1016/j.reprotox.2011.10.018. Epub 2011 Dec 9.
- Boyd WA, Smith MV, Freedman JH. Caenorhabditis elegans as a model in developmental toxicology. Methods Mol Biol. 2012;889:15-24. doi: 10.1007/978-1-61779-867-2_3.
- <https://invivobiosystems.com/disease-modeling/worms-flies-fish-comparison-common-model-organisms/>

References

- https://echa.europa.eu/documents/10162/21650280/oecd_test_guidelines_skin_sensitisation_en.pdf
- <https://ntp.niehs.nih.gov/whatwestudy/niceatm/accept-methods/index.html>
- Thomas RS, et al. The Next Generation Blueprint of Computational Toxicology at the U.S. Environmental Protection Agency. *Toxicol Sci.* 2019 Jun 1;169(2):317-332. doi: 10.1093/toxsci/kfz058.
- Aschner M, et al. Reference compounds for alternative test methods to indicate developmental neurotoxicity (DNT) potential of chemicals: example lists and criteria for their selection and use. *ALTEX.* 2017;34(1):49-74. doi: 10.14573/altex.1604201. Epub 2016 Jul 25.
- Sachana M, Shafer TJ, Terron A. Toward a Better Testing Paradigm for Developmental Neurotoxicity: OECD Efforts and Regulatory Considerations. *Biology (Basel).* 2021 Jan 23;10(2):86. doi: 10.3390/biology10020086.
- [http://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=ENV/JM/MONO\(2020\)25&docLanguage=en](http://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=ENV/JM/MONO(2020)25&docLanguage=en)
- <http://www.oecd.org/chemicalsafety/risk-assessment/iata-integrated-approaches-to-testing-and-assessment.htm>

References

- Estrogen Disrupting Chemical IATA references:
 - Browne P, Judson RS, Casey WM, et al. Screening Chemicals for Estrogen Receptor Bioactivity Using a Computational Model. *Environ Sci Technol*. 2015; 49(14):8804-14
 - Browne P, Noyes PD, Casey WM, Dix DJ. Application of Adverse Outcome Pathways to U.S. EPA's Endocrine Disruptor Screening Program. *Environ Health Perspect*. 2017 Sep 1;125(9):096001. doi: 10.1289/EHP1304.
 - Judson RS, Magpantay FM, Chickarmane V, et al. Integrated Model of Chemical Perturbations of a Biological Pathway Using 18 In Vitro High Throughput Screening Assays for the Estrogen Receptor. *Toxicol Sci*. 2015; 148(1):137-54
 - Kleinstreuer NC, Ceger PC, Allen DG, Strickland J, Chang X, Hamm JT, Casey WM. A Curated Database of Rodent Uterotrophic Bioactivity. *Environ Health Perspect*. 2016 May;124(5):556-62. doi: 10.1289/ehp.1510183. Epub 2015 Oct 2.
 - US Environmental Protection Agency. Use of High Throughput Assays and Computational Tools; Endocrine Disruptor Screening Program; Notice of Availability and Opportunity for Comment. National Archives Federal Register. 2015; EPA-HQ-OPPT-2015-0305, 80 FR 35350:35350-55
 - Watt ED, Judson RS. Uncertainty quantification in ToxCast high throughput screening. *PLoS One*. 2018 Jul 25;13(7):e0196963. doi: 10.1371/journal.pone.0196963.
 - Webster F, et. al. (2019) Regul Toxicol Pharmacol ; OECD (2015) Test No. 455
<https://doi.org/10.1787/9789264243040-en>
 - <https://www.epa.gov/endocrine-disruption>

References

- *In Vitro to In Vivo Extrapolation (IVIVE)*
 - Linakis MW, et al. "Development and Evaluation of a High Throughput Inhalation Model for Organic Chemicals". JESEE. <https://doi.org/10.1038/s41370-020-0238-y>
 - Pearce RG, Setzer RW, Strobe CL, et al. *httk: R Package for High-Throughput Toxicokinetics*. J Stat Softw. 2017;79(4):1-26
 - Ring, Caroline L., et al. "Identifying populations sensitive to environmental chemicals by simulating toxicokinetic variability." *Environment International* 106 (2017): 105-118
 - Rotroff, DM et al., 2010. Incorporating Human Dosimetry and Exposure into High-Throughput In Vitro Toxicity Screening. *Toxicol. Sci.*, 117 (2):348-358.
 - Sipes NS, Wambaugh JF, Pearce R, et al. An Intuitive Approach for Predicting Potential Human Health Risk with the Tox21 10k Library. *Environ Sci Technol*. 2017; 51(18):10786-10796
 - Tan YM, Liao KH, Clewell HJ 3rd. Reverse dosimetry: interpreting trihalomethanes biomonitoring data using physiologically based pharmacokinetic modeling. *J Expo Sci Environ Epidemiol*. 2007 Nov;17(7):591-603. doi: 10.1038/sj.jes.7500540. Epub 2006 Nov 15.
 - Tonnelier A, Coecke S, Zaldívar JM. Screening of chemicals for human bioaccumulative potential with a physiologically based toxicokinetic model. *Arch Toxicol*. 2012 Mar;86(3):393-403. doi: 10.1007/s00204-011-0768-0. Epub 2011 Nov 17. PMID: 22089525; PMCID: PMC3282909.
 - Wilkinson RG, and Shand DG (1975). A physiological approach to hepatic drug clearance. *Clin. Pharmacol. Ther.* 18, 377–389.

References

- *In Vitro to In Vivo Extrapolation (IVIVE), continued*
 - Wambaugh JF, et al. High throughput heuristics for prioritizing human exposure to environmental chemicals. *Environmental science & technology*. 2014;48(21):12760–7.
 - Wambaugh, JF et al., 2015. Toxicokinetic Triage for Environmental Chemicals. *Toxicol Sci.*, 147(1):55-67.
 - Wetmore, BA et al., 2012. Integration of Dosimetry, Exposure and High-Throughput Screening in Chemical Toxicity Assessment. *Toxicol. Sci.*, 125(1):157-174.
 - Wetmore, BA et al., 2013. Relative Impact of Incorporating Pharmacokinetics on Predicting In Vivo Hazard and Mode of Action from High-Throughput In Vitro Toxicity Assays. *Toxicol. Sci.*, 132(2):327-346.
 - Wetmore, BA et al., 2014. Incorporating Population Variability and Susceptible Subpopulations into Dosimetry for High-Throughput Toxicity Testing. *Toxicol. Sci.*, 142(1):210-214.
 - Wetmore, BA, et al. 2015. Quantitative in vitro-in vivo extrapolation in a high-throughput environment. *Toxicol.* 332:94-101.
 - Wetmore, BA et al., 2014. Incorporating Population Variability and Susceptible Subpopulations into Dosimetry for High-Throughput Toxicity Testing. *Toxicol. Sci.*, 142(1):210-214.

References

- **Rapid Exposure Predictions**
 - Arnot, J.A.; Mackay, D.; Webster, E.; Southwood, J. 2006. Screening level risk assessment model for chemical fate and effects in the environment. Environ. Sci. Technol. 40: 2316-2323. DOI: 10.1021/es0514085
 - Arnot, J.A.; Mackay D. 2008. Policies for chemical hazard and risk priority setting: Can persistence, bioaccumulation, toxicity and quantity information be combined? Environ. Sci. Technol. 42: 4648-4654. DOI: 10.1021/es800106g
 - Dionisio, K., et al. The Chemical and Products Database, a resource for exposure-relevant data on chemicals in consumer products. Sci Data 5, 180125 (2018). <https://doi.org/10.1038/sdata.2018.125>
 - Egeghy PP, et al. Computational Exposure Science: An Emerging Discipline to Support 21st-Century Risk Assessment. Environ Health Perspect. 2016 Jun;124(6):697-702. doi: 10.1289/ehp.1509748. Epub 2015 Nov 6.
 - Isaacs KK, et al. SHEDS-HT: an integrated probabilistic exposure model for prioritizing exposures to chemicals with near-field and dietary sources. Environ Sci Technol. 2014 Nov 4;48(21):12750-9. doi: 10.1021/es502513w. Epub 2014 Oct 21. PMID: 25222184.
 - Newton, S.R., Sobus, J.R., Ulrich, E.M. et al. Examining NTA performance and potential using fortified and reference house dust as part of EPA's Non-Targeted Analysis Collaborative Trial (ENTACT). Anal Bioanal Chem 412, 4221–4233 (2020).
 - Ring, Caroline L., et al. “Consensus Modeling of Median Chemical Intake for the US Population Based on Predictions of Exposure Pathways.” Environmental science & technology 53.2 (2018): 719-732
 - Rosenbaum, R.K., Bachmann, T.M., Gold, L.S. et al. USEtox—the UNEP-SETAC toxicity model: recommended characterisation factors for human toxicity and freshwater ecotoxicity in life cycle impact assessment. Int J Life Cycle Assess 13, 532 (2008). <https://doi.org/10.1007/s11367-008-0038-4>
 - Wambaugh et al. “New Approach Methodologies for exposure science”, Current Opinions in Toxicology, 15, 76-92 (2019)
 - Williams, P., et al. “CHAPTER 3 An Overview of Exposure Assessment Models Used by the US Environmental Protection Agency.” (2009).
 - <https://www.epa.gov/chemical-research/rapid-chemical-exposure-and-dose-research>

References

- **Rapid Risk Based Prioritization**

- Judson, RS et al., 2011. Estimating Toxicity-Related Biological Pathway Altering Doses for High-Throughput Chemical Risk Assessment. *Chem. Res. Toxicol.*, 24(4):451-62.
- Kavlock RJ, Bahaduri T, Barton-Maclaren TS, et al. Accelerating the Pace of Chemical Risk Assessment. *Chem Res Toxicol.* 2018; 31(5):287-290
- Paul Friedman K, et al. Utility of In Vitro Bioactivity as a Lower Bound Estimate of In Vivo Adverse Effect Levels and in Risk-Based Prioritization. *Toxicol Sci.* 2020 Jan 1;173(1):202-225. doi: 10.1093/toxsci/kfz201.
- Shin HM et al. Risk-Based High-Throughput Chemical Screening and Prioritization using Exposure Models and in Vitro Bioactivity Assays. *Environ Sci Technol.* 2015 Jun 2;49(11):6760-71.
- Sipes NS, Wambaugh JF, Pearce R, et al. An Intuitive Approach for Predicting Potential Human Health Risk with the Tox21 10k Library. *Environ Sci Technol.* 2017; 51(18):10786-10796
- Thomas, RS et al., 2013. Incorporating New Technologies into Toxicity Testing and Risk Assessment: Moving from a 21st Century Vision to a Data-Driven Framework. *Toxicol. Sci.*, 136(1):4-18.
- Wambaugh JF, et al. Assessing Toxicokinetic Uncertainty and Variability in Risk Prioritization. *Toxicol Sci.* 2019 Dec 1;172(2):235-251.