

Machine Learning in Predictive Toxicology: An Overview and Case Study

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A frequent problem in toxicology....

- We have a large number of chemicals to screen for potential risk
- We want to know about hazard or toxicity that is hard to measure for all these chemicals (expensive, slow, unethical...)
 - We have some previously-measured examples of this information for some chemicals
- We have information that is easier to measure (rapid, inexpensive)
 - Molecular structure
 - In vitro bioactivity in high-throughput screening assays
- We don't have a clear idea of how the "hard" info relates to the "easy" info
 - no mechanistic model
- How can we use the available "easy" data to predict the "hard-to-measure" data?



Machine learning: Computational algorithms that can infer patterns from data

Data					
Target/ response	Features				
у1	x1	x2	•••		
[value1]	[value1]	[value1]			
[value2]	[value2]	[value2]			

Target/response (y): what we want to predict (e.g. toxicity or hazard)

Features (x1, x2...): Information available to predict response (e.g. structure, in vitro HTS bioactivity, etc.)



Two categories of patterns to be inferred

Supervised: Infer relationshipbetween target and features.Goal: predict target from features.

Data					
Target/ response	Features				
у1	x1	x2	•••		
[value1]	[value1]	[value1]			
[value2]	[value2]	[value2]			

Unsupervised: No target to predict. Infer descriptive patterns in features (e.g., clustering).

Data					
Features					
x1	x2				
[value1]	[value1]				
[value2]	[value2]	•••			



This presentation will focus on supervised machine learning

Supervised: Infer relationship between target and features. Goal: predict target from features.

Data					
Target/ response	Features				
у1	x1	x2			
[value1]	[value1]	[value1]			
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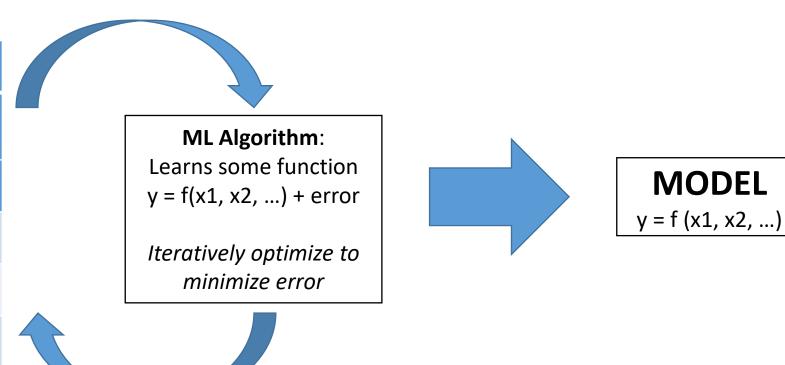
Unsupervised: No target to predict. Infer descriptive patterns in features (e.g., clustering).

Data					
Features					
x1	x2				
[value1]	[value1]	•••			
[value2]	[value2]	•••			



United States Environmental Protection Training a supervised ML model

Training data				
Target/ response	Features (predictors)			
у1	x1	x2		
[value1]	[value1]	[value1]		
[value2]	[value2]	[value2]	•••	



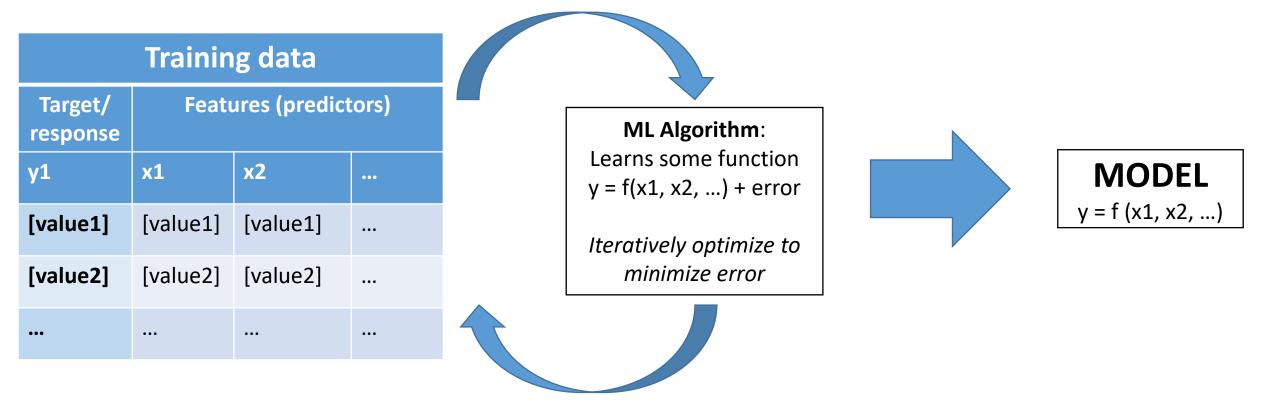
Algorithms can include: • naïve Bayes

- k nearest neighbors
- decision trees
- support vector machine

- random forest
- artificial neural networks
- etc.



Two types of model: regression and classification



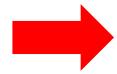
Regression model: y is numeric & continuous (e.g. LD50) **Classification model:** y is categorical (e.g. hepatic toxicity yes/no)



This presentation focuses on classification models

	Trainin	g data			
Target/ response	Featı	ures (predic	tors)	ML Algorithm:	
/1	x1	x2		Learns some function y = f(x1, x2,) + error	
alue1]	[value1]	[value1]		Iteratively optimize to	
alue2]	[value2]	[value2]		minimize error	
•					

Regression model: y is numeric & continuous (e.g. LD50)



Classification model: y is categorical (e.g. hepatic toxicity yes/no)



"*Classification model:* Usually predicts *probability* of positive/negative for a category (e.g., hepatotoxicity)

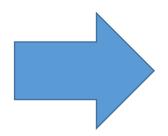
	Training da	ata
Target/	Features (pre

Target/ response	Features (predictors)				
y1: Positive?	x1	x2			
0	[value1]	[value1]			
1	[value2]	[value2]			



Learns some function y = f(x1, x2, ...) + error

Iteratively optimize to minimize error



MODEL

y = f(x1, x2, ...)

Predicted prob. positive

0.4

0.5

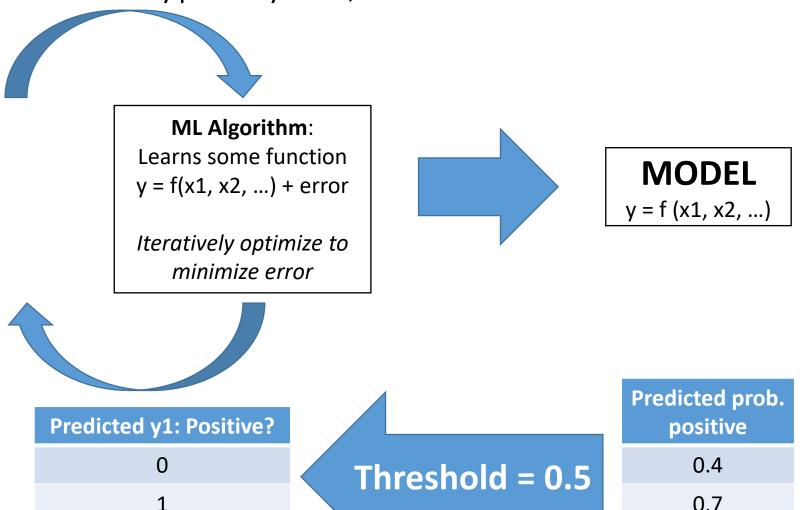
0.7

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Make predictions categorical by applying a threshold on predicted probabilities — typically 0.5, but doesn't have to be

Training data					
Target/ response	Features (predictors)				
y1: Positive?	x1	x2			
0	[value1]	[value1]			
1	[value2]	[value2]			
•••					



• • •



Evaluating performance of a ML model

	Test	data	
Target/	Features (predictors)		tors)
response	x1	x2	
•			•••
[value1]	[value1]	[value1]	•••
[value2]	[value2]	[value2]	
		Metric:	Error b

For regression, this could be sum of squared errors $\sum (y - y_p)^2$



Error metrics for (binary) classification models: confusion matrix

	Predicted negative	Predicted positive
Observed negative	True negatives (TN)	False positives (FP)
Observed positive	False negatives (FN)	True positives (TP)

Accuracy: (TN + TP) / (TN + TP + FN + FP)

Sensitivity (true positive rate, TPR): TP/(TP + FN)

Specificity (true negative rate, TNR): TN/(TN + FP)

Balanced Accuracy: (Sensitivity + Specificity)/2

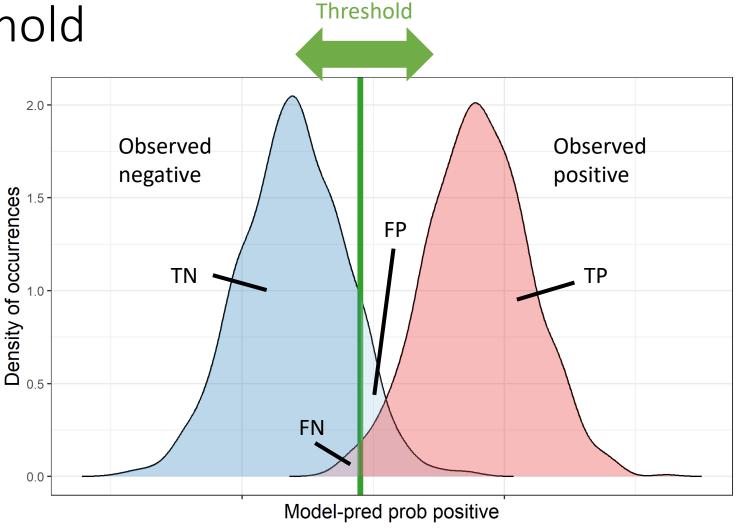
Positive Predictive Value (PPV): TP / (TP + FP)

False Discovery Rate: 1 – PPV

[...lots more!]



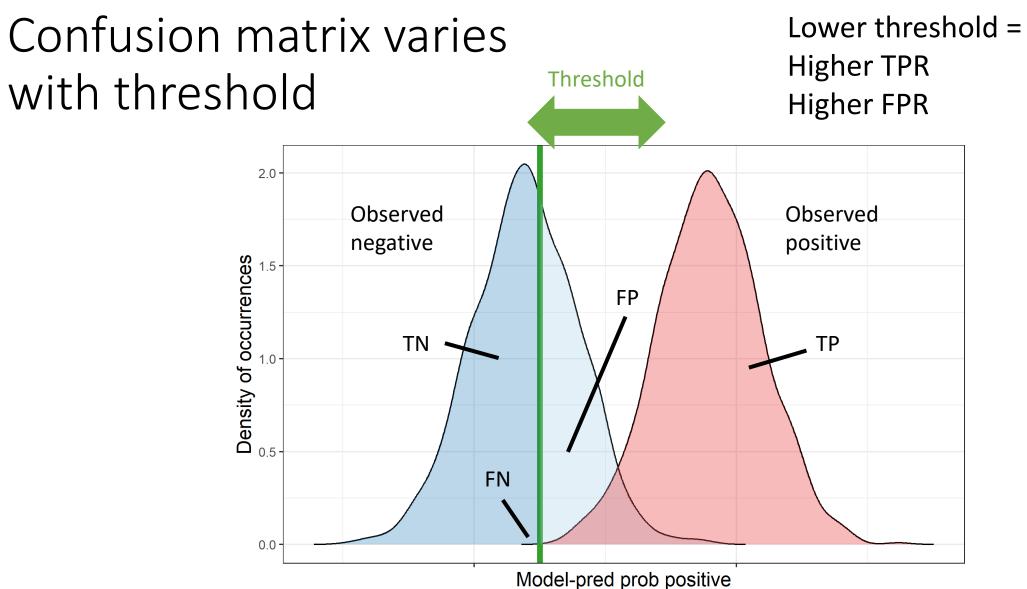
Confusion matrix varies with threshold



More separation between peaks = more informative model

(synthetic example data)

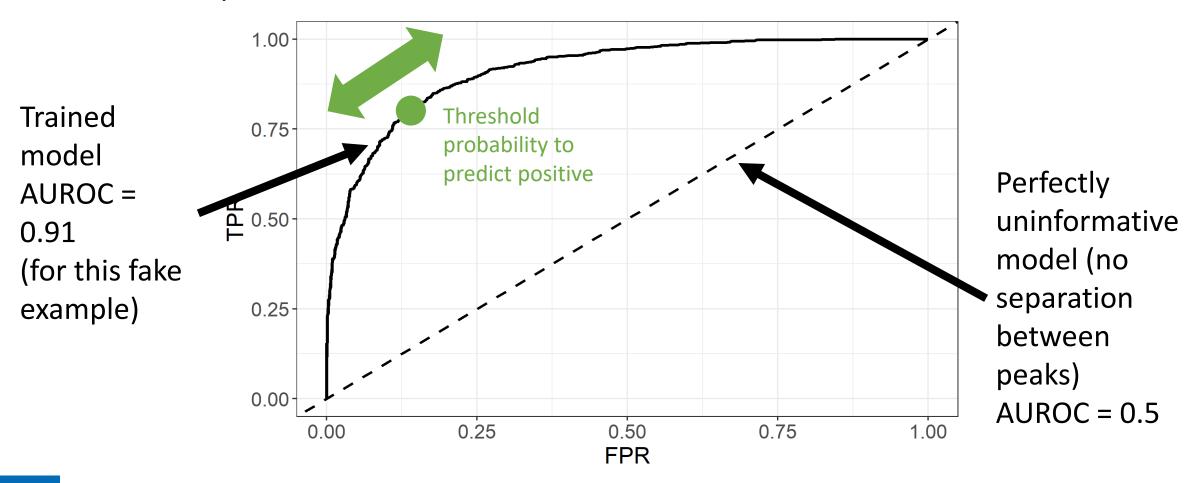






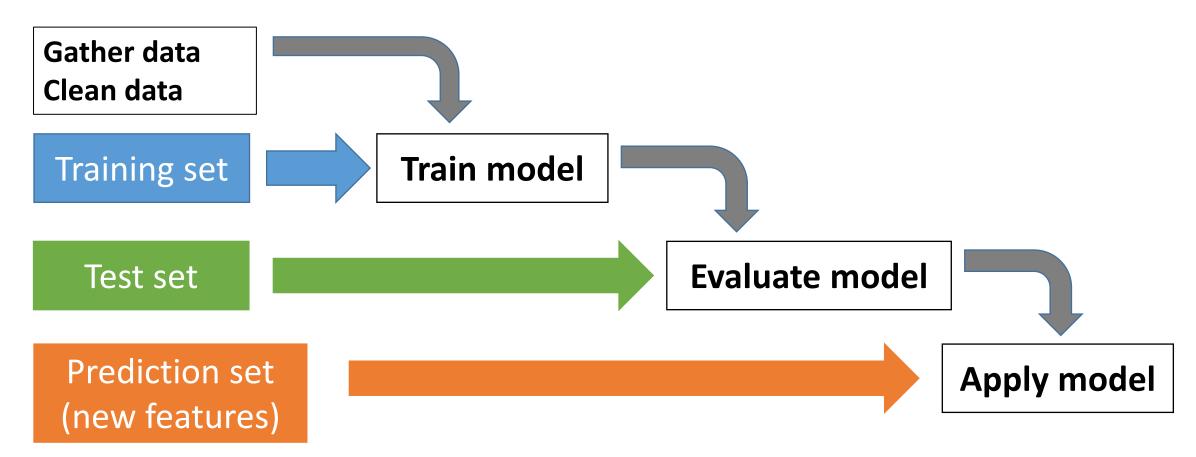
Confusion matrix varies Higher threshold = **Lower TPR** Threshold with threshold Lower FPR 2.0 Observed Observed negative positive Density of occurrences FP TN TP FN 0.0 Model-pred prob positive

Area under receiver-operator characteristic (ROC) curve (AUROC) tells us about separation between peaks & model performance over all thresholds





Summary of machine-learning model process





Challenge in classification models for predictive toxicology: *imbalanced data*

From Mansouri et al. 2020: ComPARA training set (response = *in vitro* androgen activity in ToxCast, yes/no)

Table 1. Training set chemicals for binding, agonist and antagonist data sets.

	-		
Number of	Binding	Agonist	Antagonist
Actives Inactives	198 1,464	43 1,616	159 1,366
Total	1,662	1,659	1,525
	88% inactive for binding	97% inactive for agonism	90% inactive for antagonism

Problem:

A ML model that simply predicted "inactive" for *everything* would have a 97% accuracy rate for agonism!

Many toxicology-related data sets are imbalanced like this (Idakwo et al. 2018; Wang et al. 2020)

How can we build a ML model that properly predicts the minority class?



Strategies to address imbalanced training data

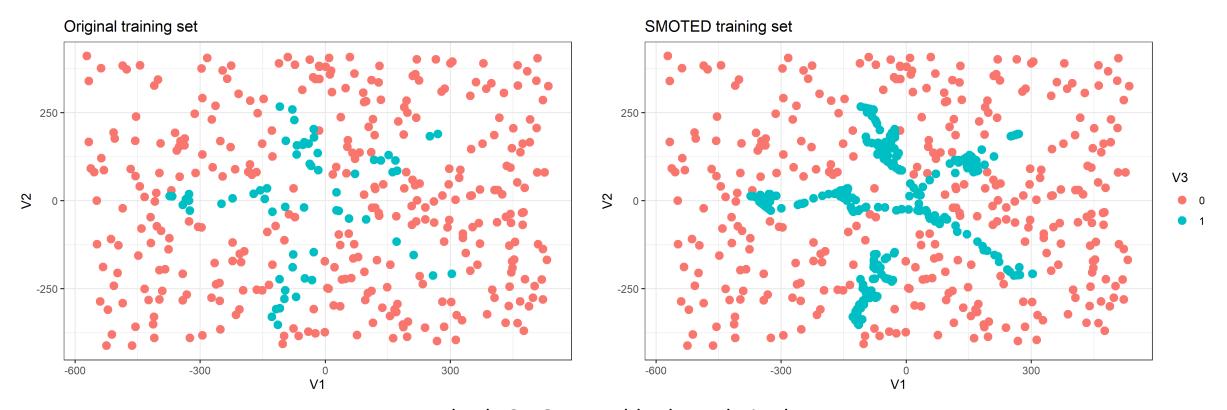
(Branco et al., 2016)

- Algorithm-based: Make the model less sensitive to imbalance
 - Boosting: iteratively correct misidentified instances in the training class
 - Bagging: trains multiple versions of the model on subsets or bootstrap-resampled versions of the data set
 - Cost function: During model training, weight errors more heavily for minority-class examples
- Sampling-based: Pre-process training data to balance out the classes
 - *Undersampling*: Remove some majority-class examples
 - Oversampling: Repeat some minority-class examples or create synthetic new ones
- Synthesizing new minority-class examples
 - Generative Adversarial Networks (GAN): train a second ML model to synthesize plausible minority examples (Douzas & Bacao, 2018; Green et al. 2021)
 - Interpolate between minority-class examples & nearest neighbors: e.g. SMOTE (Synthetic Minority Over-sampling TEchnique) (Chawla et al., 2002)



Example of SMOTE

"clover" data from https://sci2s.ugr.es/keel/datasets.php (Alcalá-Fdez et al. 2011)



Drawback: SMOTE can blur boundaries by interpolating to majority-class near neighbors



It turns out that ML algorithms and imbalanced data strategies perform very differently for different data sets



Figure 9A from Wang et al. 2020 (literature review of ML methods in predictive toxicology)

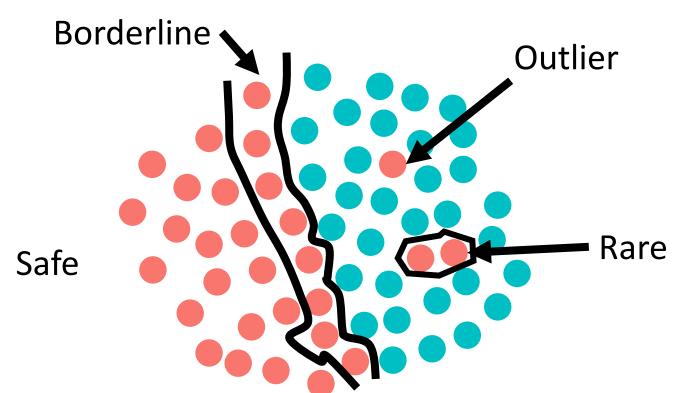
Variability in model performance *not* explained by training dataset size or by ML algorithm used

Authors suspect dataset-specific difficulties



"Data difficulty factors": ML performance depends on frequency of 4 different types of data points

(Napierla & Stefanowski 2015; Garcia et al. 2020; Stefanowski 2016)



Napierla & Stefanowski 2015:

Undersampling seems to work better for borderline examples

SMOTE seems to work better for outlier & rare examples

Adapted from Garcia et al. (2020)



Suggestion: Develop a more systematic approach to characterize these "data difficulty factors" in predictive toxicology datasets

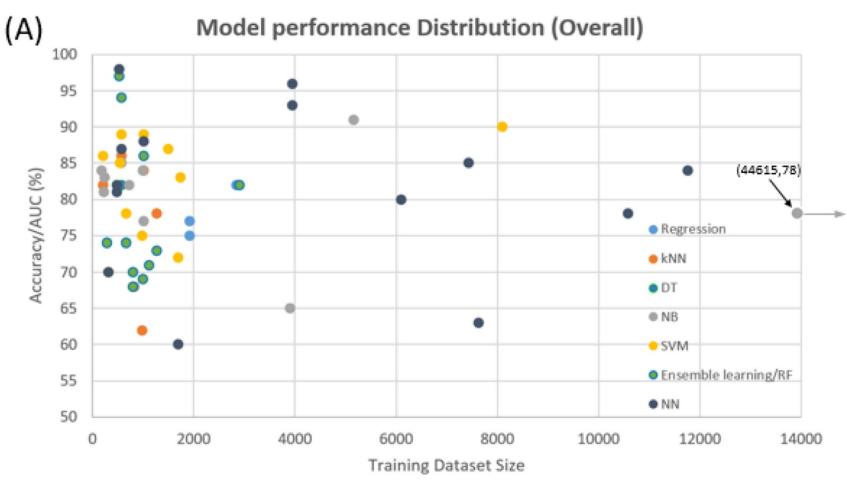


Figure 9A from Wang et al. 2020 (literature review of ML methods in predictive toxicology)

by proportion of safe, borderline, rare, and outlier data?

Could we identify "best practices" based on these dataset characteristics?

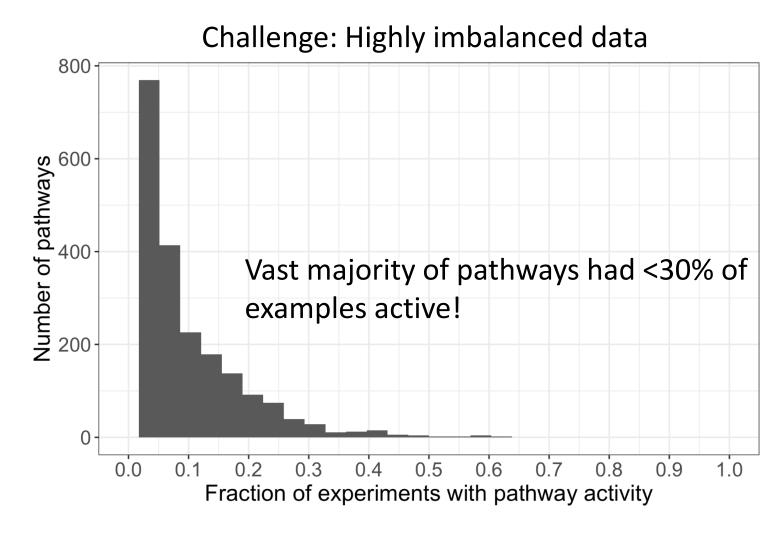


Case study: Machine learning for *in vitro-in vivo* extrapolation (Ring, Rager, et al. 2021)

 Target: in vivo pathway-level transcriptomic activity (active/inactive) in rat liver, for a given chemical & dose (DrugMatrix and TG-Gates datasets)

Features:

- in vitro Tox21 bioactive concentrations (AC50) for 144 assays
- phys-chem properties
- in vivo dose
- toxicokinetic model predictions of plasma & liver concentration at in vivo dose

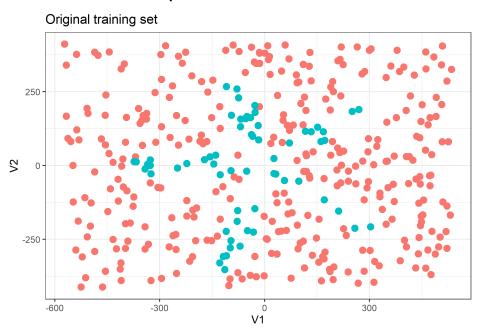


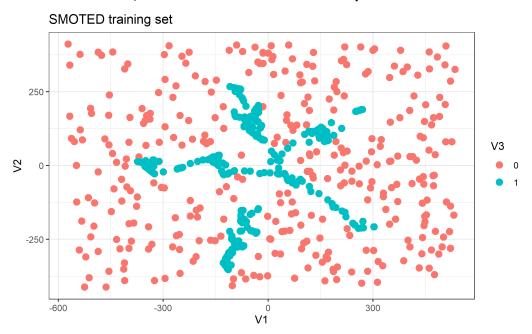


Approach to imbalanced data: SMOTE (Ring, Rager, et

al. 2021)

(Illustration of SMOTE on example data from earlier, **not** our actual data)



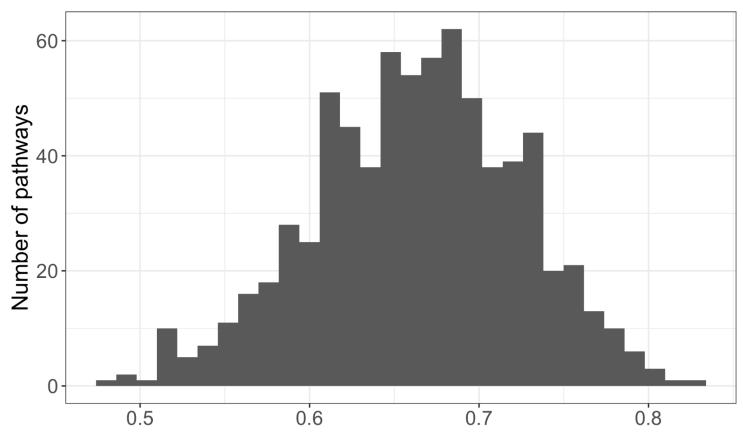


- High-dimensional feature set: in vitro bioactivity for 144 Tox21 assays interpolating along all of these dimensions
- We did not evaluate "data difficulty factors" in this analysis, so we don't know about safe, borderline, rare, or outliers



Result: Pathway models with decent AUROC

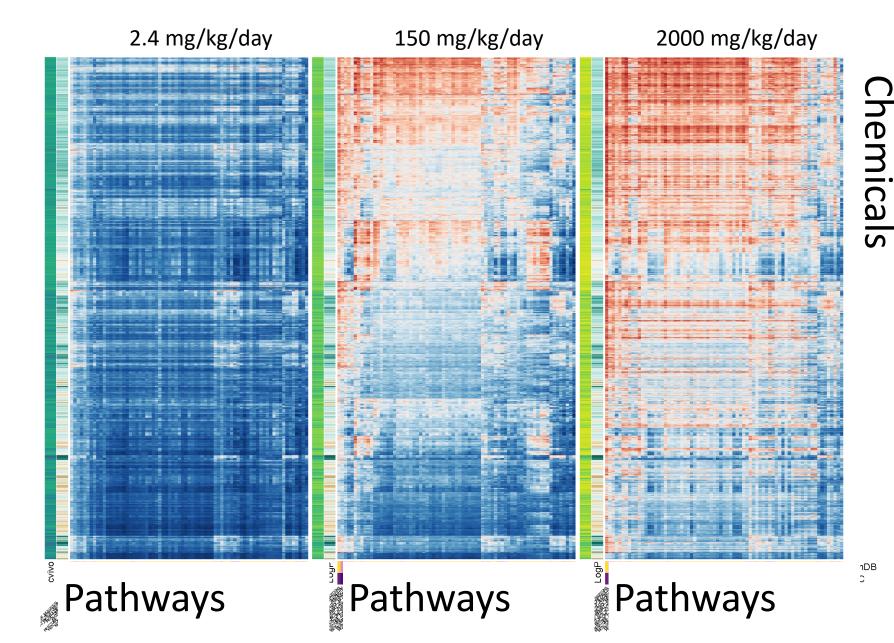
(Ring, Rager, et al. 2021)



AUROC of trained ML model for each pathway

al. 2021)

Result: Apply models to predict pathway activity for 6617 Tox21 chemicals at three doses spanning DrugMatrix dose range (Ring, Rager, et





Summary

- Machine learning is a powerful tool for predictive toxicology ...
- ... But its performance is affected by data difficulty factors
 - Imbalanced data
 - Safe, borderline, rare, and outlier data points
- Strategies to address imbalanced data exist & are fairly successful
 - e.g. SMOTE, GAN
 - but data difficulty factors affect these strategies as well
- Suggestion: Develop a more systematic approach to characterizing data difficulty factors for predictive toxicology datasets
- Case study: Machine learning for in vitro-in vivo extrapolation
 - Applying SMOTE to address highly imbalanced training data



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