



CSS Webinar

The In Vitro Developmental Neurotoxicity Testing Battery

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Disclosure Statement

This work has been funded by the US. Environmental Protection Agency. I have no conflicts to declare.

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- I. The Need for Alternative Approaches for Developmental Neurotoxicity Hazard Assessment
- II. The DNT In vitro battery (DNT-IVB)
- III. Applying the DNT-IVB



Drivers for NAMs



UNITED STATES ENVIRONMENTAL PROTECTION AGENCY
WASHINGTON, D.C. 20460

September 10, 2019

THE ADMINISTRATOR

MEMORANDUM

SUBJECT: Directive to Prioritize Efforts to Reduce Animal Testing

FROM: Andrew R. Wheeler
Administrator

TO: Associate Deputy Administrator
General Counsel
Assistant Administrators
Inspector General
Chief Financial Officer
Chief of Staff
Associate Administrators
Regional Administrators

During my March 2019 all-hands address, I reiterated the U.S. Environmental Protection Agency's commitment to move away from animal testing. We are already making significant efforts to reduce, replace and refine our animal testing requirements under both statutory and strategic directives. For example, the *Toxic Substances Control Act*, amended June 22, 2016, by the Frank R. Lautenberg Chemical Safety for the 21st Century Act, requires the EPA to reduce reliance on animal testing. Also, Objective 3.3 of the *FY 2018-2022 U.S. EPA Strategic Plan* outlines a commitment to further reduce the reliance on animal testing within five years. More than 200,000 laboratory animals have been saved in recent years as a result of these collective efforts.

Scientific advancements exist today that allow us to better predict potential hazards for risk assessment purposes without the use of traditional methods that rely on animal testing. These new approach methods (NAMs), include any technologies, methodologies, approaches or combinations thereof that can be used to provide information on chemical hazard and potential human exposure that can avoid or significantly reduce the use of testing on animals. The benefits of NAMs are extensive, not only allowing us to decrease animals used while potentially evaluating more chemicals across a broader range of potential biological effects, but in a shorter timeframe with fewer resources while often achieving equal or greater biological predictivity than current animal models.

Lautenberg Amendment to TSCA requires EPA to assess more chemicals

- Authorizes the use of alternative approaches

USEPA Administrator Memo Prioritizing Efforts to Reduce Animal Testing, September 10, 2019

- Sets goals for EPA to reduce and ultimately eliminate requests for, and our funding of, mammal studies
- Form a working group of agency experts in this field who will provide a work plan within six months. <https://www.epa.gov/chemical-research/new-approach-methods-work-plan>
- <https://www.epa.gov/environmental-topics/administrator-memo-prioritizing-efforts-reduce-animal-testing-september-10-2019>

In Europe, REACH legislation requires data on all compounds used in commerce

Under FIFRA, Office of Pesticides wants information regarding DNT that can help design more focused in vivo DNT studies (when they are conducted)



Requirements of EPA 870.6300 (OECD TG 426/443)

- 6 Pregnant females/dose (20 litters/dose recommended)
- 10 pups/litter (5 male/5 female)
- Minimum 3 doses + control
- Dosing period GD6-PND10
- Assessments on PND 4, 11, 21, 35, 45, 60
- Signs of Maternal Toxicity
- Developmental landmarks
- Brain/body weights (4, 11, 17, 21 PND)
- Motor activity (13, 17, 21, 60 PND)
- Auditory Startle (weaning, PND 60)
- Learning and memory (weaning, PND 60)
- Neuropathology (PND 11 and termination)
 - Major brain regions

<https://beta.regulations.gov/document/EPA-HQ-OPPT-2009-0156-0042>

https://www.oecd-ilibrary.org/environment/test-no-426-developmental-neurotoxicity-study_9789264067394-en

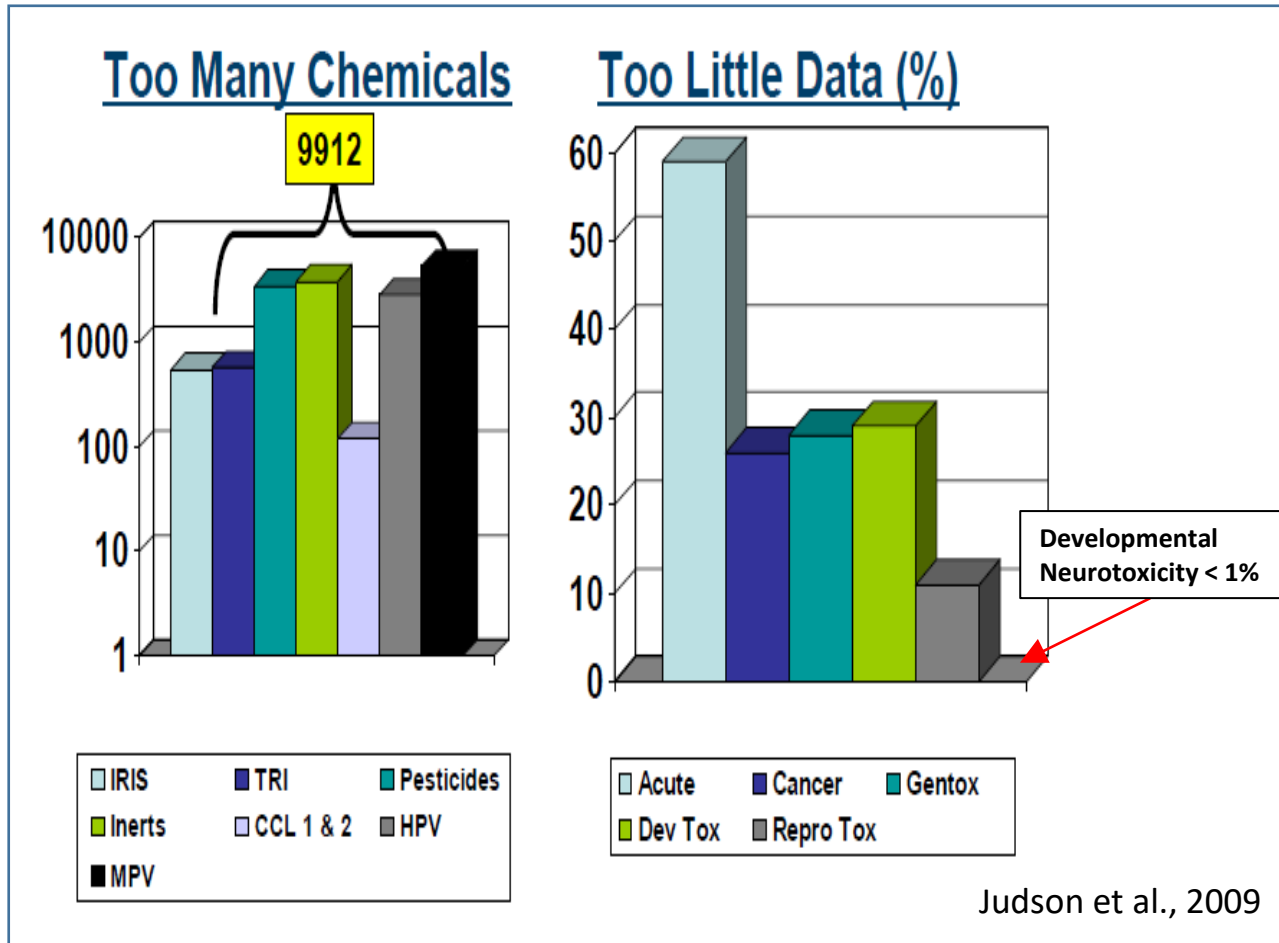
<https://www.oecd.org/chemicalsafety/test-no-443-extended-one-generation-reproductive-toxicity-study-9789264185371-en.htm>

- “Triggered” test- Only requested if concern for neurotoxicity
- Expensive- ~\$1,000,000/chemical
- Time-consuming- takes 1-2 years to complete
- Ethically questionable- Estimated ~1000 animals/test
- Value of Information
 - Quality of data varies considerably
 - Not often used for point of departure values for risk assessment*

*Raffaele et al. [The use of developmental neurotoxicity data in pesticide risk assessments](#). Neurotoxicol Teratol. 2010 Sep-Oct;32(5):563-72.



Data on Developmental Neurotoxicity (DNT) Hazard is Not Available for Too Many Chemicals



Current testing too slow

- Not Required under FIFRA or TSCA
- At current pace, ~150 chemicals in 20+ yrs

The absence of DNT hazard data on chemicals impedes consideration of this adverse outcome in environmental decision-making.

Reports of the potential involvement of environmental chemicals in increased rates of neurodevelopmental disease contributed to increasing public concern about DNT hazard of chemicals

*Raffaele et al. [The use of developmental neurotoxicity data in pesticide risk assessments](#). Neurotoxicol Teratol. 2010 Sep-Oct;32(5):563-72.



New Approach Methodologies (NAMs) are Needed for Developmental Neurotoxicity (DNT) Hazard

Solution: Faster, inexpensive and predictive methods are needed to detect and characterize compounds with developmental neurotoxicity hazard

- Develop high throughput, *in vitro* assays,
- Characterize chemicals for developmental neurotoxicity hazard
- Use human models whenever possible
- Data from these assays can provide information for decision-making



Encouraging Regulatory Use of NAMs

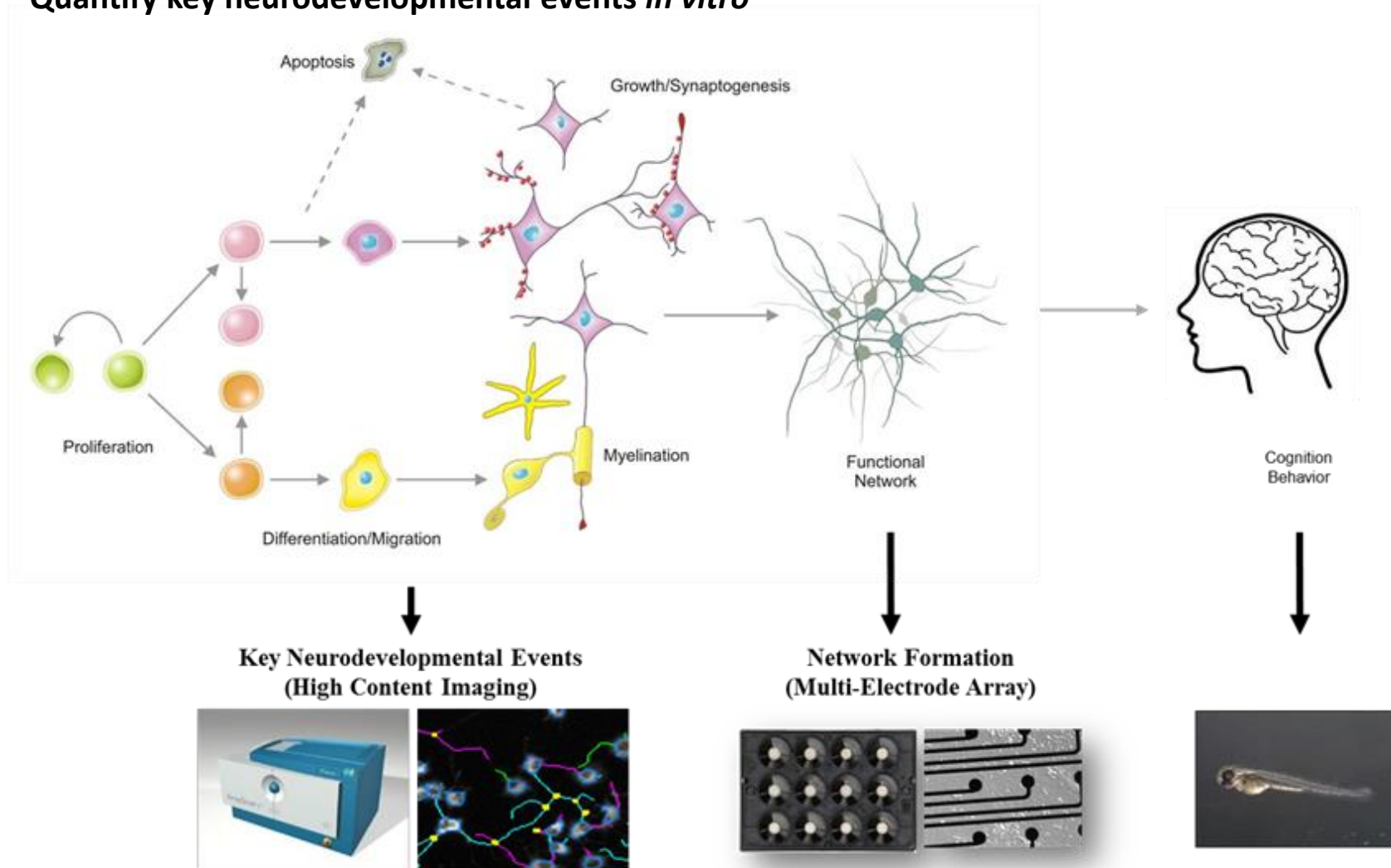
- Understanding of how the assays work and what they measure
- Evaluation of individual assays and the battery of assays
- Data from alternative assays
- Understanding of what can be done with the data
- Accessibility to the data (Toxcast)

Regulatory decision-makers must have confidence in the assays and data in order to incorporate them into the decision-making process

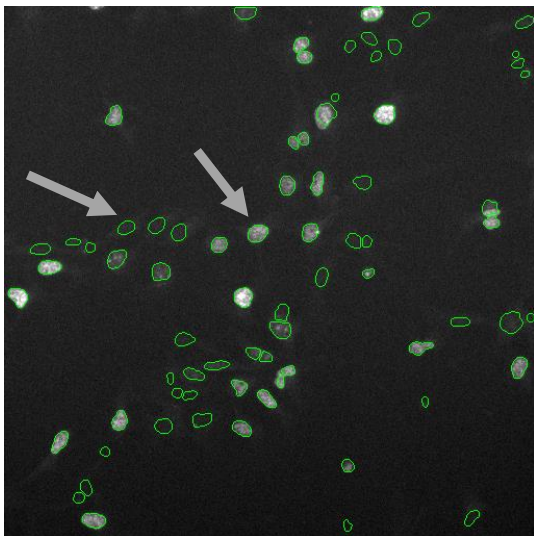
- Central nervous system development is complex
 - Multiple potential targets
 - Time-dependent processes
 - Spatially-dependent processes
- Which target? Where? When?

Therefore, focus research on *key neurodevelopmental processes*

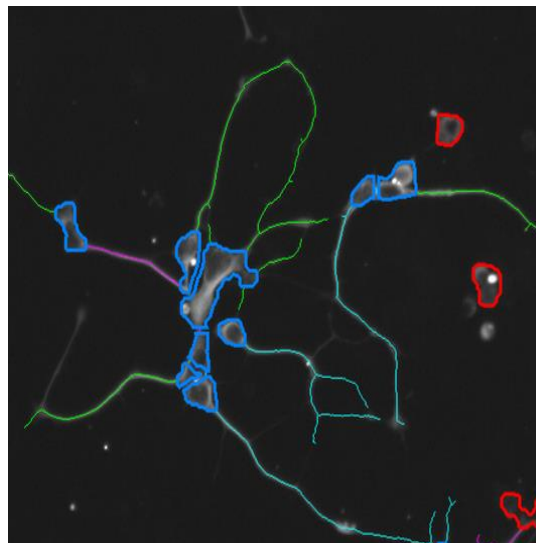
Quantify key neurodevelopmental events *in vitro*



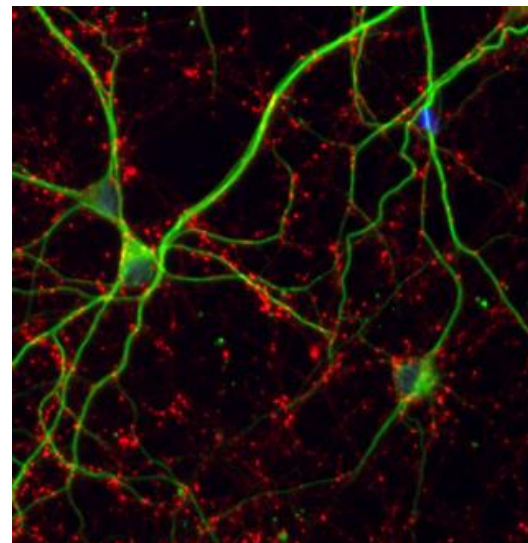
Proliferation



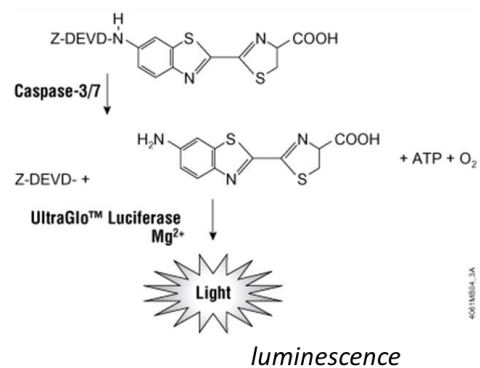
Neurite Outgrowth



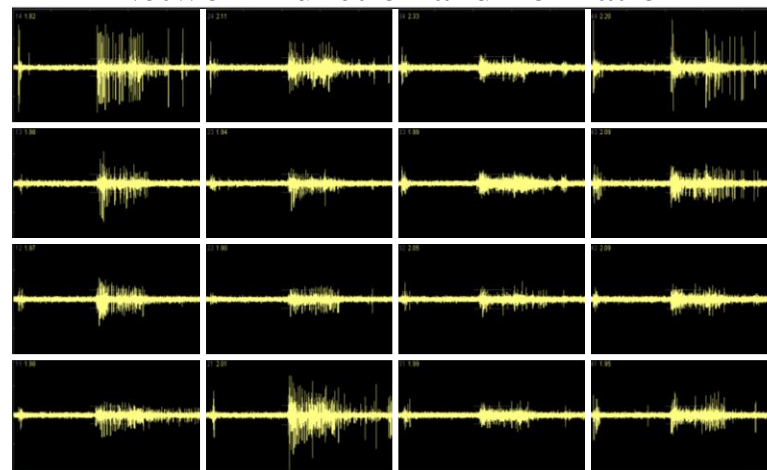
Synaptogenesis



Apoptosis

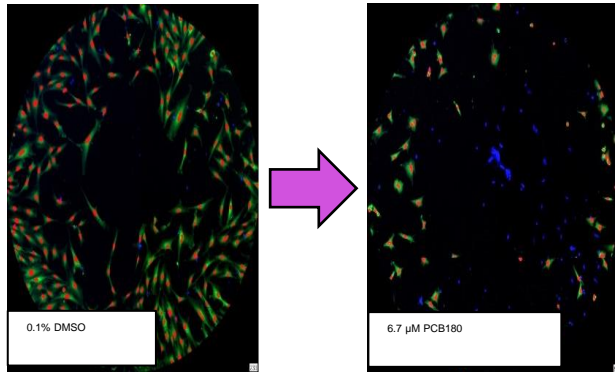


Network Function and Formation

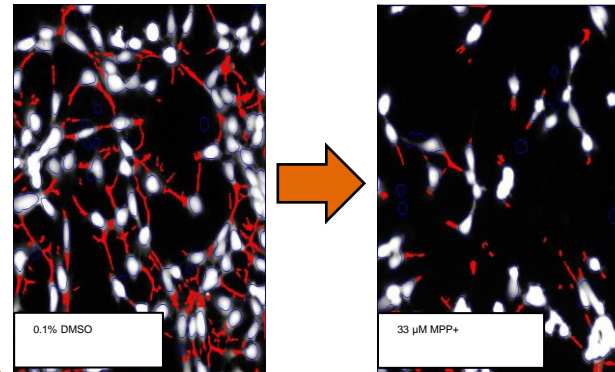


“EFSA Sponsored” DNT NAM Assays

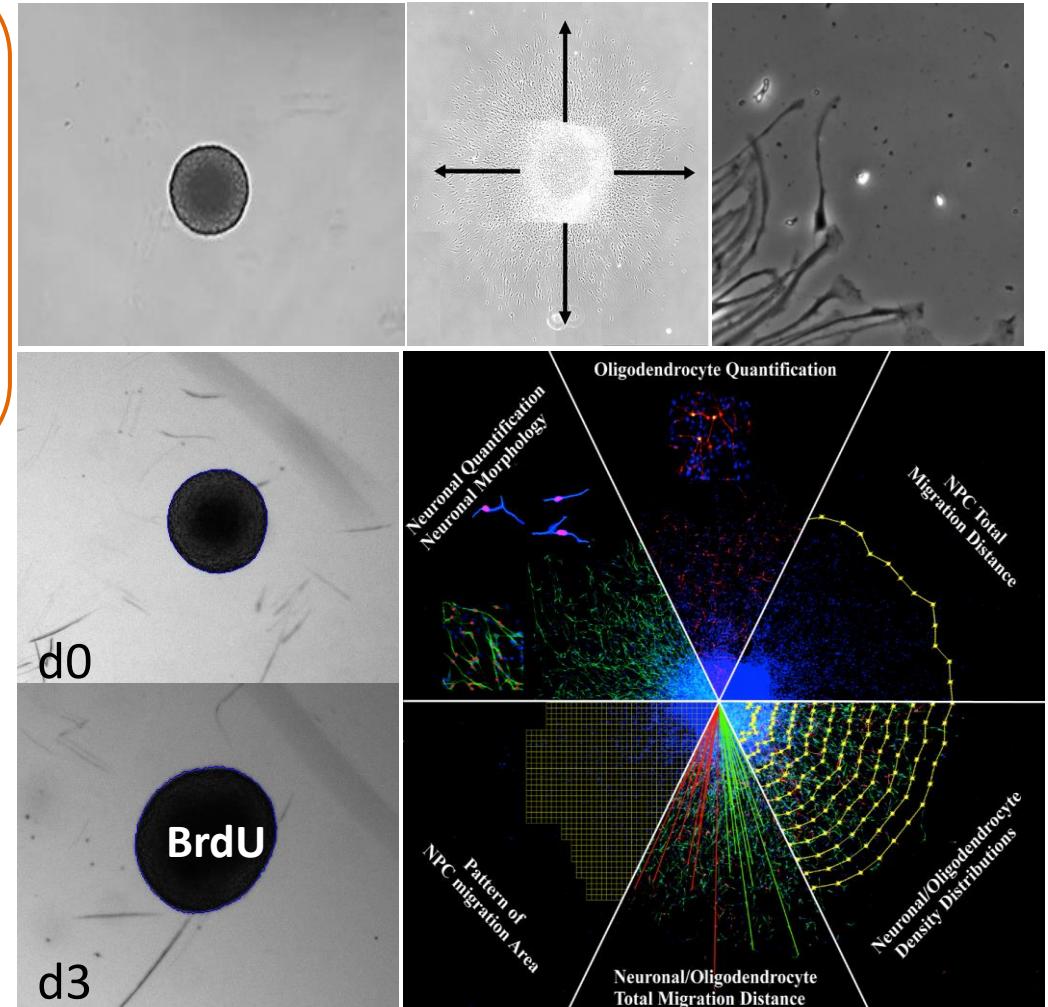
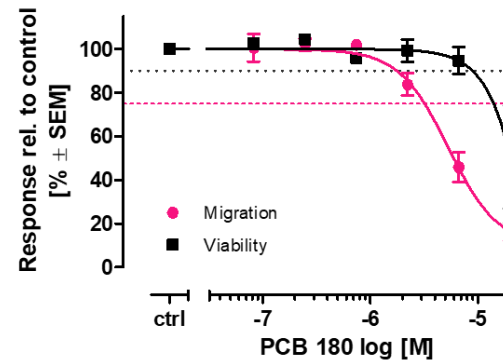
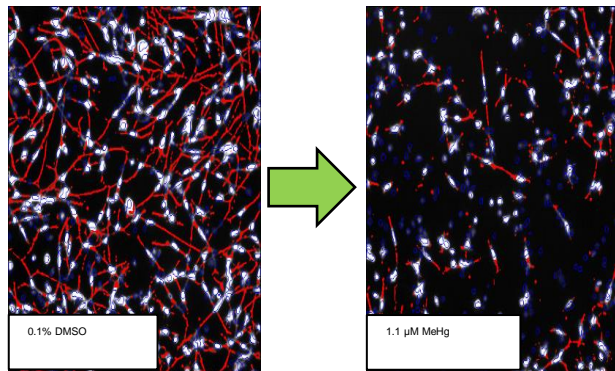
Effect on migration of neural crest cells



Effect on neurite outgrowth of CNS neurons



Effect on neurite outgrowth of PNS neurons



FORUM

International Regulatory and Scientific Effort for Improved Developmental Neurotoxicity Testing

Magdalini Sachana,^{*,1} Anna Bal-Price,[†] Kevin M. Crofton,[‡] Susanne H. Bennekou,[§] Timothy J. Shafer,^{||} Mamta Behl,^{||} and Andrea Terron^{|||}

Towards regulatory DNT testing: Alternative methods

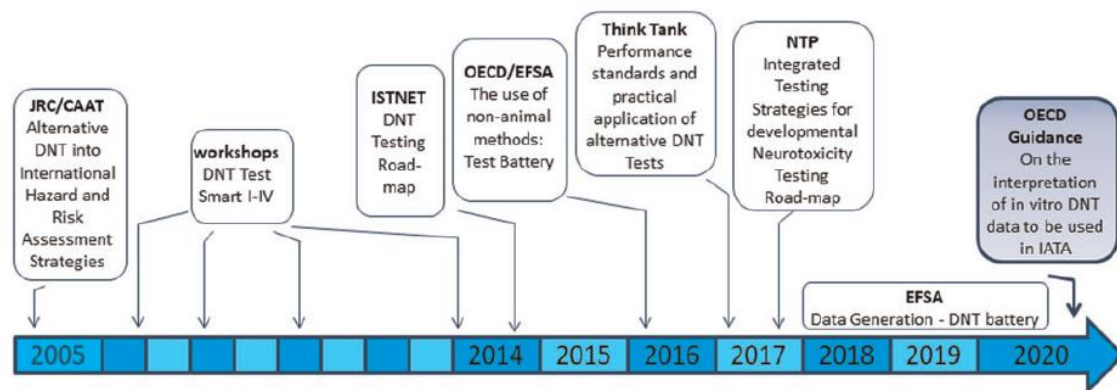
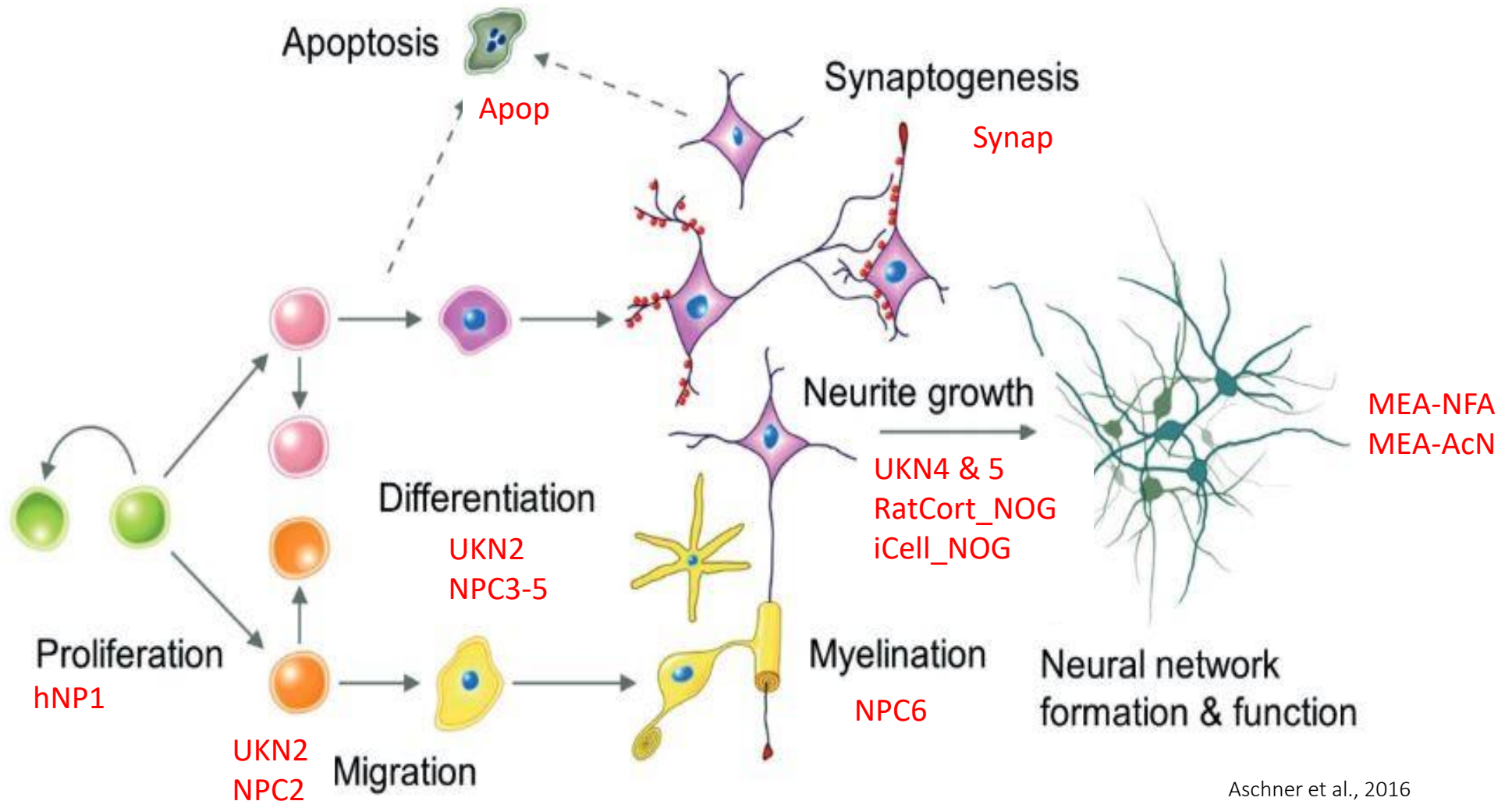


Figure 1. Timeline of efforts to develop and implement new alternative methods for developmental neurotoxicity.

Table 2. Proposed Assays for Evaluation As an *In Vitro* DNT Battery

Process	Assays	References
Proliferation	hNP1	Harrill et al. (2018)
	NPC1	Baumann et al. (2016) and Barenys et al. (2017)
Apoptosis Migration	UKN1	Balmer et al. (2012)
	hNP1	Harrill et al. (2018)
	NPC2	Baumann et al. (2016) and Barenys et al. (2017)
Neuron differentiation	UKN2	Nyffeler et al. (2017)
	NPC3	Baumann et al. (2016) and Barenys et al. (2017)
Oligodendrocyte differentiation & maturation Neurite outgrowth	NPC5/6	Baumann et al. (2016) and Barenys et al. (2017)
	iCell gluta hN2	Harrill et al. (2018)
	UKN 4 & 5 NPC4	Krug et al. (2013) Baumann et al. (2016) and Barenys et al. (2017)
Synaptogenesis	Rat primary synaptogenesis	Harrill et al. (2018)
Network formation	MEA-NFA	Brown et al. (2016) and Frank et al. (2018)





Encouraging Regulatory Use of NAMs

- Understanding of how the assays work and what they measure
- Evaluation of individual assays and the battery of assays
- Data from alternative assays
- Understanding of what can be done with the data
- Accessibility to the data

Regulatory decision-makers must have confidence in the assays and data in order to incorporate them into the decision-making process

Several different approaches can be taken to evaluate the performance of the DNT-NAMS

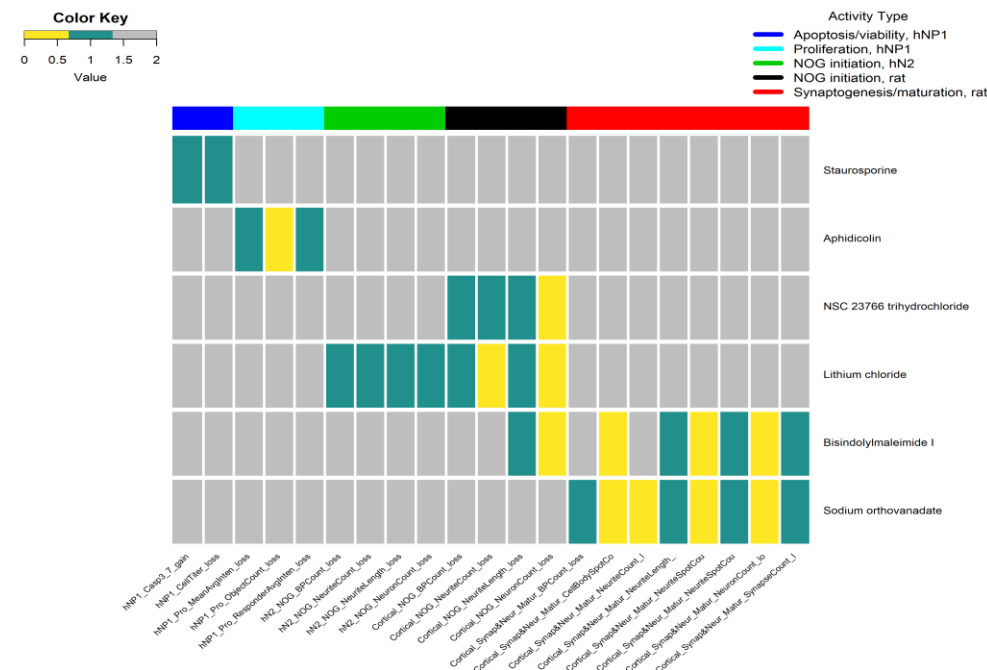
- Demonstrate that they recapitulate the *in vivo* neurobiology
- Evaluate Performance of Positive Controls
- Evaluate the Reproducibility when retesting compounds
- Evaluation of the Sensitivity/Specificity

True Positive Rate (sensitivity) = True positives/Known Positives

True Negative Rate (specificity) = True negatives/Known Negatives

Precision = True positives/(True Positives + False Positives)

Accuracy = (True Positives + True Negatives)/(Known Positives + Known Negatives)





The Challenge of Evaluating DNT NAMs Sensitivity and Specificity

NTP Report on Human Carcinogens (2016)

- 62 recognized, human carcinogens
- >170 “Anticipated” human Carcinogens
- >1000 compounds evaluated

By Contrast, for DNT:

- 12 recognized human developmental neurotoxicants (Grandjean and Landrigan, Lancet Neurol. 2014).
- ~150 compounds evaluated in Guideline DNT studies (rodents).

As a result, benchmarking the sensitivity/specificity of *in vitro* DNT assays against *in vivo* data is challenging, but still needs to be done



Goal: Assess the level of information in the literature that a chemical has *DNT hazard* or does not cause DNT

Evaluating 500 putative “DNT Reference” Compounds..

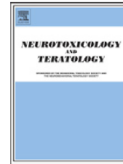
Neurotoxicology and Teratology 52 (2015) 25–35



Contents lists available at ScienceDirect

Neurotoxicology and Teratology

journal homepage: www.elsevier.com/locate/neutera



Review article

Expanding the test set: Chemicals with potential to disrupt mammalian brain development



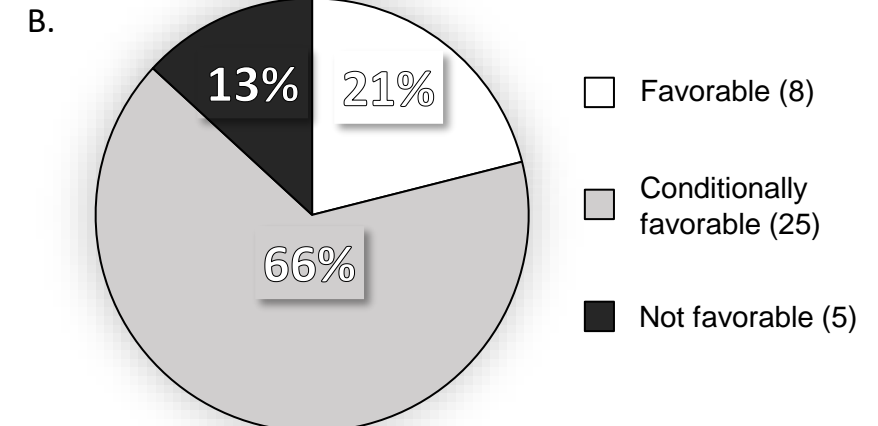
William R. Mundy ^{a,*}, Stephanie Padilla ^a, Joseph M. Breier ^{a,1}, Kevin M. Crofton ^b, Mary E. Gilbert ^a, David W. Herr ^a, Karl F. Jensen ^a, Nicholas M. Radio ^{a,2}, Kathleen C. Raffaele ^c, Kelly Schumacher ^d, Timothy J. Shafer ^a, John Cowden ^b

Identified ~100 compounds that have evidence of causing DNT in mammals

These are *scientific* summaries of evidence, not *regulatory* decisions. Summaries reflect *hazard*, not exposure or risk.

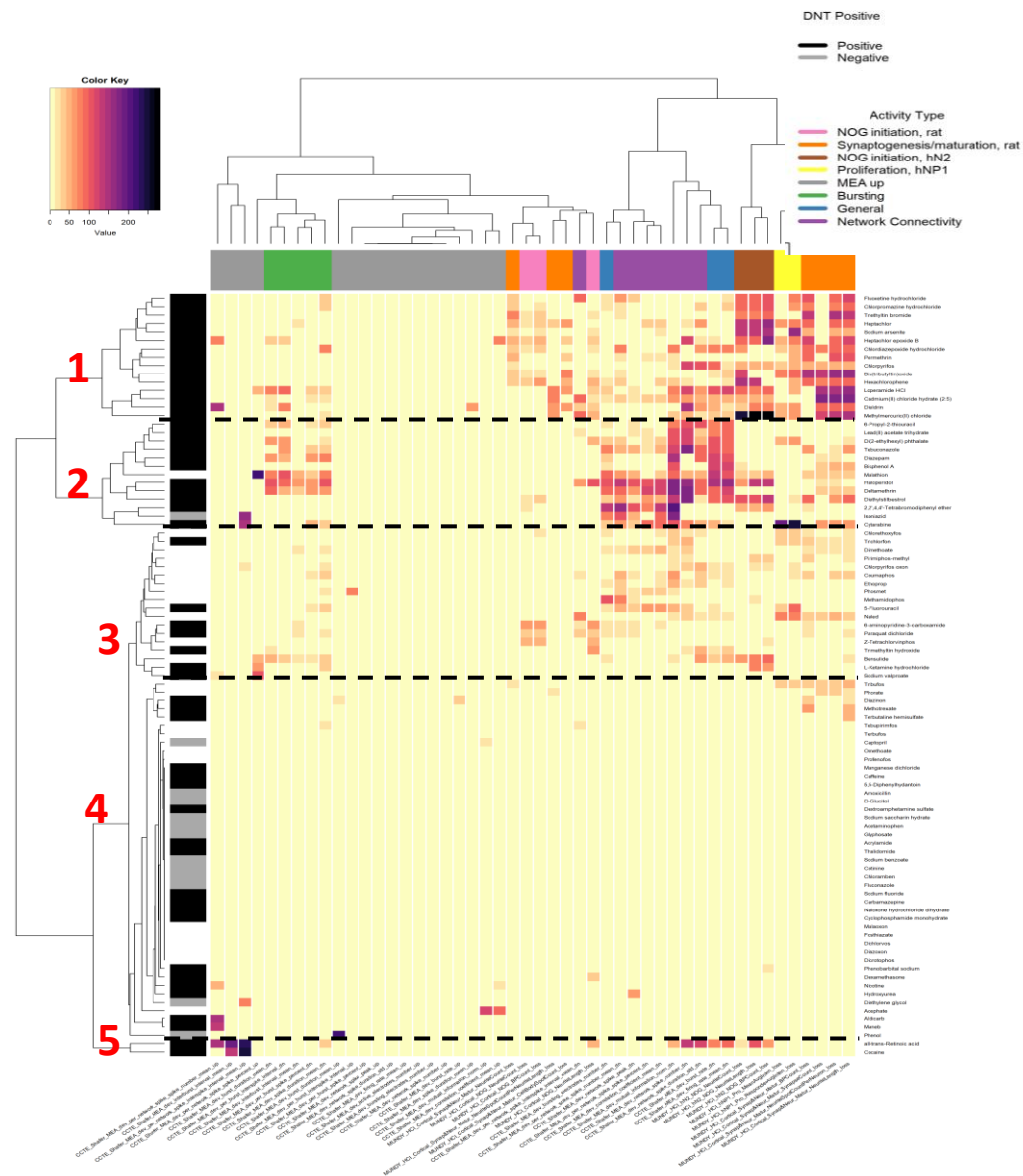
... and 38 putative “DNT Negative” Compounds

A. Categorical definitions	
Favorable	Compounds in this category have convincing evidence for a lack of developmental neurotoxicity (DNT). If there are caveats, they are considered minor.
Conditionally favorable	Compounds in this category may be used as negative DNT reference compounds as there was no clear evidence that they cause DNT in vivo. However, caveats such as lack of (or conflicting) data, experimental design challenges, and/or chemistry/toxicokinetic uncertainties should be duly considered
Not favorable	These compounds have evidence that indicates they may have potential effects on the developing nervous system.



Sensitivity/Specificity Analysis for EPA DNT NAMs

55 DNT Reference and 13 DNT Negative Compounds



		DNT Reference	
		Negative	Positive
1	Proliferation, synaptogenesis, NOG (hN2 cells)	0	15
2	Decreased network formation activity	1 Isoniazid	11
3	Moderate to low activity across endpoints	0	7
4	Inactive/ equivocal	12	19
5	Increased mean inter-spike interval for network spikes	0	2

		Negatives	Positives
Results from DNT-NAM battery	Selective activity (Clusters 1,2,3,5)	False positive:1	True positive: 35
	Inactive/ equivocal (Cluster 4)	True Negative: 12	False negative: 19

Sensitivity= 65%, Specificity= 92%,

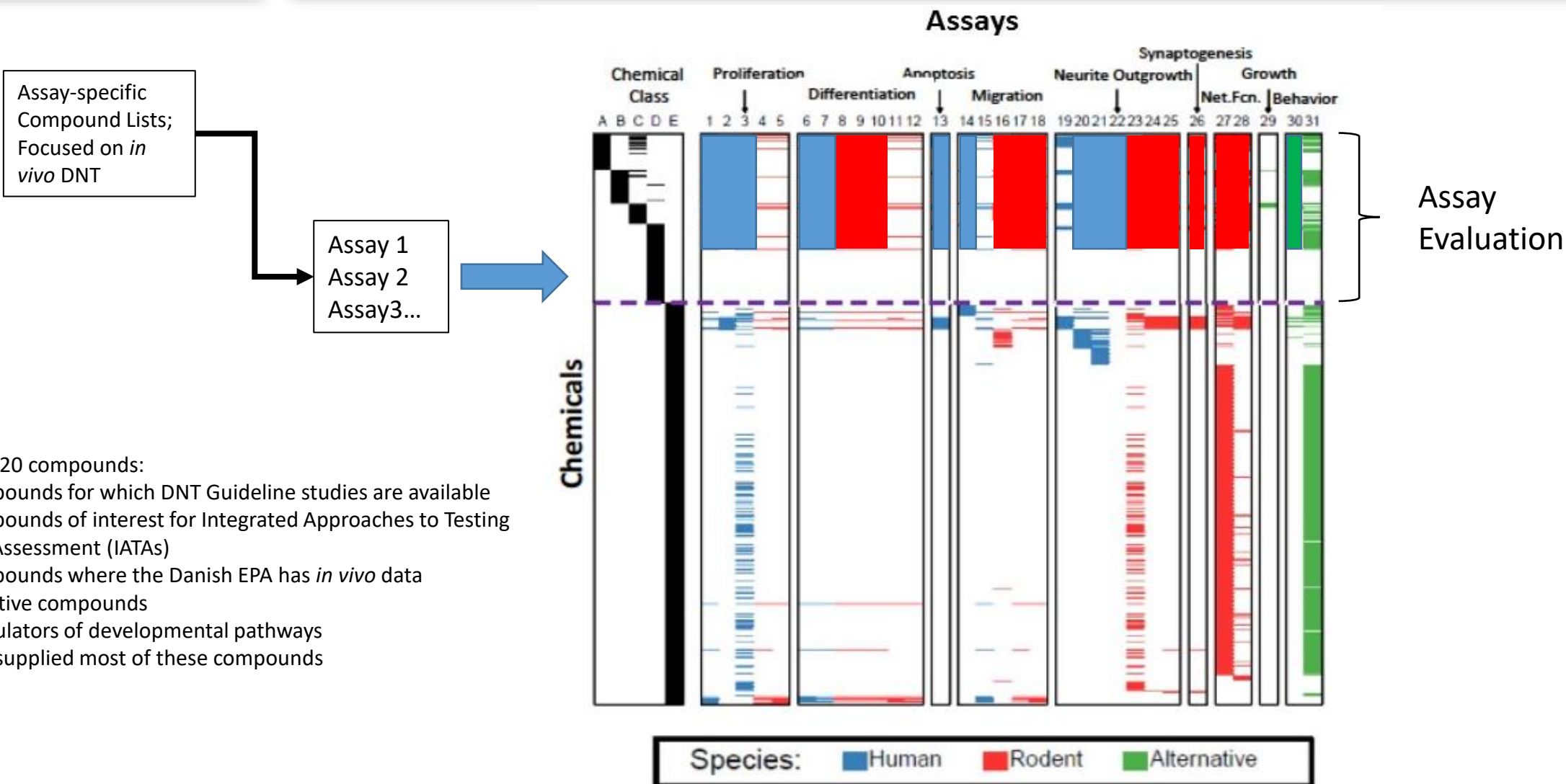
Carstens et al., in preparation

- Understanding of how the assays work and what they measure
- Evaluation of individual assays and the battery of assays
- **Data from alternative assays**
- Understanding of what can be done with the data
- Accessibility to the data

Regulatory decision-makers must have confidence in the assays and data in order to incorporate them into the decision-making process



The Need for More Data: Priority on compounds with *in vivo* DNT information



- Identified ~120 compounds:
 - Compounds for which DNT Guideline studies are available
 - Compounds of interest for Integrated Approaches to Testing and Assessment (IATAs)
 - Compounds where the Danish EPA has *in vivo* data
 - Negative compounds
 - Modulators of developmental pathways
- ToxCast has supplied most of these compounds

- Partners have received ToxCast compounds.
 - Testing is Completed at Konstanz and Düsseldorf
 - Report has been released to the public.
 - <https://www.efsa.europa.eu/en/supporting/pub/en-1938>
- EPA testing is Completed
 - Data expected in Mid-2021
- Zebrafish behavioral testing
 - Focus on ~30 IATA compounds
 - Data collection has started and will be completed later in 2021.
- NTP is distributing a set of ~100 compounds for testing, to begin in mid-2021



Encouraging Regulatory Use of DNT NAMs

- Understanding of how the assays work and what they measure
- Evaluation of individual assays and the battery of assays
- Data from alternative assays
 - Particularly for compounds that will be used for IATA case studies
- Understanding of what can be done with the data
- Accessibility to the data

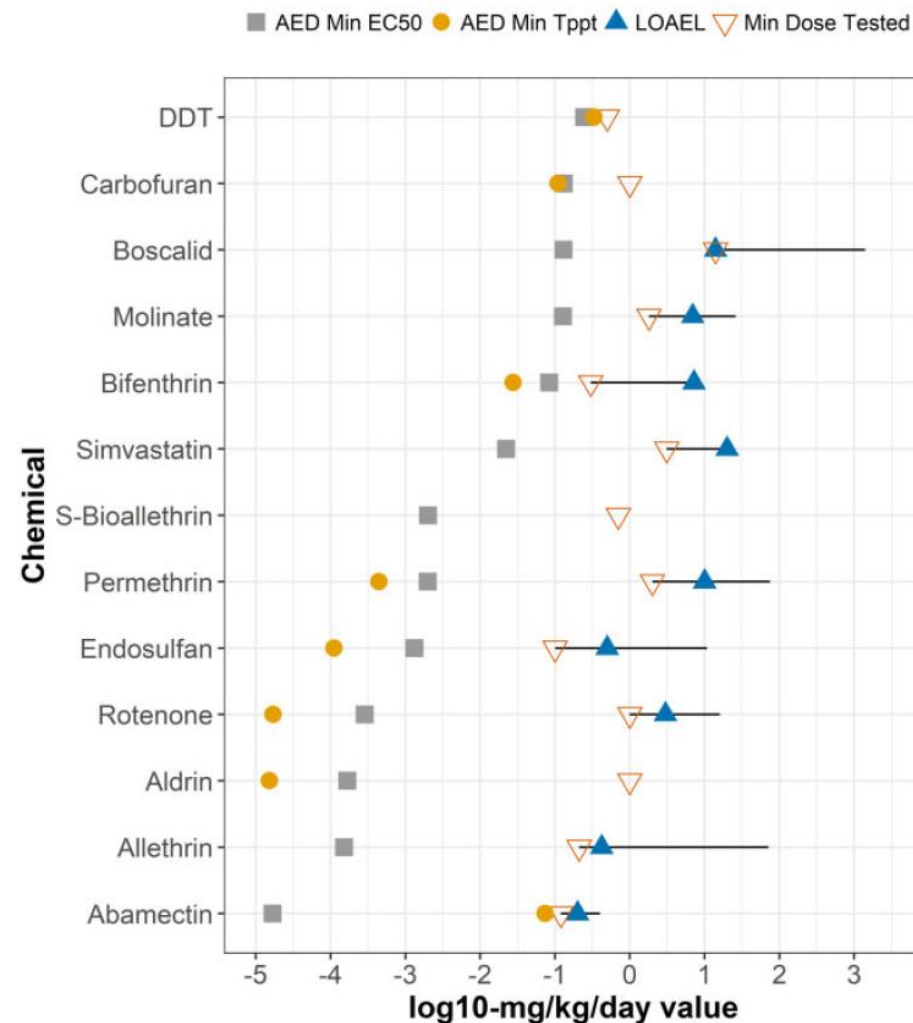
Regulatory decision-makers must have confidence in the assays and data in order to incorporate them into the decision-making process

Evaluation of Chemical Effects on Network Formation in Cortical Neurons Grown on Microelectrode Arrays

Timothy J. Shafer,^{*,1} Jasmine P. Brown,^{*,2} Brittany Lynch,[†]
Sylmarie Davila-Montero,[‡] Kathleen Wallace,^{*} and Katie Paul Friedman[§]

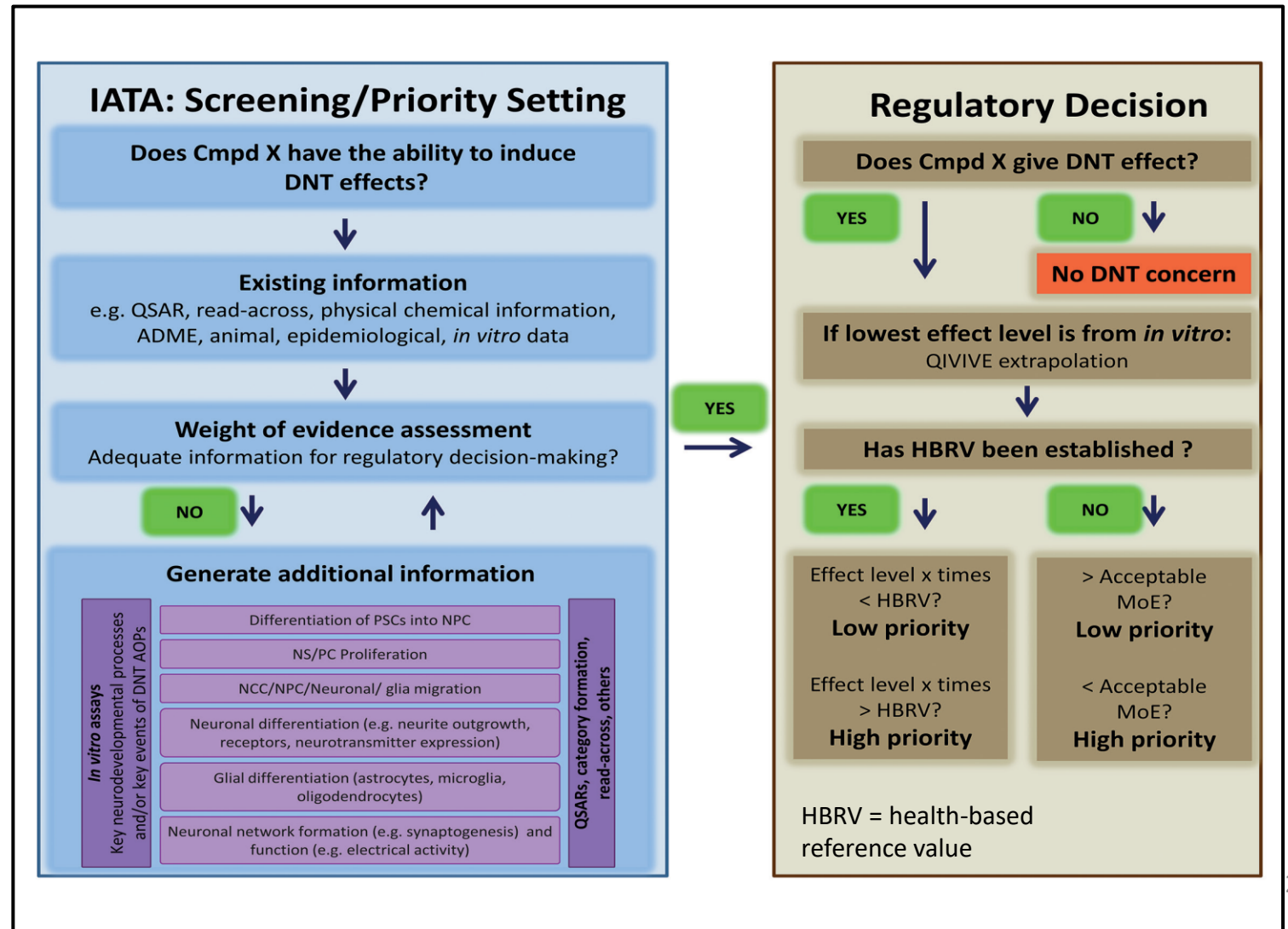
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These data indicate that DNT NAMs can be sensitive indicators of potential disruption of nervous system development



OECD DNT Expert group is Developing a Guidance Document for the use of DNT alternative assays in Integrated Approaches for Testing and Assessment (IATAs)

- Introduction and Rationale
- Issues with the Current Guideline testing approaches
- **Guidance for incorporation of *in vitro* assays into IATAs**
- **Case Studies**
 - Prioritization
 - Weight of Evidence decisions
 - Chemical specific decisions
- **Expected late 2021**



I. Screening Level information

- APCRA, TSCA, PFAS

II. Understanding species differences

- Data from DNT NAMs provided to OPP to help understand rodent-human differences in response to chemicals since the battery has both rodent and human assays

III. Structure-activity relationships

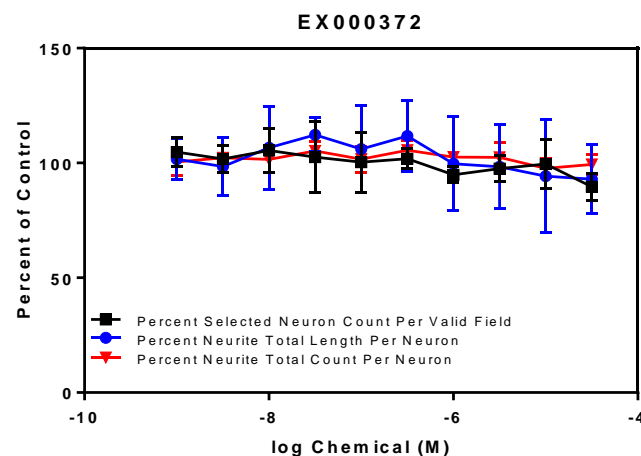
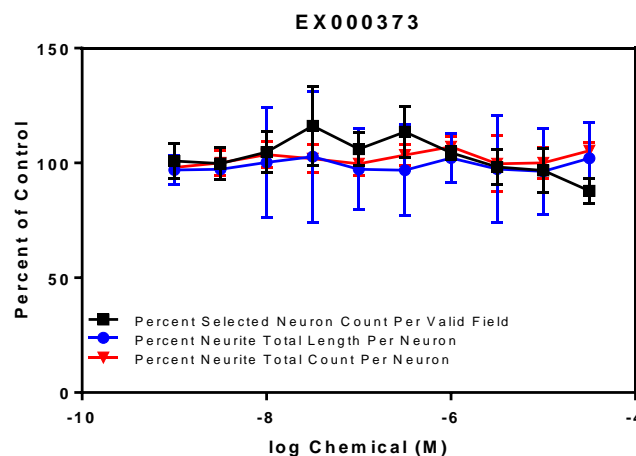
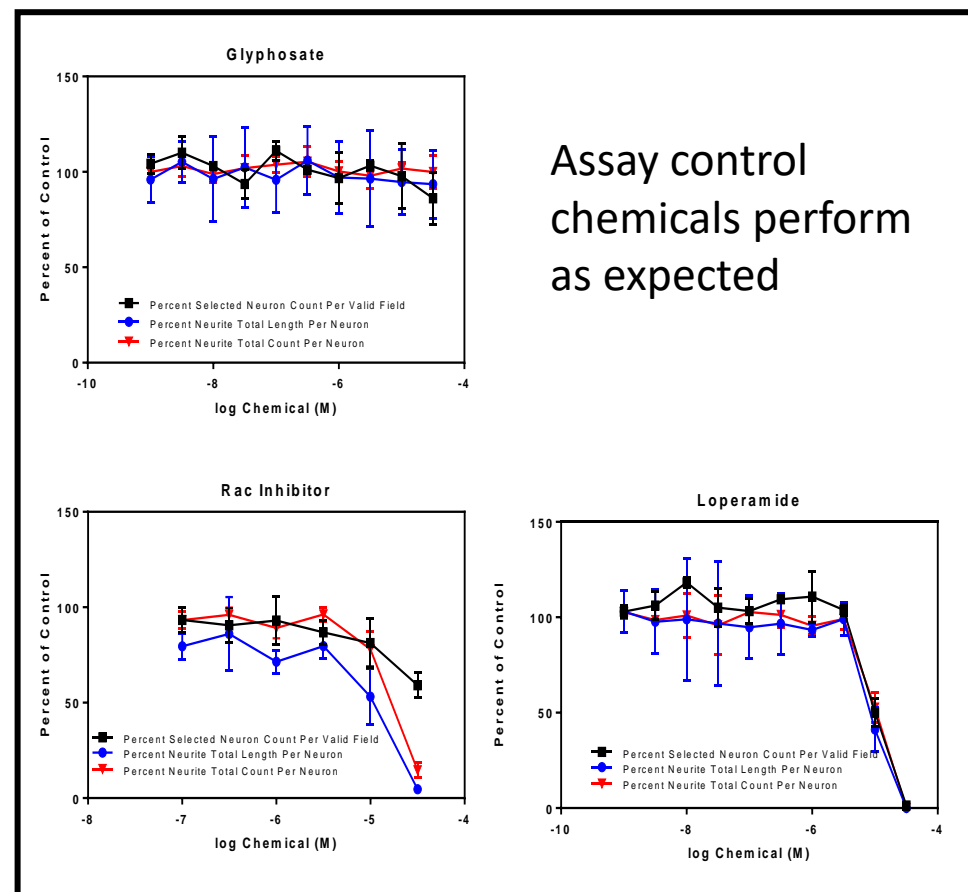
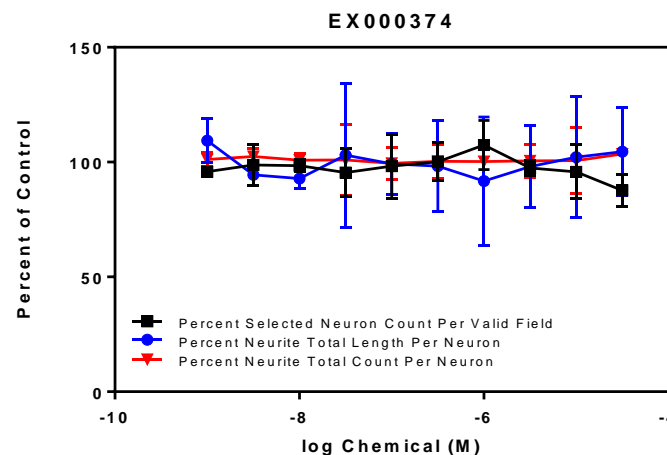
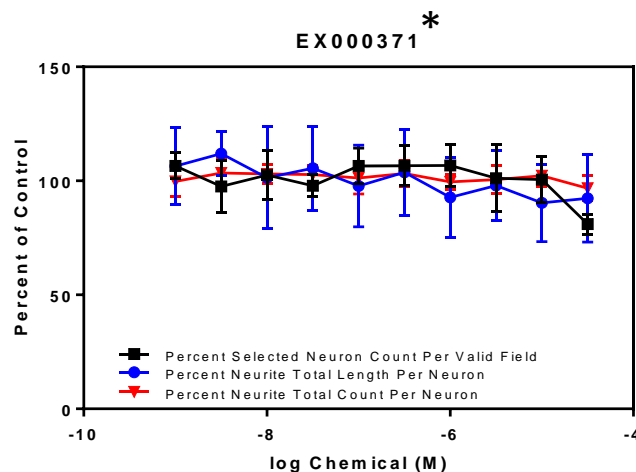
- If we have a guideline study for one compound, can we use DNT NAMs to understand structurally similar compounds?
 - Example with compound X and structurally similar analogs
 - DNT Guideline exists for X, should it be required for the analogs?

IV. Weight of Evidence approaches

- Organophosphates



DNT NAMs data for Compound X and analogs on Neurite Outgrowth

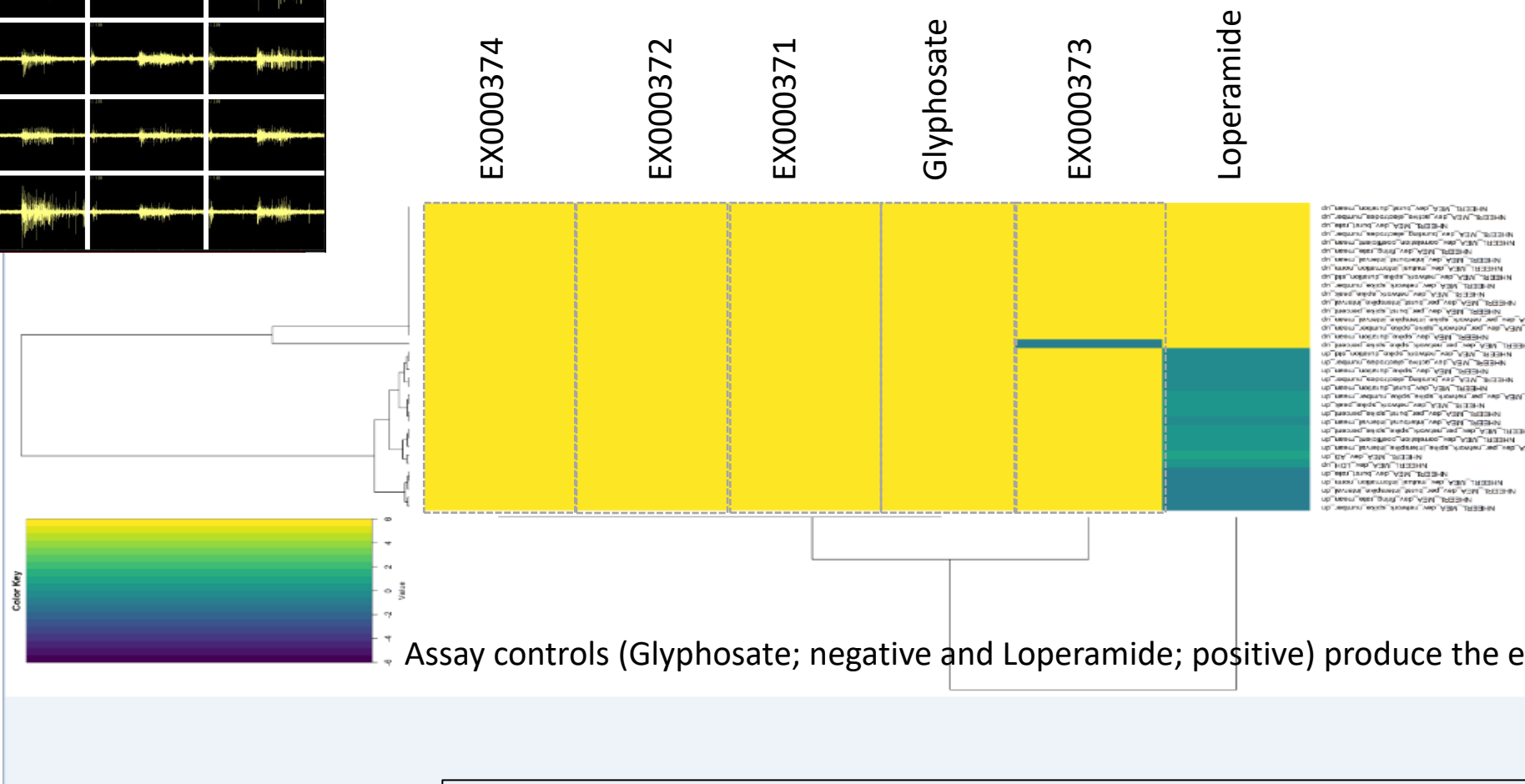
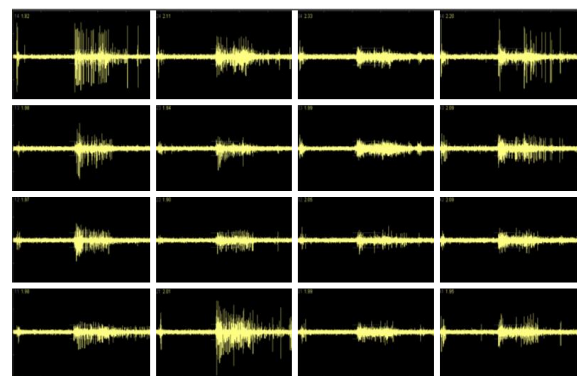


*EX000371 = Compound X

Conclusion: Compound X and analogs have no effects on neurite outgrowth

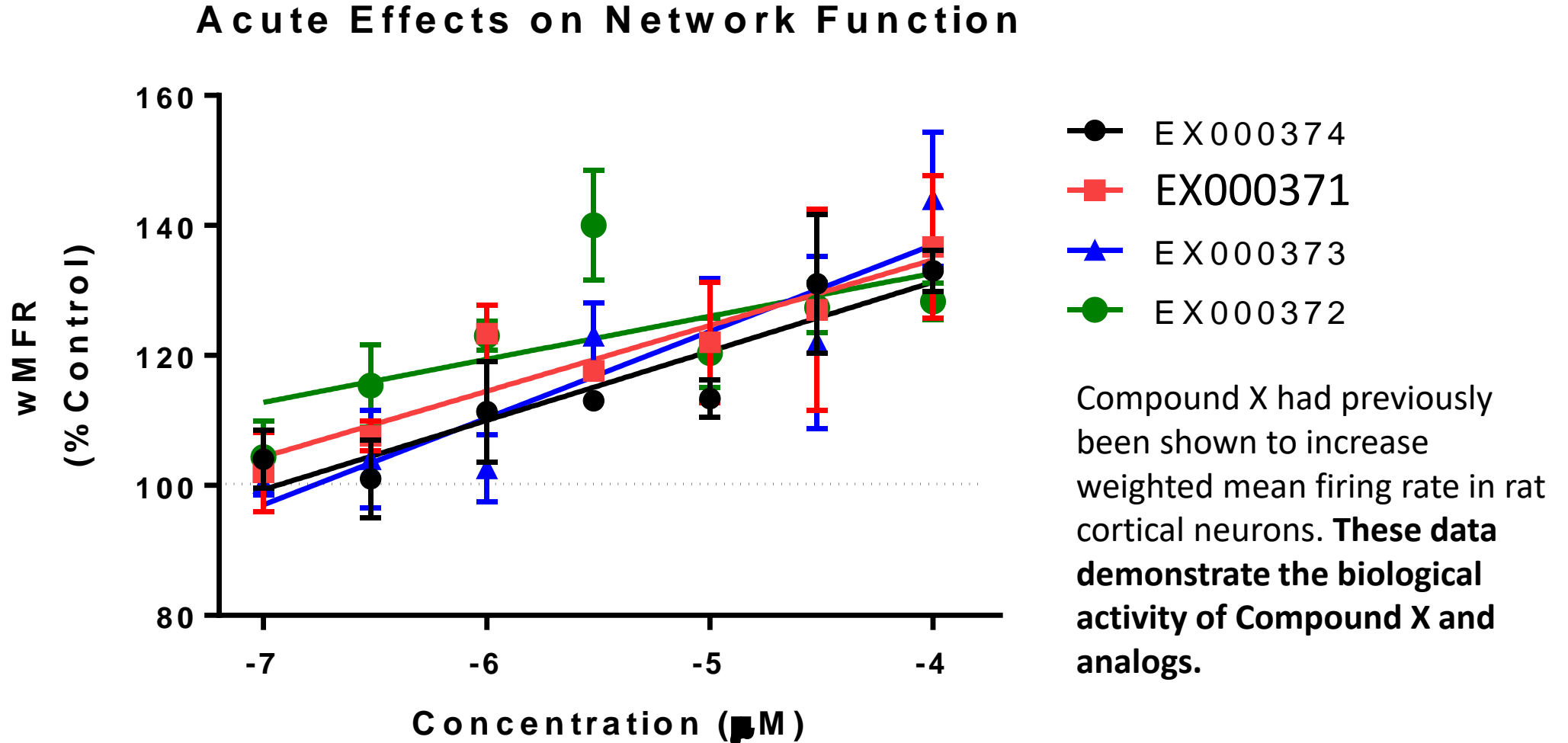


DNT NAMs data for Compound X and analogs on Network Formation



Conclusion: Compound X and analogs have no effects on Network Formation

Acute Effects of Compound X and analogs on Network Function





IVIVE Approaches indicate that appropriate concentration ranges were tested

From Guideline study, LOAEL of Compound X = **14 mg/kg/day**

Using HTTK and IVIVE

- 1 mg/kg/day = C_{ss} values of 0.66 and 2.21 μ M in rats and humans, respectively
- 30 μ M Compound X = AED of **45 mg/kg/day** (rats) and 13.5 mg/kg/day (humans)

Summary: At concentrations equivalent to or above the LOAEL from in vivo studies, Compound X and analogs did not alter Neurite Outgrowth or Network Formation, but did have acute effects on Network Function

Organophosphate insecticides are currently regulated based on inhibition of acetylcholinesterase (AChE).

Primary Questions:

- 1) Does the DNT battery indicate that regulation based on AChE inhibition may not be health protective?
- 2) Can data from the DNT battery contribute to a WOE approach for OPs?



Organophosphates and DNT

Study Design:

Test 27 Organophosphate insecticides in the EPA DNT assays
8 Parent/oxon pairs
Concentration-response up to 100 μM
Pipeline results through TCPL to generate AC_{50} values
Use HTKK to convert AC_{50} values to AED_{50} values
Compare to BMD/BMDL10 values based on AChE inhibition

Assays:

Proliferation	-	human neuroprogenitors (hNP1)
Apoptosis	-	human neuroprogenitors (hNP1)
Neurite initiation	-	human neurons (hN2)
Neurite initiation	-	rat primary neural culture
Neurite maturation	-	rat primary neural culture
Synaptogenesis	-	rat primary neural culture
Network formation (MEA)	-	rat primary neural culture
Behavior/Anatomy	-	zebrafish (data analysis pending)



In vitro assays to identify DNT Hazard: Promises and challenges

Promises:

- Data on DNT hazard for many more chemicals
- Characterization of DNT hazard on biologically-relevant processes
- Data from human models
- Substantially lower cost and faster results than *in vivo* studies

Challenges/Future Directions:

- Further evaluation of the battery
- Development of additional case-studies using *in vitro* DNT assays
- Development of additional AOPs related to DNT that will increase confidence in using these assays
- Development of assays that cover areas of neurodevelopmental processes not well covered in the current battery



Thank you! Questions?

EPA Colleagues:

- Kathleen Wallace
- Theresa Freudenrich
- Bill Mundy (retired)
- Josh Harrill
- Jasmine Brown
- Katie Paul Friedman
- Melissa Martin
- Kelly Carstens (ORISE)
- Amy Carpenter (ORISE)
- Seline Choo (ORISE)

EFSA Collaborators

- Ellen Fritsche
- Marcel Leist

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- Andrea Terron