

# Findings from EPA's Non-Targeted Analysis Collaborative Trial (ENTACT)

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# Why Does EPA Need Measurement Data?

- **Measurement data needed to ensure chemical safety**

- Characterize risk
- Regulate use & disposal
- Manage human & ecological exposures
- Ensure compliance under federal statutes

## Toxic Substances Control Act (TSCA) Compliance Monitoring

To protect federal, state, and tribal health and the environment from unreasonable risks of chemicals, EPA monitors chemical substances under the Toxic Substances Control Act (TSCA).

## Safe Drinking Water Act (SDWA) Compliance Monitoring

Providing safe drinking water to the public is one of EPA's primary responsibilities. Under the Safe Drinking Water Act (SDWA), EPA monitors public water systems to ensure they are providing safe drinking water.

## Federal Insecticide, Fungicide and Rodenticide Act Compliance Monitoring

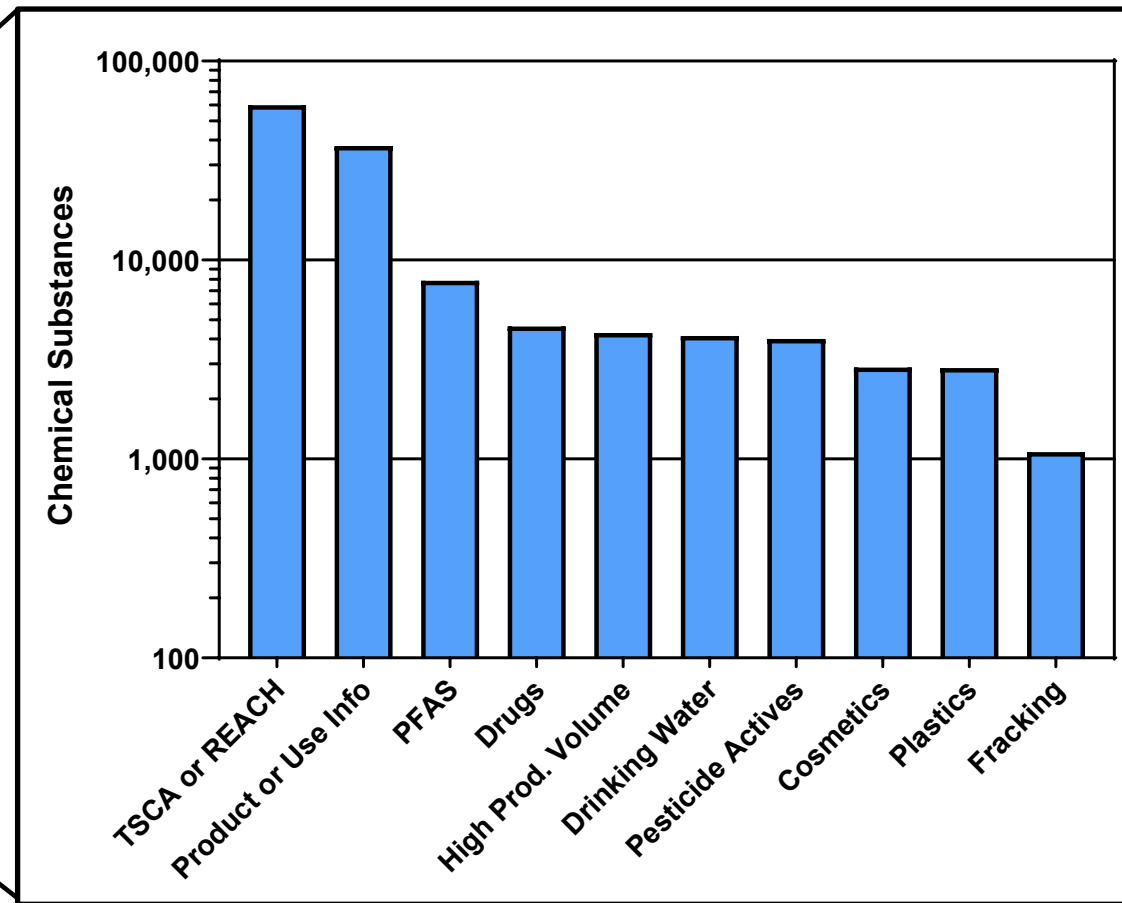
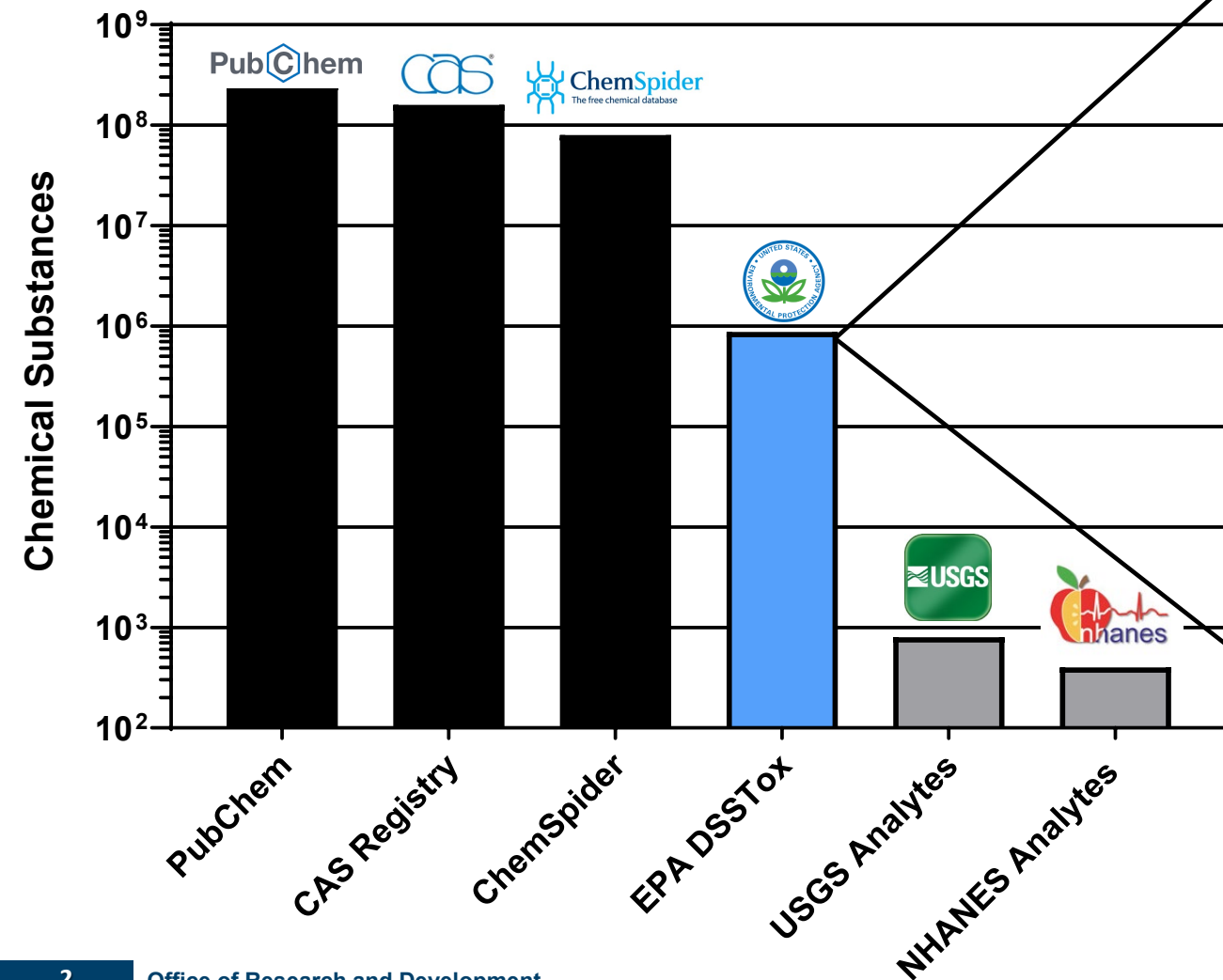
The Federal Insecticide, Fungicide and Rodenticide Act (FIFRA) gives EPA the authority to regulate the registration, distribution, sale and use of pesticides. FIFRA applies to all types of pesticides, including:

Resources and  
Guidance  
Documents

## Chemical Monitoring Needs



# Data Disparity: Have vs. Need



# Challenges

- High-quality exposure data are unavailable for most chemicals
- Measurement data traditionally generated using “targeted” methods
- Targeted analytical methods:
  - Require *a priori* knowledge of chemicals of interest
  - Produce data for few selected analytes (10s-100s)
  - Require standards for method development & compound quantitation
  - Are blind to emerging contaminants
  - Can't keep pace with the needs of 21<sup>st</sup> century chemical safety evaluations

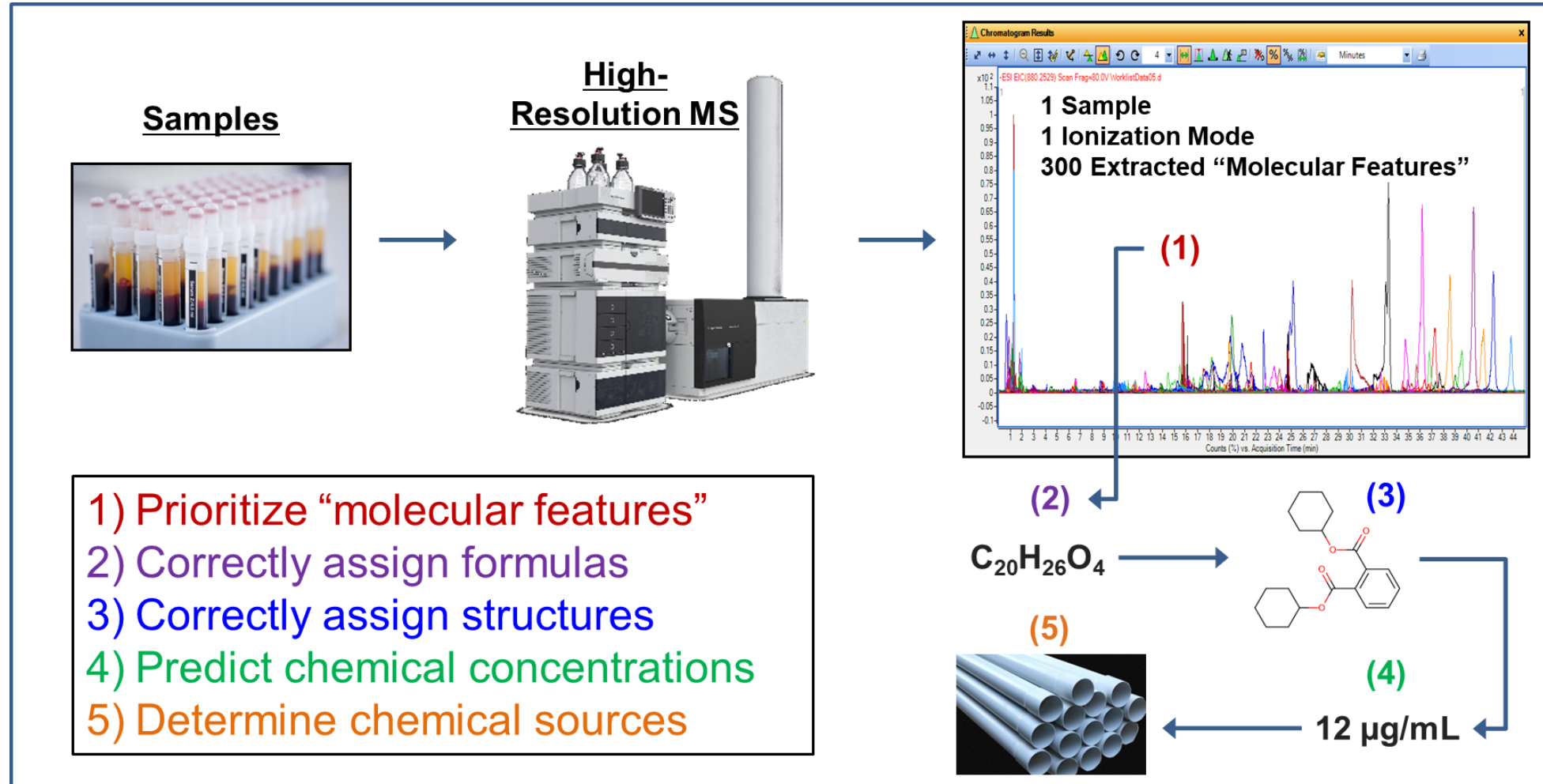
# What's So Great About NTA?

Rapidly screen  
for “knowns”

Discover  
“unknowns”

Uncover historical  
exposures

Generate source  
fingerprints...



# NTA State-of-the-Science

## Environmental Science & Technology

Viewpoint

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### Is Nontargeted Screening Reproducible?

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journal homepage: [www.elsevier.com/locate/scitotenv](http://www.elsevier.com/locate/scitotenv)



Prioritizing potential endocrine active high resolution mass spectrometry (HRMS) features in Minnesota lakewater

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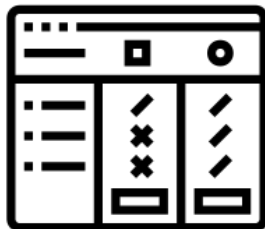
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*“No single analytical technique is suitable for the analysis of all compounds, and successful nontargeted screening will require the development of multiplatform approaches, facilitated and validated through interlaboratory collaborations.”*

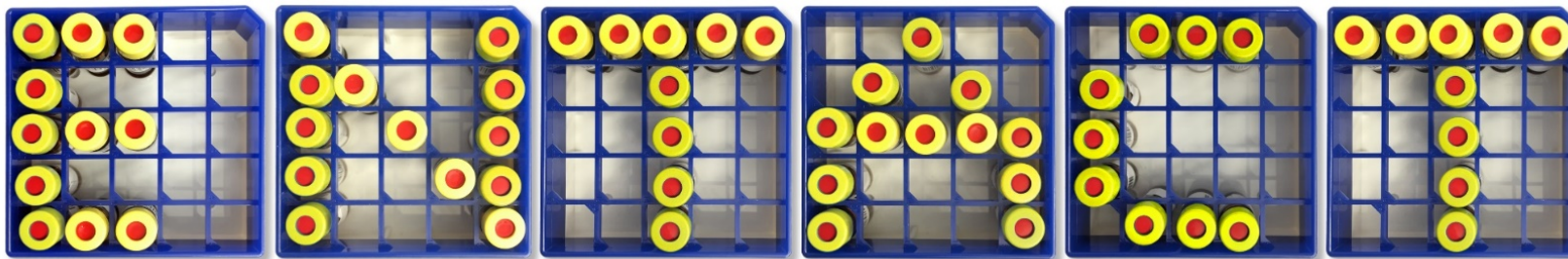
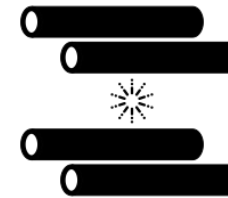
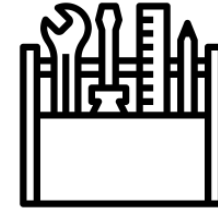


*“The novelty of nontarget analysis, particularly its current lack of implementation by regulatory agencies, has prevented the establishment of streamlined quality assurance and quality control (QA/QC) procedures.”*



# Science Questions for Research Community

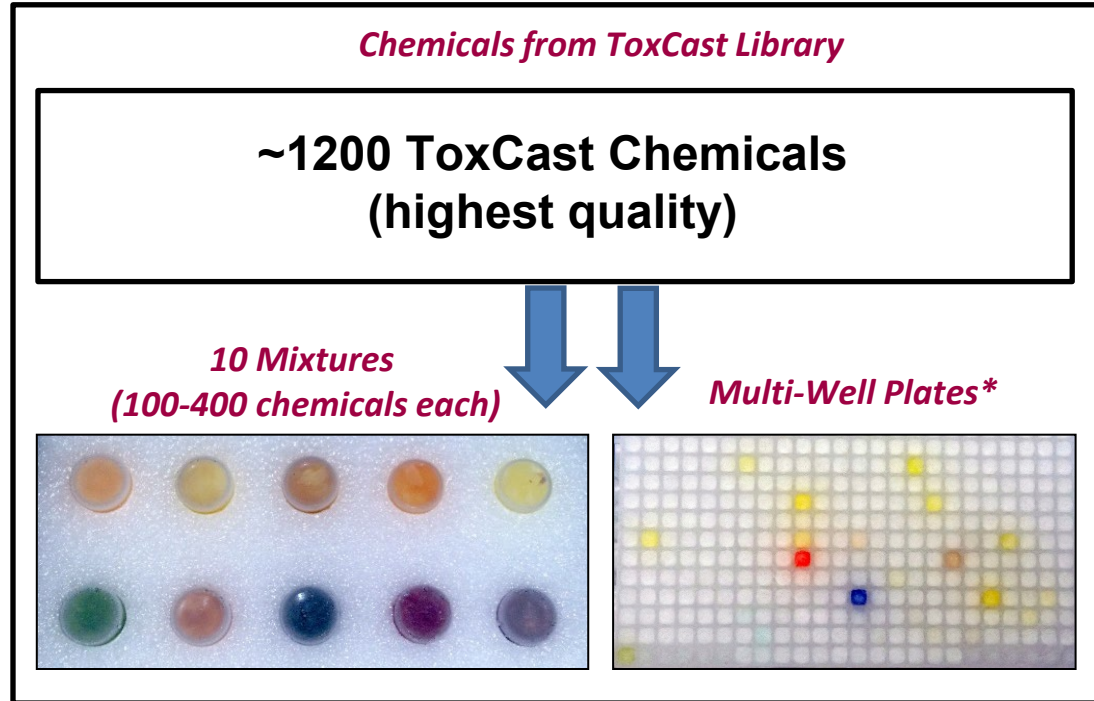
- How variable are tools and results from lab to lab?
- Are some methods/workflows better than others?
- How does sample complexity affect performance?
- What chemical space does a given method cover?
- How sensitive are specific instruments/methods?



*EPA's Non-Targeted Analysis Collaborative Trial*



## ENTACT Part 1



~25 Collaborators & 6 Contractors\*:

1<sup>st</sup>: Blinded analysis

2<sup>nd</sup>: Unveiling of chemicals

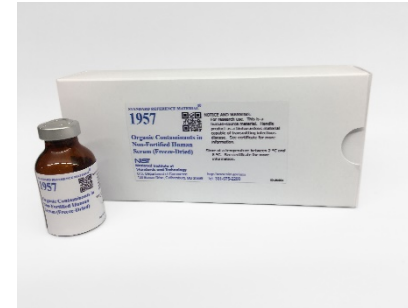
3<sup>rd</sup>: Unblinded evaluation

## ENTACT Part 2

*Reference & Fortified House Dust*



*Reference & Fortified Human Serum*

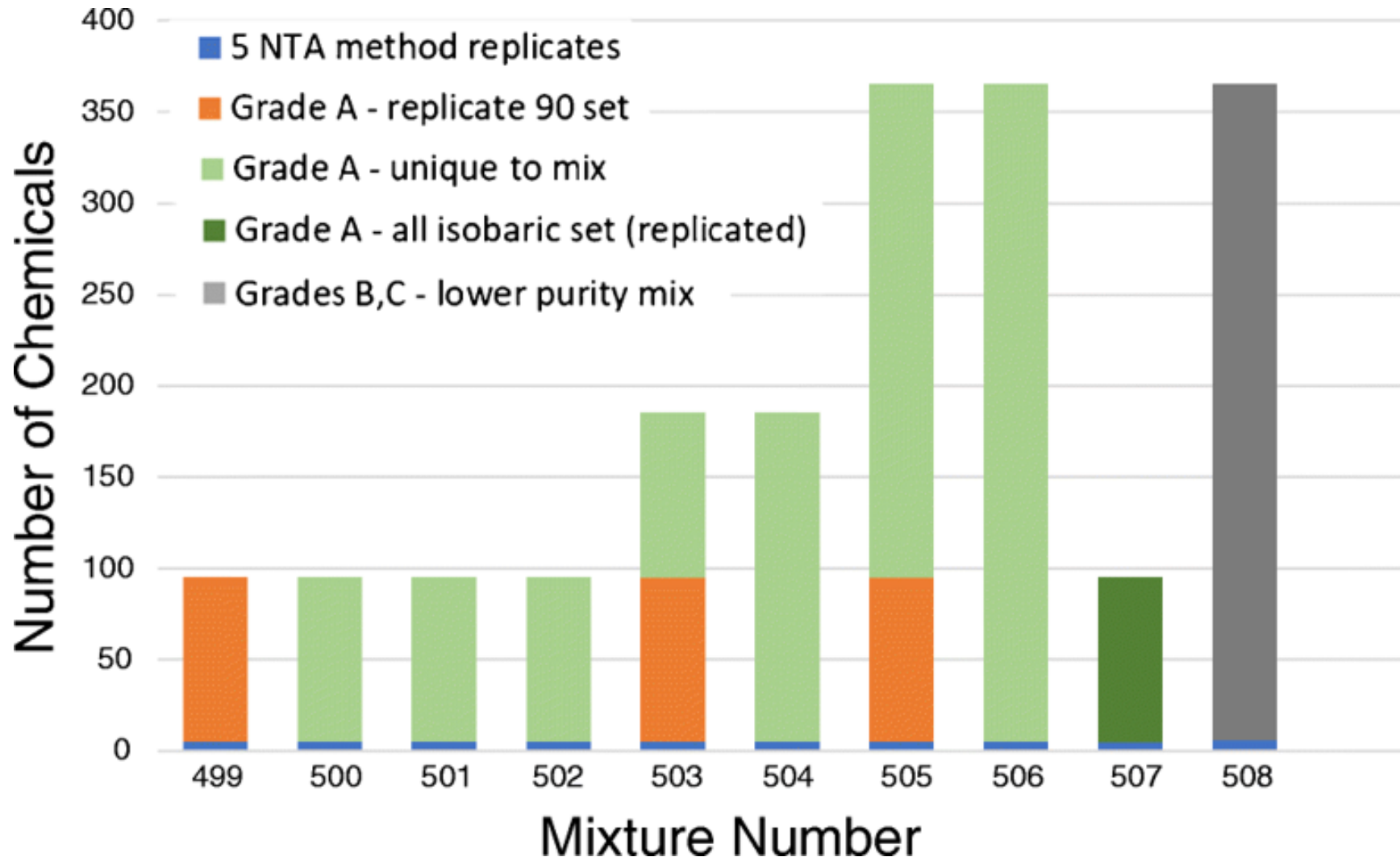


*Reference & Fortified Silicone Wristbands*





# Design of ENTACT Mixtures

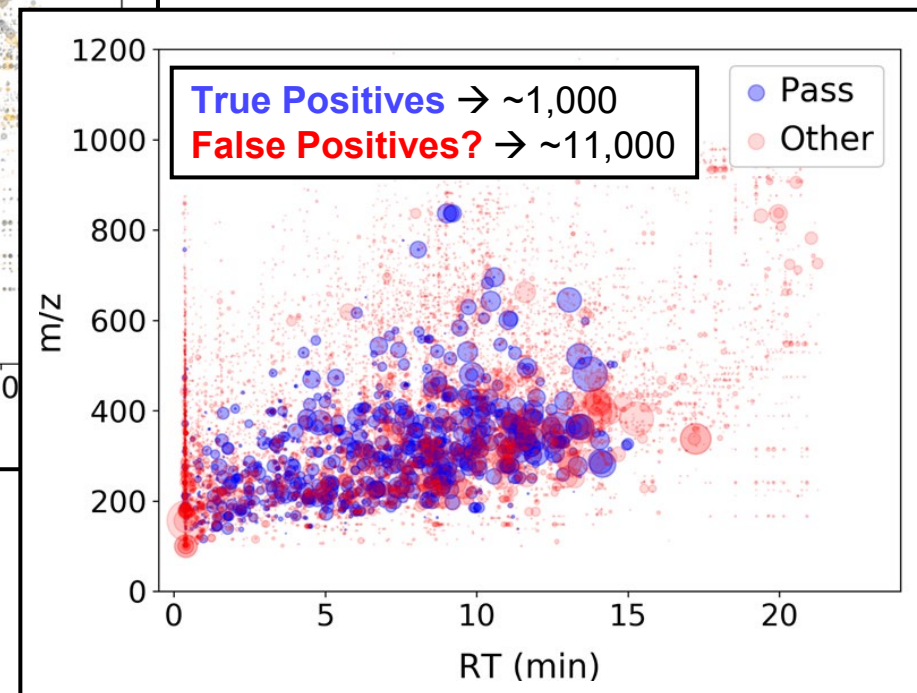
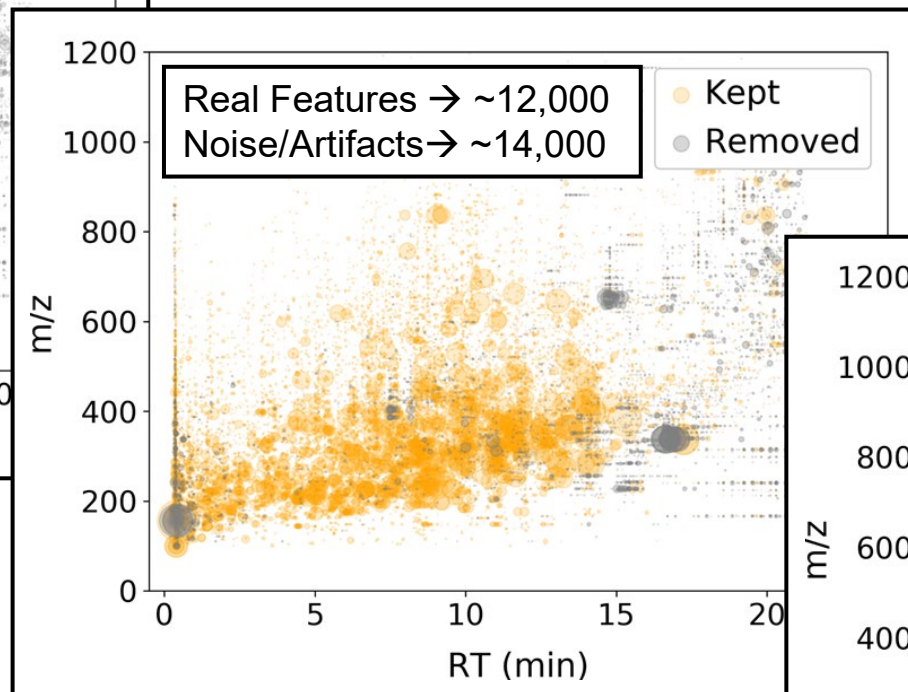
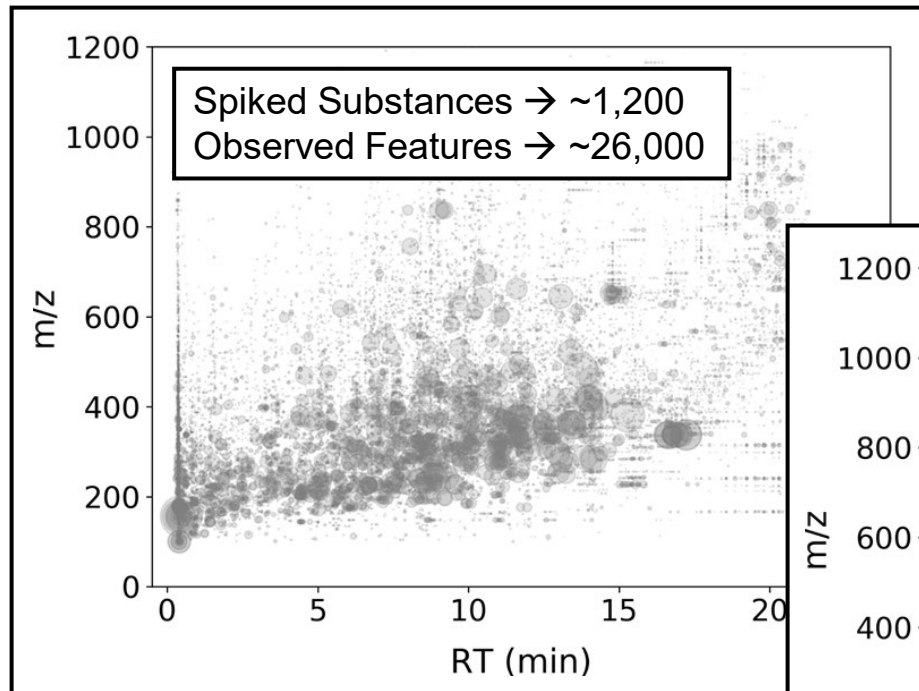


**Replication in  
substance spikes  
offers a unique  
means to assess  
NTA method  
reproducibility!**

# EPA Lab Results for ENTACT Mixtures



**LC-QTOF HRMS  
(ESI+ and ESI-)**



**Substance Spiked?**

**Yes**

**No**

**Substance Identified?**

**Yes**

**True Positives  
(≤ 65%)**

**False  
Positives?**

**No**

**False Negatives  
(≥ 35%)**

**True  
Negatives?**

# Who Else is Working on ENTACT?

## Contractors:



**19 Blind  
submissions**

**15 Unblinded  
submissions**

## Vendors:



## General Participants:



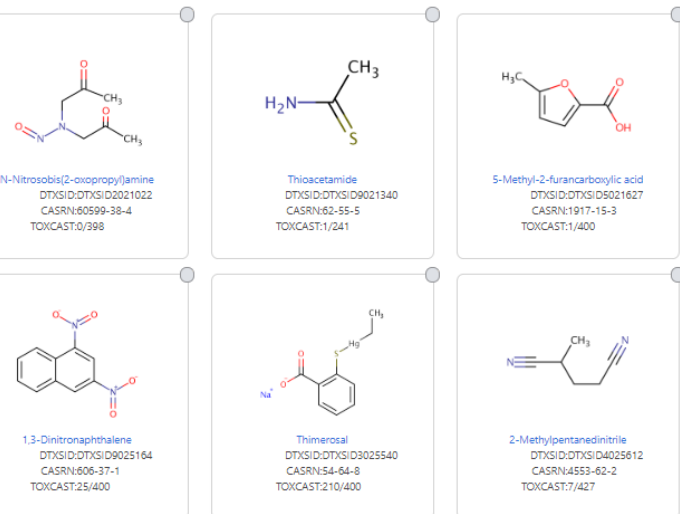
# Processing ENTACT Data Submissions

- Individual methods treated separately (if appropriate)
- One candidate mass/formula/compound per feature
- Confidence level revised as needed (with consensus)
- Matching to spiked substances by mass, formula & structure
- “**Observed**” if structure or formula (no spiked isomers) match
- “**Identified**” if structure match
- “**Reproducible**” if correctly ID’d >50% of the time
  - For compounds spiked >1 time and identified  $\geq 1$  time

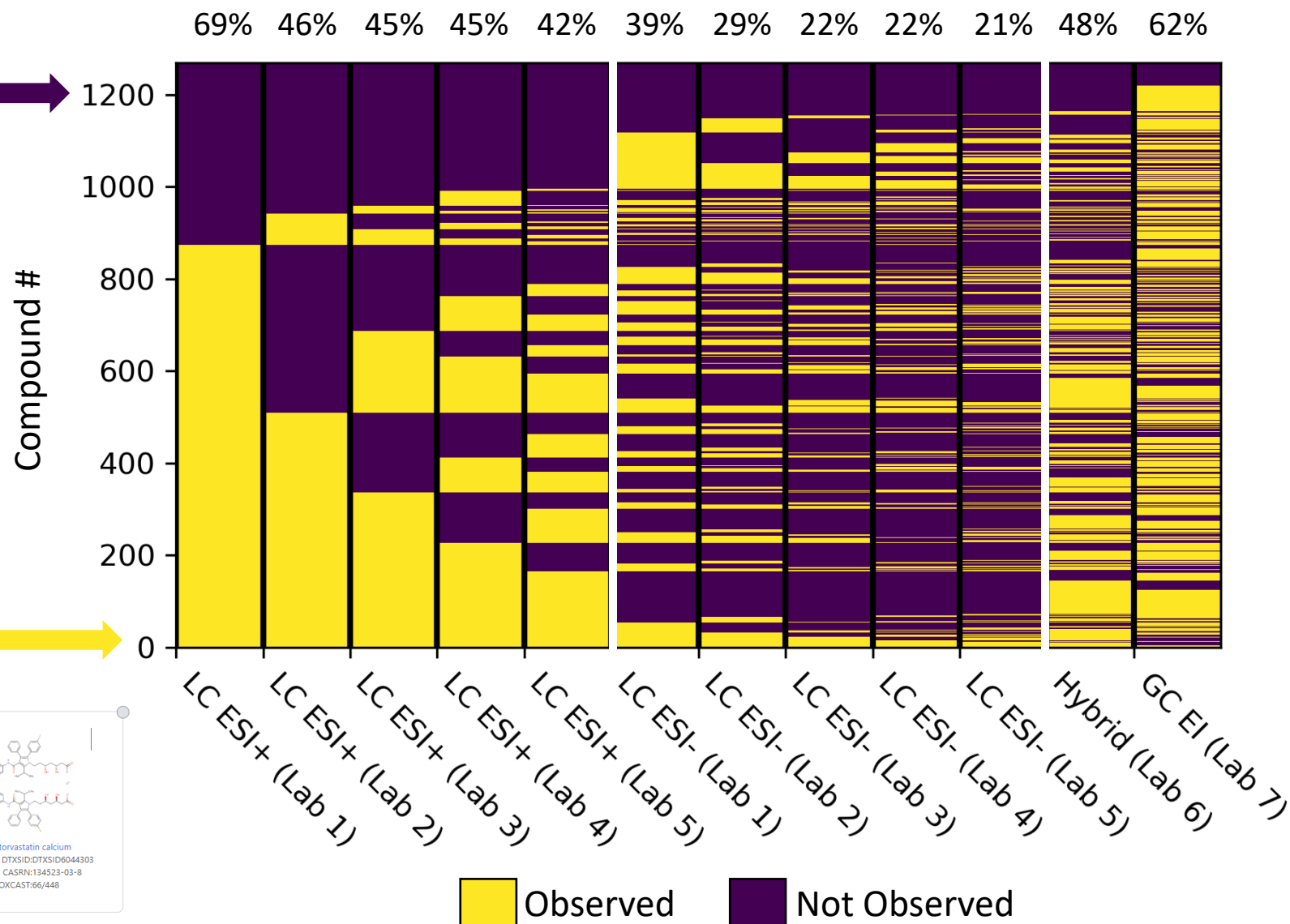
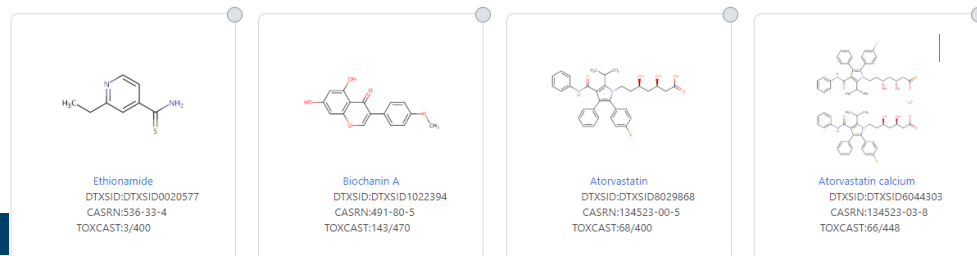
# Method Comparison: “Observed” Compounds

7 Labs, 12 Methods

~5% Not Observed by Any Method

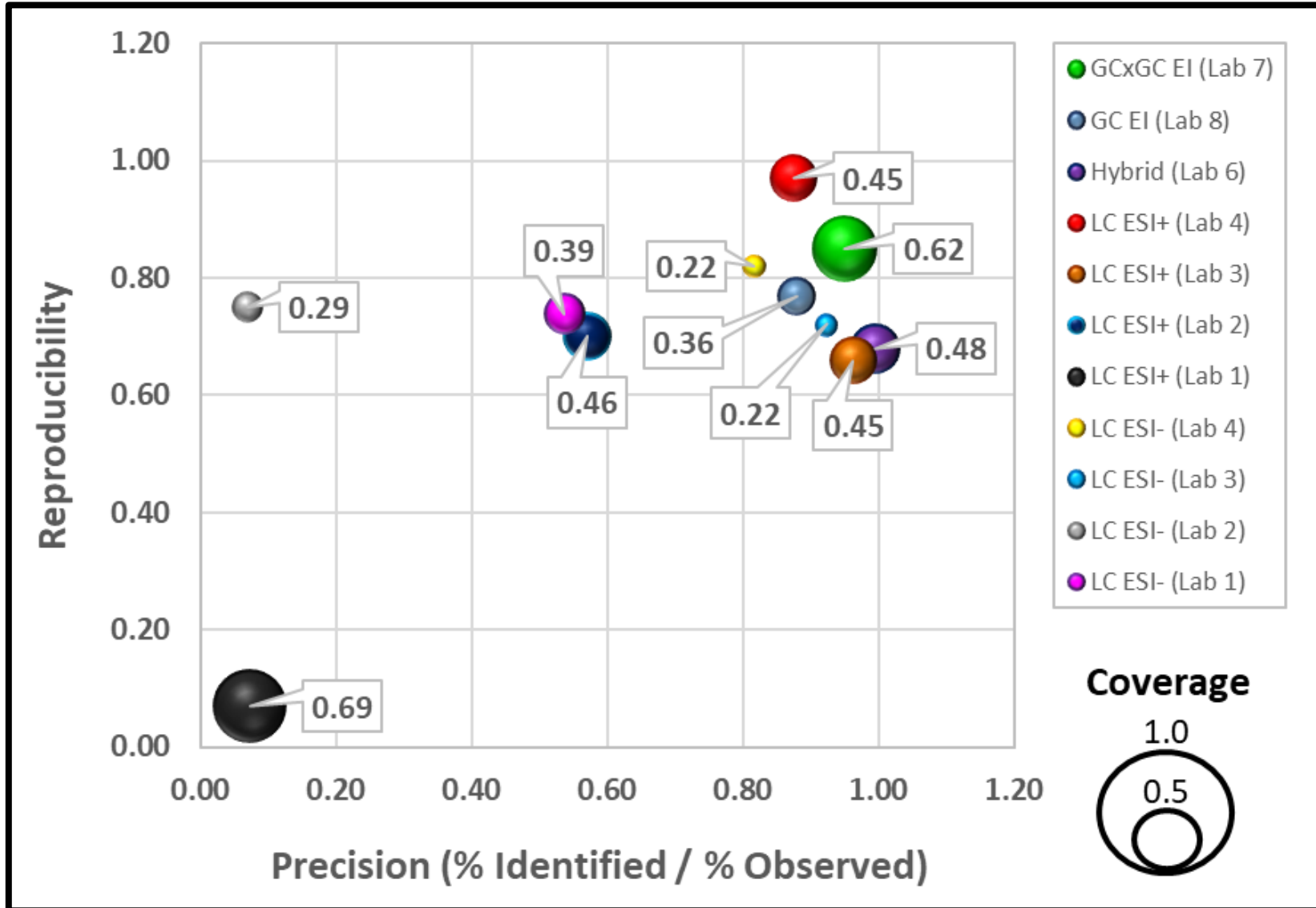


<1% Observed by All 12 Methods





# Method Comparison: Total Performance



**Metrics (all %):**

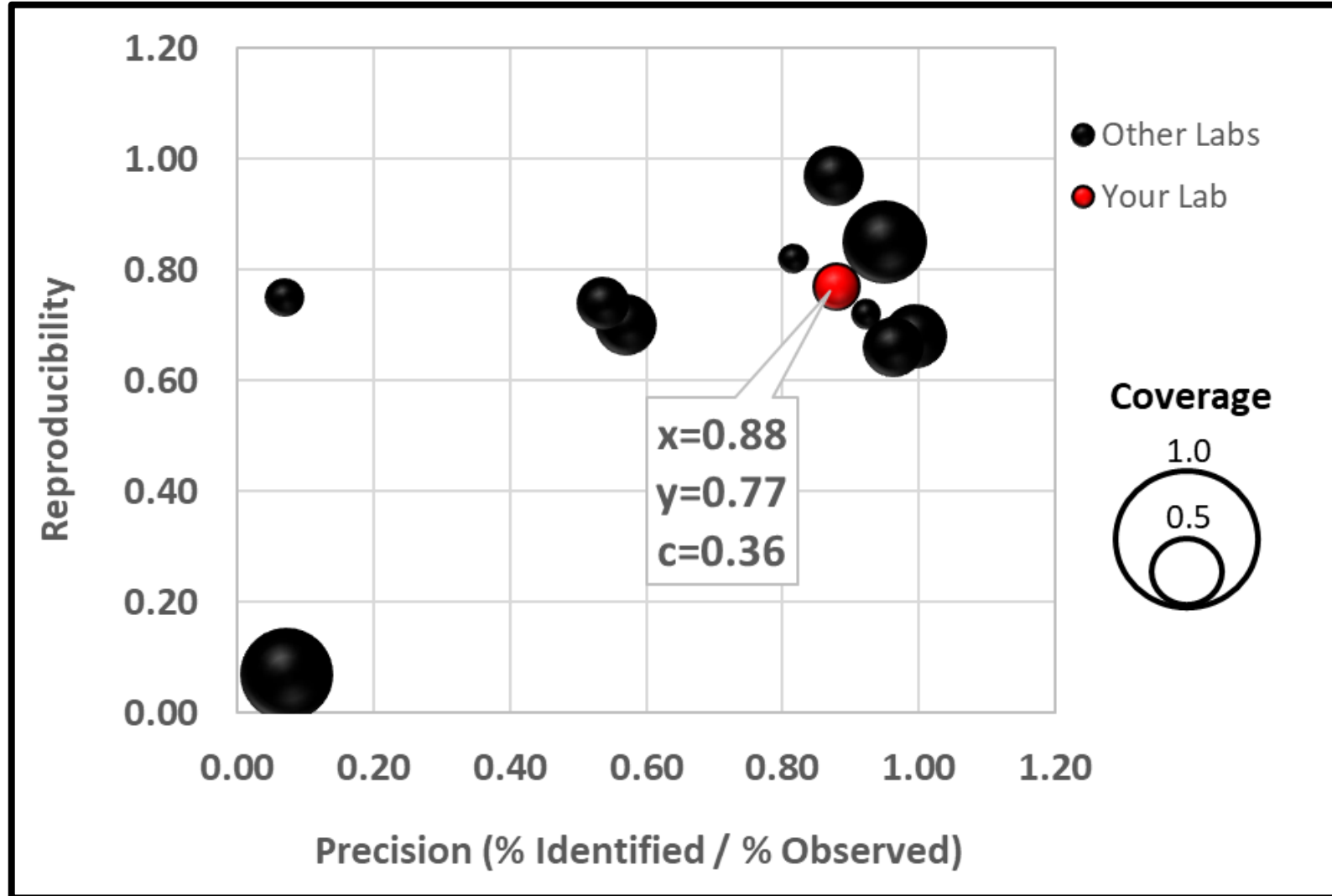
**X-Axis** →  
How often correct?

**Y-Axis** →  
How consistent?

**Bubble Size** →  
How much coverage?



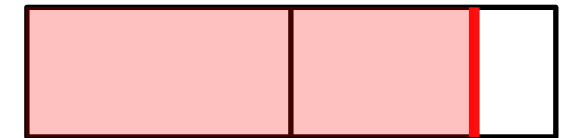
# Example Performance Report



## Performance Scores: (% of max score)

Precision: **88%**

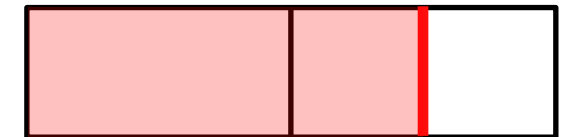
min



max

Reproducibility: **78%**

min



max

Coverage: **30%**

min



max

# Additional Results for Collaborators

- Simple performance summary file (n=1 per method):
  - # and % correct identifications per sample
- Individual results files (n=10 per method):
  - Mass match (yes/no), formula match (yes/no), compound match (yes/no)
  - Highest confidence level (as reported or after consensus revision)
- Composite results file (n=1 per method):
  - For each spiked substance (n=1,269)
    - # of spikes (1-10), # of isomer spikes (1-5)
    - # mass hits, # formula hits, # compound hits
    - Observed (yes/no/undetermined), Correct ID (yes/no), Reproducible (yes/no)

# Some Challenges (to date)

- Multiple chemical candidate submissions per feature
- Inconsistent & inaccurate use of scoring metrics
- Inconsistent & inaccurate chemical reporting procedures
- Inconsistent and unclear feature filtering protocols
- Limited engagement regarding collaborator follow-up
- Determining false positives vs. unanticipated true positives
- Determining true negatives and dependent metrics
- Slow evaluation process vs. rapid method development processes

# Summary of ENTACT Findings

- NTA methods are suitable for many ToxCast chemicals
  - ~5% of ENTACT compounds not observed by any method
- Multiple methods required for broad characterization
  - No “one size fits all” method
  - <1% of ENTACT compounds observed using all methods
- Performance determined across 3 categories:
  - **Coverage** = Ability to Observe → (Range = 22% to 69%)
  - **Precision** = Ability to Identify those Observed → (Range = 7% to 99%)
  - **Reproducibility** = Ability to Consistently Identify → (Range = 7% to 97%)

# Take-Away Messages from ENTACT (to date...)

- Lack of transparency in methods/results reporting
- Method procedures change over short time increments
- Biased self-reporting → highlight strengths, mask weaknesses
- Blinded ToxCast mixtures allow for NTA performance assessment
- Performance measures highly variable across labs/methods
- Standard performance assessment methods/benchmarks must be adopted
- Benchmarks require input/consensus from NTA community
- Community focus must be on QA/QC

# Developing and Disseminating Guidance Materials

- BP4NTA → Borne out of 2018 ENTACT workshop
- ~100 U.S. and international members
  - Government, academia, and industry
- Working Group Objectives:
  - Short term → define common NTA terms, concepts, and performance metrics
  - Short term → provide recommendations on research & reporting best practices
  - Long term → enable proficiency testing
- Products (including 3 manuscripts):
  - Website with key resources and links: <https://nontargetedanalysis.org/>
  - Guidance documents with definitions & supporting info
  - “NTA Study Reporting Tool” to standardize reporting (proposals & manuscripts)





# Building Tools to Ensure Transparency & Reproducibility

## The “NTA Study Reporting Tool” (NTA SRT):

- Standardized framework for reviewing quality of NTA reporting
- Aids NTA study design and review (proposals & manuscripts)
- Follows chronology of typical NTA studies with detailed examples
- Scale-based scoring (numeric & colorimetric) for individual study attributes
- HTML interactive version via BP4NTA website (hyperlinks → supporting docs.)
- Fillable PDF version available for download (via website)
- Comment box for periodic updates/revisions (via website)
- Working with journal editors for initial testing and deployment

# NTA Study Reporting Tool (draft version)

NTA Study Chronology

Study Sections & Categories			Example Information to Report	Numeric & Colorimetric Scoring	Rationale/Notes
Methods	Study Design	Objectives & Scope	<ul style="list-style-type: none"> <li>Study goals and hypotheses</li> <li>Scope of the study with respect to use of NTA / suspect screening</li> <li>Expected chemical coverage of approach and potential limitations</li> </ul>	1	
		Sample Information & Preparation	<ul style="list-style-type: none"> <li>Sample collection/replication, handling/storage, preparation, extraction, &amp; clean-up methods (and related QA practices)</li> <li>Intended use of samples (e.g., method development, compound identification, etc.)</li> <li>Development and intended use of blanks</li> </ul>	2	
		QC Spikes & Controls	<ul style="list-style-type: none"> <li>Development of spikes/controls (e.g., isotopically labeled standards/spikes, native standard spikes, matrix pools)</li> <li>Intended use of QC or other spikes/controls (e.g., to monitor instrument performance, data normalization, etc.)</li> </ul>	2	
	Data Acquisition	Analytical Sequence	<ul style="list-style-type: none"> <li>Sample randomization and use of replicate injections</li> <li>Inclusion of blanks and QC samples in the acquisition sequence</li> <li>Information about single vs. multiple analytical batches</li> </ul>	3	
		Chromatography	<ul style="list-style-type: none"> <li>Instrument specifications</li> <li>Method settings (e.g., column/guard, mobile phases, gradient, injection techniques)</li> </ul>	3	
		Mass Spectrometry	<ul style="list-style-type: none"> <li>Instrument specifications</li> <li>Instrument calibration and/or tuning procedures</li> <li>Method settings (e.g., ...)</li> </ul>	3	
		Software	<ul style="list-style-type: none"> <li>File conversion information</li> <li>Software program(s) used</li> <li>Workflow steps (e.g., ...)</li> <li>Feature detection thresholds</li> <li>Data correction or not</li> <li>Software programs(s) used</li> </ul>	2	
	Data Outputs	Statistical & Chemometric Outputs	<ul style="list-style-type: none"> <li>Basic statistical outputs (e.g., adj. p-values, standard deviations, test statistics)</li> <li>Results of chemometric analyses (e.g., reported classifications/groupings of features or samples, observed trends in the data)</li> <li>Visuals/plots (e.g., Venn diagrams, heatmaps, clustering dendrograms, volcano plots, network diagrams, PCA and loading plots)</li> <li>New statistical metrics, algorithms, packages, and/or scripts</li> </ul>	NA	
		Identification & Confidence Levels	<ul style="list-style-type: none"> <li>Reported identifications and associated confidence levels (e.g., levels described by Schymanski et al.)</li> <li>Supporting data for annotation/identification (e.g., formula match scores, fine isotope pattern, retention time match, MS/MS match scores, source of MS/MS spectra)</li> <li>For features with lower confidence IDs, (i.e., not standard-confirmed), proposed tentative structures and other annotated data</li> <li>Semi-quantification or quantification data</li> <li>Exported MS/MS spectra (e.g., as a library, database, or deposition into online repository)</li> </ul>	3	
		Data Acquisition QA/QC	<ul style="list-style-type: none"> <li>Quality: Adherence to QA/QC protocols for sample preparation and data acquisition</li> <li>Boundary: Description of the potential impacts of methods (sample prep, chromatographic, MS) on observable chemical space</li> <li>Accuracy: Reported chromatographic and mass accuracy</li> <li>Precision: Variability of observed retention time, precursor mass error, and abundance</li> </ul>	1	
Results	QA/QC Metrics	Data Processing & Analysis QA/QC	<ul style="list-style-type: none"> <li>Quality: Outcomes of QC checks along the data processing &amp; analysis workflow</li> <li>Boundary: Impact of data processing &amp; analysis method(s) on observed chemical space, observed limits of detection/ID</li> <li>Accuracy: Performance measures (True Positive Rate, False Positive Rate, etc.) for known compounds or samples with known classification</li> <li>Precision: Reproducibility/repeatability of performance measures for known compounds or samples with known classification; Calculations such as False Discovery Rate, F1 score, etc.</li> </ul>	0	

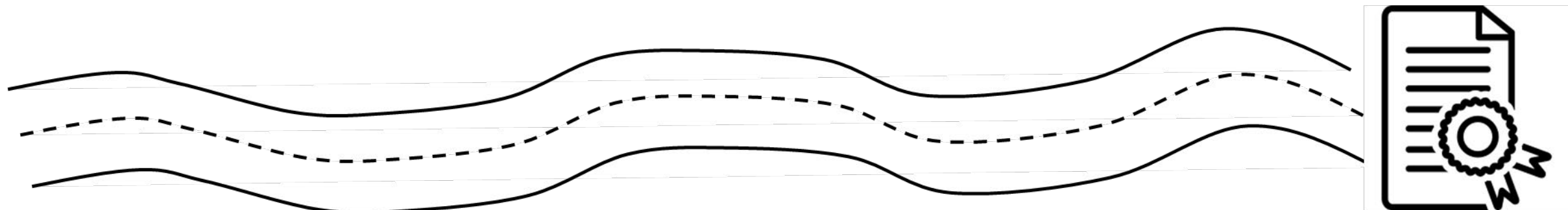
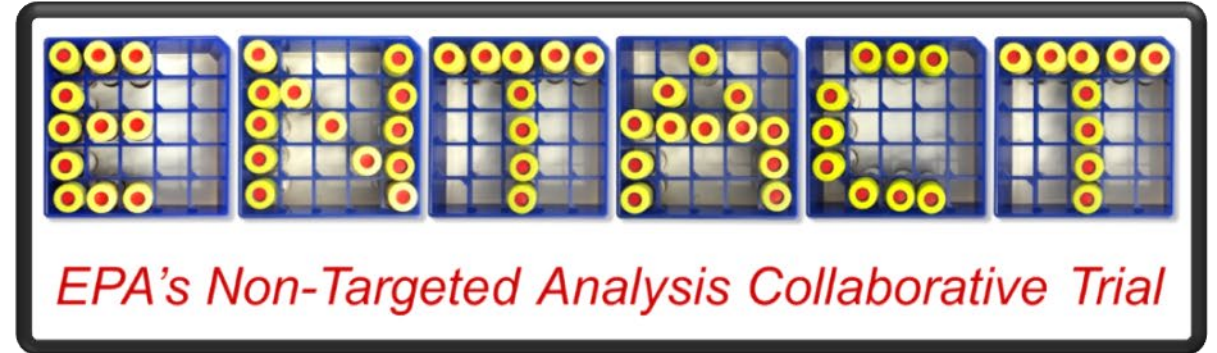
Hyperlinked  
(HTML version)  
to supporting  
information

3-4 bullet point examples for each of the 13 sub-categories

Not exhaustive – intended to guide reviewers;  
relies on reviewer expertise/discretion.

Space for  
reviewer to  
explain  
assigned  
score

# The Path to NTA Lab Credentialing



# Contributing Researchers

## (EPA Affiliation Unless Otherwise Noted)

- **ENTACT:**

- ***Co-leads:*** E. Ulrich and J. Sobus
- ***Research Team:*** A. Williams, A. Chao, S. Newton, C. Lowe, C. Grulke, A. Richard, J. Grossman (ORISE)

- **BP4NTA:**

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- ***Website Co-leads:*** S. Newton and S. Nason (CAES)

- **NTA SRT:**

- ***Co-leads:*** K. Peter (NIST) and A. Phillips
- ***Research Team:*** P. Gardinali (FIU), A. Knolhoff (FDA), C. Manzano (SDSU), K. Miller, M. Pristner & B. Warth (U. of Vienna), L. Sabourin & M. Sumarah (Agri-Food Canada), J. Sobus



# Additional EPA Contributors



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Credit: the Research Triangle Foundation

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# Questions?

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*The views expressed in this presentation are those of the author and do not necessarily represent the views or policies of the U.S. Environmental Protection Agency.*