

# Category-Based Toxicokinetic Evaluations of Data-Poor Per- and Polyfluoroalkyl Substances (PFAS) by Gas Chromatography Coupled with Mass Spectrometry

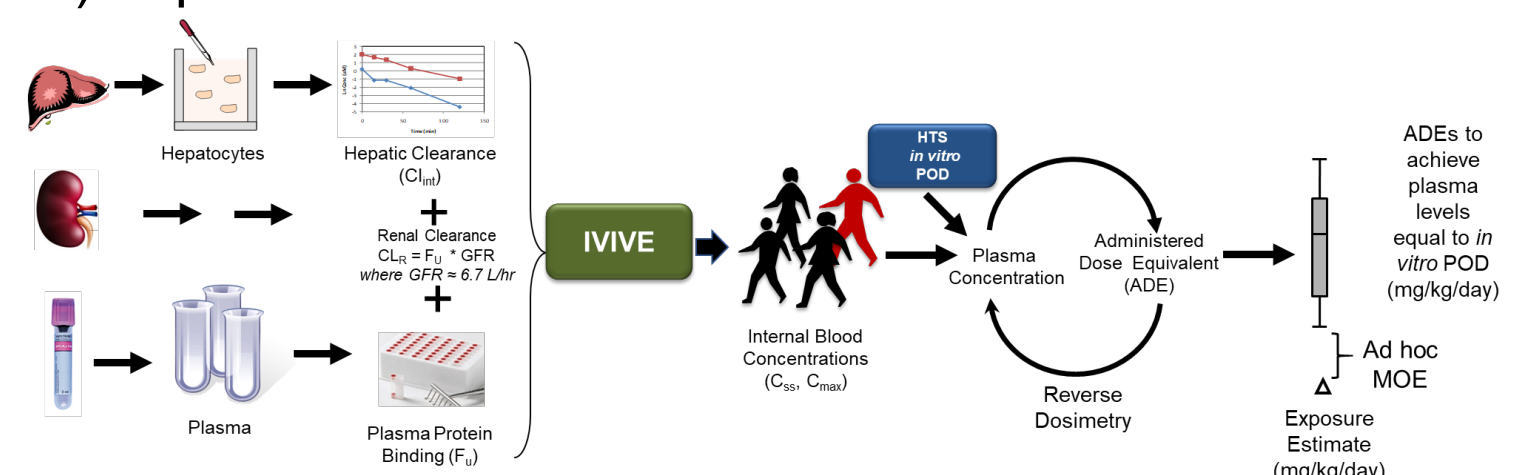
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## Background

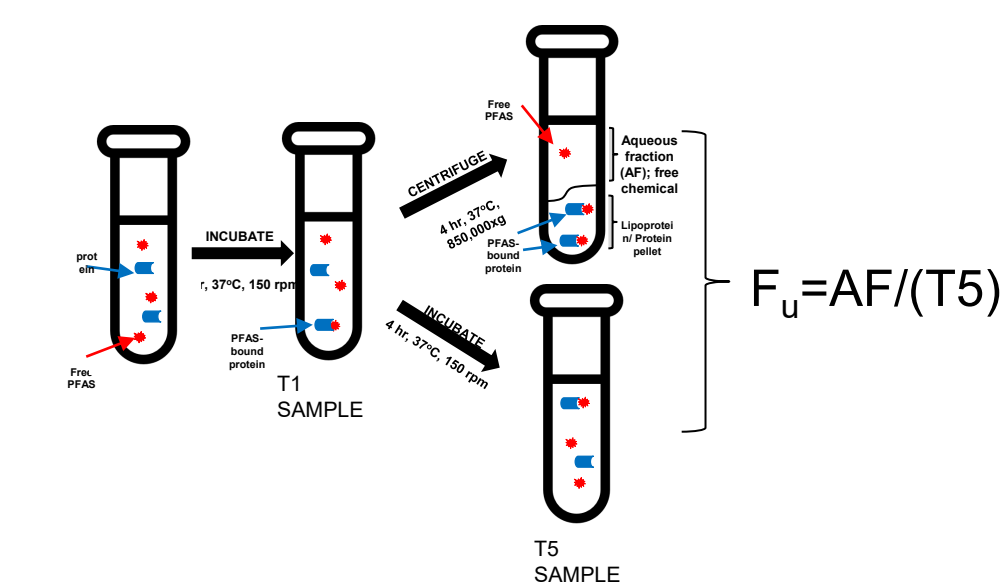
- Despite widespread concern over PFAS exposure, only limited biomonitoring, toxicity and toxicokinetic (TK) data are available.
- New approach methods (NAMs) that make use of *in vitro* and *in silico* methodologies are available to provide toxicity and TK data efficiently.
- Read-across approaches that use such data to categorize PFAS in specific groupings can be used to inform risk across the broader set of PFAS.
- This poster describes:
  - Evaluation and generation of *in vitro* TK data (plasma protein binding (PPB), hepatic clearance) of a set of 75 PFAS by gas chromatography mass-spectrometry.
  - Trends analyses to aid in developing grouping strategies
  - Preliminary findings with regard to *in vitro-in vivo* extrapolation (IVIVE) to predict *in vivo* TK.



TK NAM approach using *in vitro* TK data to predict *in vivo* TK. Using TK NAMs, *in vitro* points of departure (POD) can be converted to an administered equivalent dose (AED), useful in margin of exposure (MOE) evaluations employed during risk assessment.

## In Vitro Toxicokinetic Assays

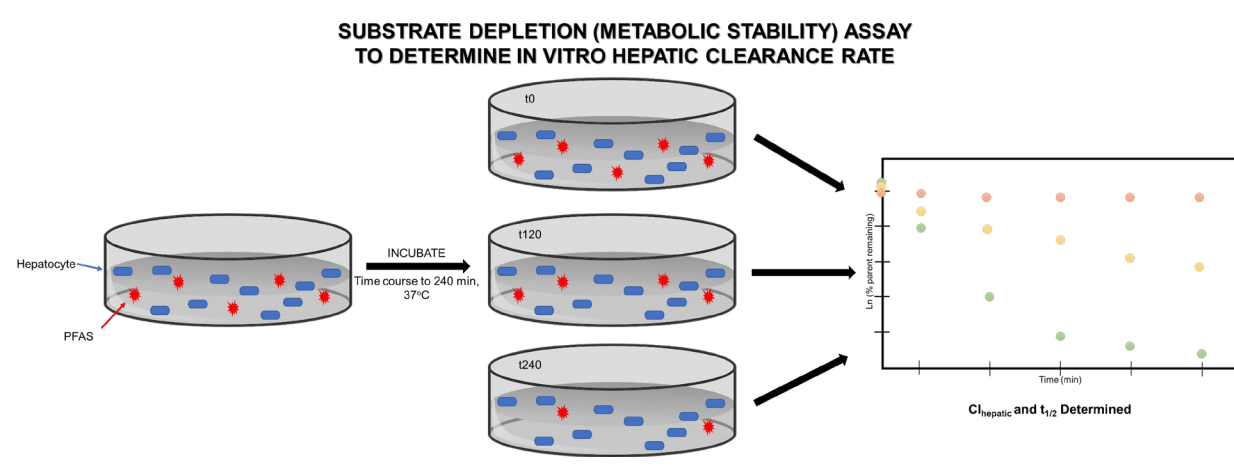
### ULTRACENTRIFUGATION PLASMA PROTEIN BINDING ASSAY



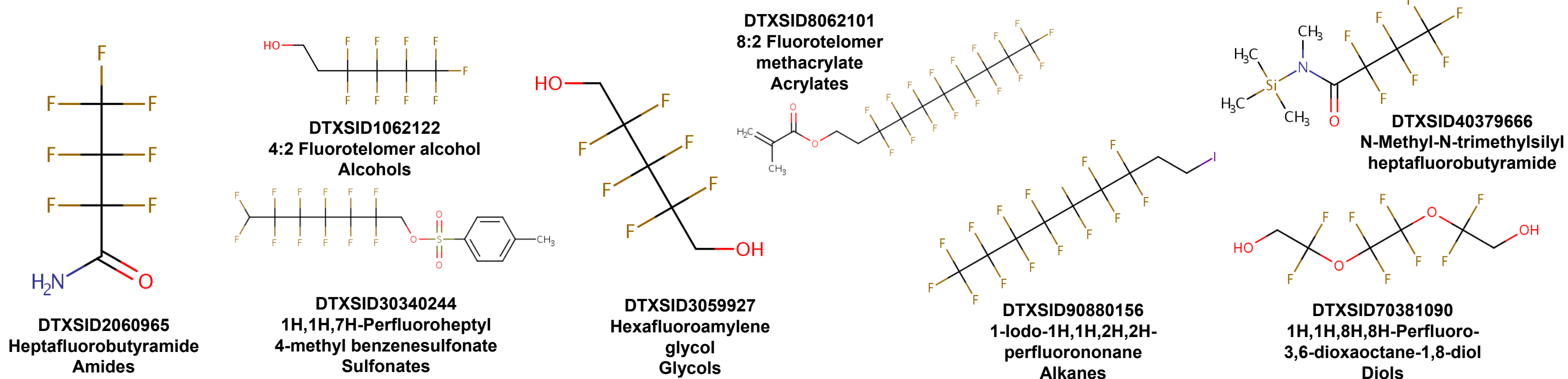
- Human plasma (10-donor pool, mixed sex) centrifuged at 850,000xg to separate aqueous fraction from plasma proteins, lipoproteins, and fatty acids
- Fraction unbound ( $f_u$ ) derived by analyte quantitation in aqueous fraction (AF) and whole plasma, performed by gas chromatography-tandem mass spectrometry (GC-MS/MS) on Agilent 8890/7010B, 7890A/7010A, and 7890B/7010B instrumentation

### SUBSTRATE DEPLETION (METABOLIC STABILITY) ASSAY TO DETERMINE IN VITRO HEPATIC CLEARANCE RATE

- Substrate depletion approach using primary human hepatocytes (50-donor pool, mixed sex) at 1  $\mu$ M PFAS.
- Time course: 0, 15, 30, 60, 90, 120, and 240 min with linear regression fit to determine intrinsic clearance ( $CL_{int}$ )
- Media only and heat-inactivated hepatocyte controls to monitor abiotic loss



## PFAS Chemical Space Evaluated



75 PFAS containing functional groups displayed above were selected for evaluation. All DMSO stocks passed an analytical QC check prior to study start. Targeted mass spectrometric methods have been developed for 62 to date. PPB, hepatic clearance, and associated stability assays have been conducted; TK data generated to date is discussed below.

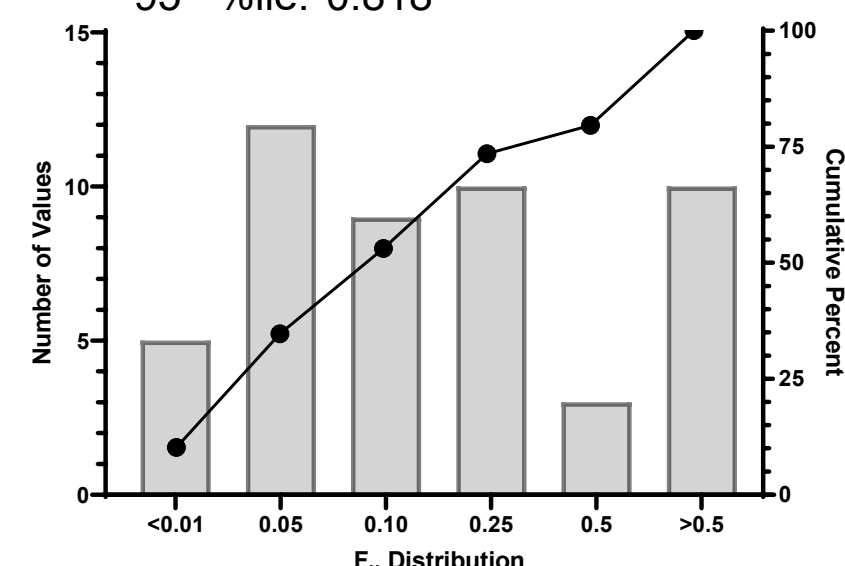
## Results

### Category-Based Evaluations of PFAS Plasma Protein Binding (PPB)

#### Distribution of $f_u$ values for 48 GC-MS PFAS

##### $f_u$ Summary Statistics

25<sup>th</sup> %ile: 0.028  
Median: 0.098  
Mean: 0.218  
75<sup>th</sup> %ile: 0.248  
95<sup>th</sup> %ile: 0.818

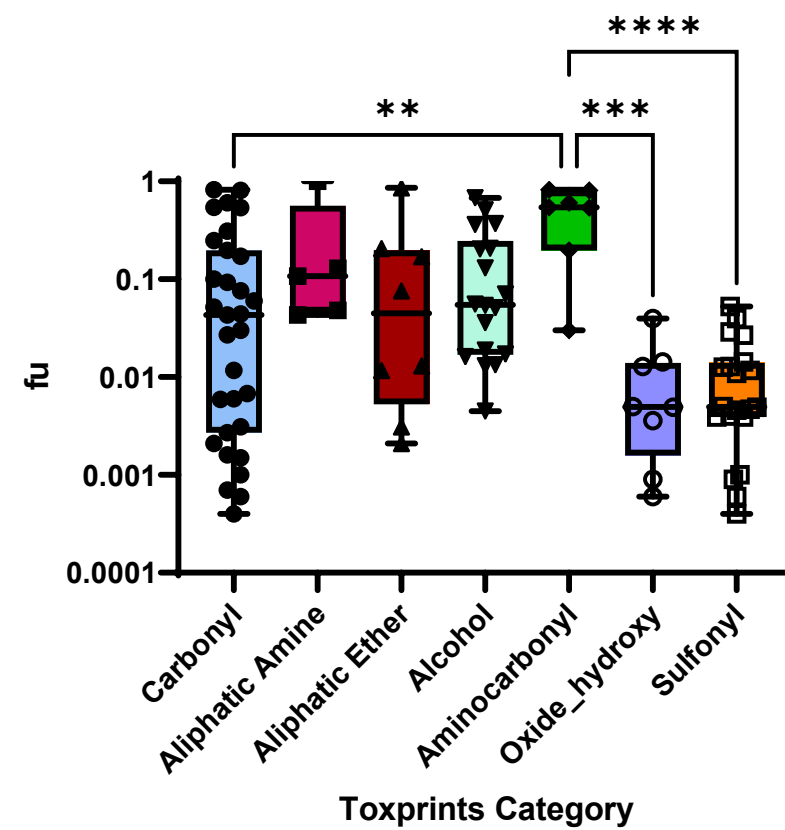


#### Distribution of $f_u$ values measured for 48 GC-MS PFAS.

A parallel  $f_u$  study on 58 LC-MS PFAS showed a median  $f_u$  = 0.01; i.e., 10-fold higher binding compared to this set (75<sup>th</sup> %ile:  $f_u$  < 0.05; LCMS PFAS include PFOA, PFOS, PFHxS).

See Smeltz et al., poster 3093; P196

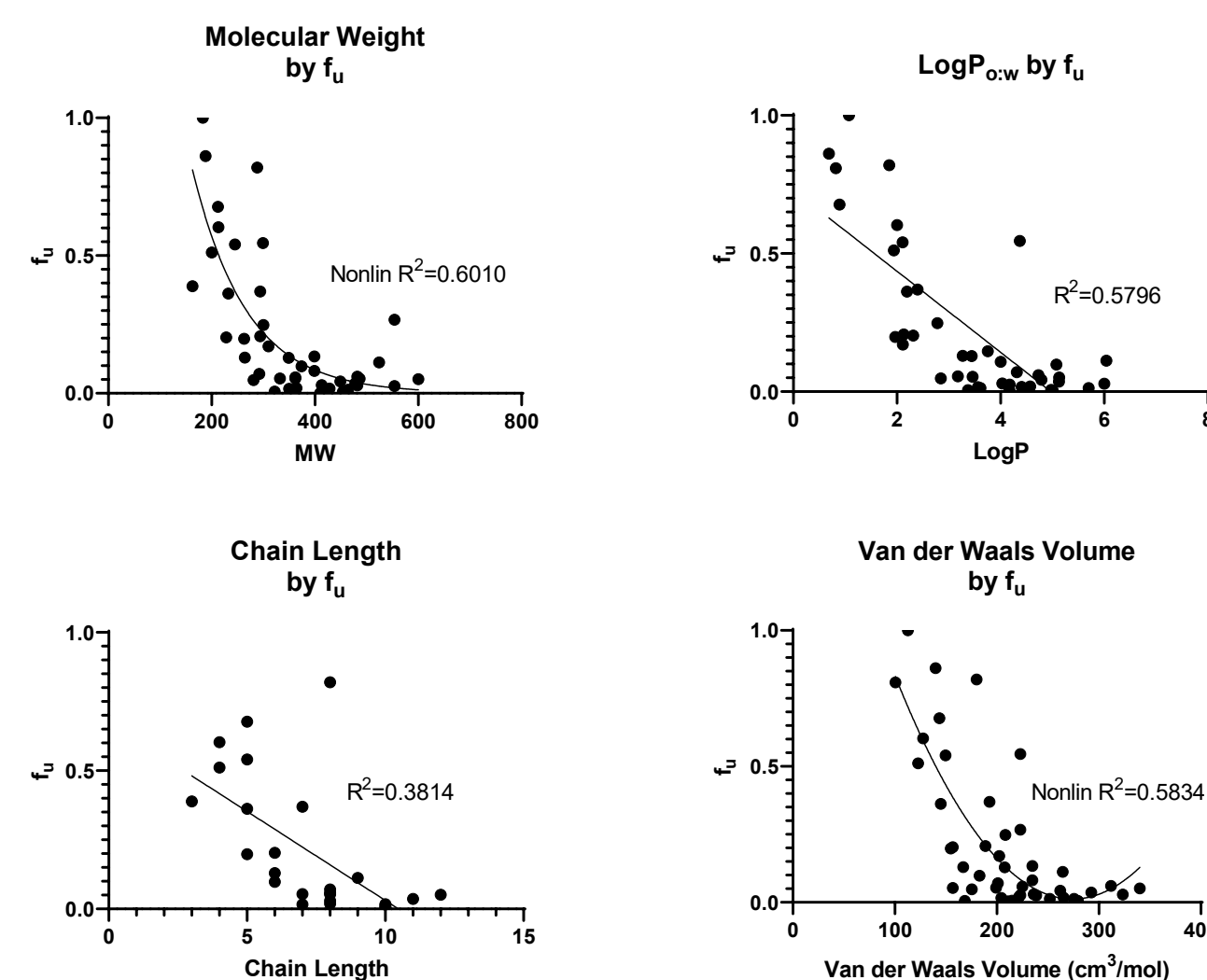
#### Grouping $f_u$ based on ToxPrint ChemoTypes



**Using ToxPrint ChemoTypes to evaluate  $f_u$ .** A one-way ANOVA was performed on the major ToxPrint chemotypes using a Tukey test for multiple comparisons to determine between-group differences. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001.

Note the ToxPrint ChemoType names are condensed for display purposes.

#### Identifying trends between $f_u$ and physicochemical properties



**Physicochemical properties trends analysis.** Plots depict physicochemical properties that showed a significant correlation with  $f_u$ . Regression analyses were fit with simple linear, or non-linear regression, using whichever better fit the data. Chemicals were manually categorized for chain length and only included straightforward cases.

## Results: IVIVE Modeling

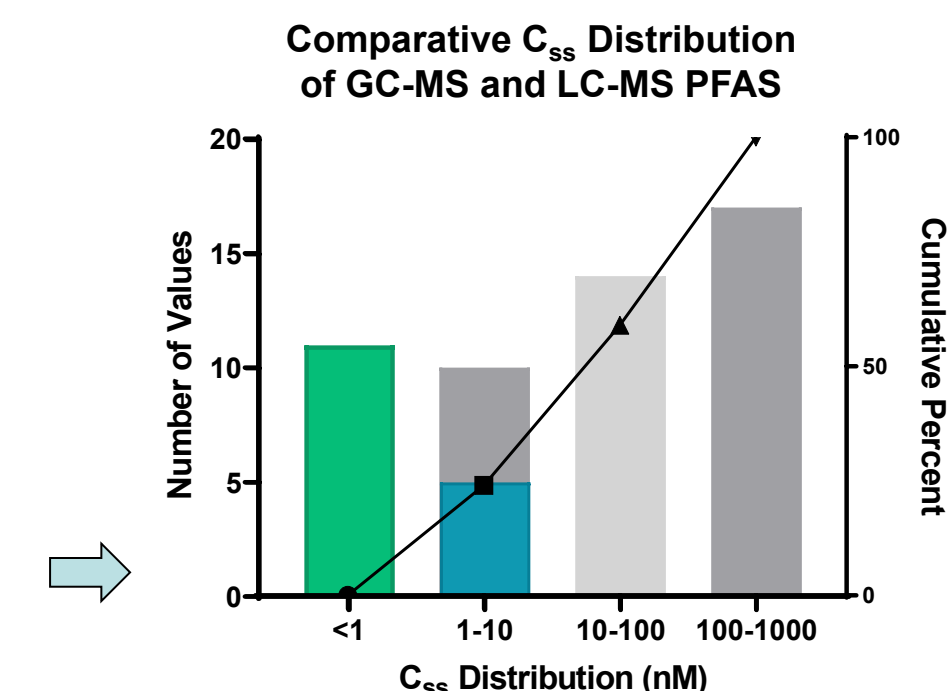
| DTXSID         | Compound Name                                     | Avg MW (g/mol) | $f_u$  | in vitro $CL_{int}$ ( $\mu$ L/min/ $10^6$ cells) | $CL_{renal}$ (L/h) | $CL_{hep}$ (L/h) | $C_{ss}$ (nM) |
|----------------|---|----------------|--------|--|--------------------|------------------|---------------|
| DTXSID0059871  | Pentafluoropropionamide                           | 163.047        | 0.809  | 0  | 5.4203             | 0.000            | 3.33          |
| DTXSID3059927  | Hexafluoroamylene                                 | 212.091        | 0.677  | 0  | 4.5359             | 0.000            | 3.06          |
| DTXSID50369896 | 1H,1H,10H,10H-Perfluorodecane-1,10-diol           | 462.13         | 0.004  | 56.6   | 0.0268             | 2.323            | 2.71          |
| DTXSID00380798 | 1H,1H,11H,11H-Perfluorotetraethylene glycol       | 410.112        | 0.013  | 19.93  | 0.0871             | 2.649            | 2.62          |
| DTXSID60379269 | 3-(Perfluoropropyl)propanol                       | 228.11         | 0.203  | 2.69   | 1.3601             | 5.407            | 1.90          |
| DTXSID30340244 | 1H,1H,7H-Perfluoroheptyl 4-methylbenzenesulfonate | 486.27         | 0.029  | 23.85  | 0.1943             | 6.740            | 0.87          |
| DTXSID90880156 | 1-Iodo-1H,1H,2H,2H-perfluorononane                | 524.012        | 0.1123 | 5.75   | 0.7524             | 6.324            | 0.79          |
| DTXSID30396867 | 1H,1H,8H,8H-Perfluorooctane-1,8-diol              | 362.115        | 0.058  | 20.006   | 0.3886             | 10.761           | 0.73          |
| DTXSID70381090 | 1H,1H,8H,8H-Perfluoro-3,6-dioxaoctane-1,8-diol    | 294.097        | 0.207  | 6.72   | 1.3869             | 12.601           | 0.71          |
| DTXSID10382147 | 3-(Perfluoro-2-butyl)propane-1,2-diol             | 294.117        | 0.37   | 5.88   | 2.4790             | 18.266           | 0.48          |
| DTXSID80310730 | Octafluoroadipamide                               | 288.097        | 0.82   | 2.95   | 5.4940             | 19.858           | 0.40          |
| DTXSID1062122  | 4:2 fluorotelomer alcohol                         | 264.091        | 0.1300 | 37.71  | 0.8703             | 32.796           | 0.33          |
| DTXSID60400587 | Nonafluoropentamide                               | 263.063        | 0.198  | 26.026   | 1.3266             | 33.960           | 0.32          |
| DTXSID70366226 | Perfluoropentamide                                | 245.072        | 0.54   | 14.68  | 3.6180             | 43.315           | 0.26          |
| DTXSID2060965  | Heptafluorobutyramide                             | 213.055        | 0.603  | 19.95  | 4.0401             | 52.624           | 0.24          |
| DTXSID40379666 | N-Methyl-N-trimethylsilylheptafluorobutyramide    | 299.264        | 0.545  | 19.95  | 3.6515             | 50.397           | 0.18          |

$C_{ss}$  values calculated for 16 PFAS for which both  $f_u$  and  $CL_{int}$  values were available.  $CL_{renal}$  =  $GFR \times f_u$ .

#### Key Findings Compared to LC-MS PFAS, this set of 16 had much lower $C_{ss}$ values, due to:

- higher  $f_u$  values (on average 10-fold higher);
- higher hepatic clearance rates (only 2 with no  $CL_{int}$ ; for LC-MS PFAS, most <1  $\mu$ L/min/ $10^6$  cells)
- Highest  $C_{ss}$  for GC-MS PFAS 3.33 nM

**Distribution of steady-state concentration ( $C_{ss}$ ) values for GC-MS PFAS in comparison to LC-MS PFAS.** The number of GC-MS PFAS (shown in color) are overlaid on top of the distribution of LC-MS PFAS (gray), which show much higher  $C_{ss}$ .



## Summary

- PPB data on 48 GC-MS PFAS measured to date indicate lower binding than LC-MS PFAS, with  $f_u$  values ranging from 0.004 -1, and a mean  $f_u$  of 0.22.
- Evaluation of PFAS PPB using ToxPrint ChemoTypes showed variations based on presence of certain functional groups. Lower  $f_u$  was noted for sulfonyl/sulfur-containing PFAS and oxide hydroxy-containing PFAS. Higher  $f_u$  was noted for amino-carbonyl and nitrogen-containing PFAS.
- Correlations between increased PPB and higher molecular weight, chain length, log  $P_{ow}$ , and van der Waals volumes were noted during trends analyses.
- Due to lower PPB and higher hepatic clearance, these PFAS are less bioaccumulative than legacy PFAS with carboxylic acid and sulfonate functional groups (e.g., PFOA, PFOS).
- Given the relatively high rate of metabolic clearance for this set, ongoing work is now evaluating PFAS metabolic biotransformation through non-targeted and other analyses.
- Data generation across additional PFAS and toxicokinetic assays evaluating renal reuptake (i.e., *in vitro* transporter assays) are currently underway.

## References

Smeltz, M., Clifton, M., McMillan, L., Wetmore, B. Application of a Quality Scoring System for Assessing Per- and Polyfluoroalkyl Substances (PFAS) in Organic Solvents for *In Vitro* Toxicokinetic Testing. 2022 SOT Annual Meeting, PFASI Poster Session, #3093, P196.