The views expressed in this presentation are those of the author and do not necessarily reflect the views or policies of the US EPA

A mesocosm approach to test eDNA monitoring of ballast tanks

Courtney Larson, Donn Branstrator, Megan Corum, Meagan N Aliff, Abigail Latanich, Erik Pilgrim, Chelsea Hatzenbuhler, Matthew Julius, Matthew Etterson, and Euan Reavie





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A mesocosm approach to test <u>traditional</u> morphological and eDNA monitoring of Spiny Water Flea in ballast tanks

Courtney Larson, Donn Branstrator, Megan Corum, Meagan N Aliff, Abigail Latanich, Erik Pilgrim, Chelsea Hatzenbuhler, Matthew Julius, Matthew Etterson, and Euan Reavie

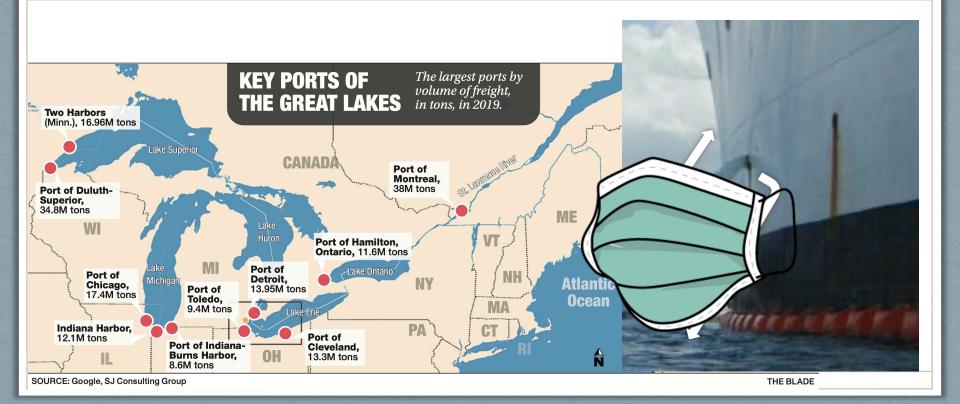




Ballast water

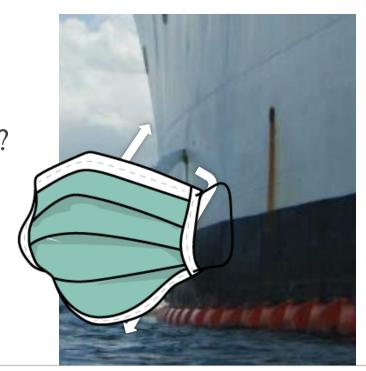


Ballast treatment and monitoring



Question: How clean is clean?

- What is the risk-release relationship for potential aquatic invaders?
- How can potential aquatic invaders be detected in ballast and ballast discharge?
 - Identify individuals morphologically
 - Adults
 - Larvae
 - Eggs
 - Identify populations genetically (eDNA)



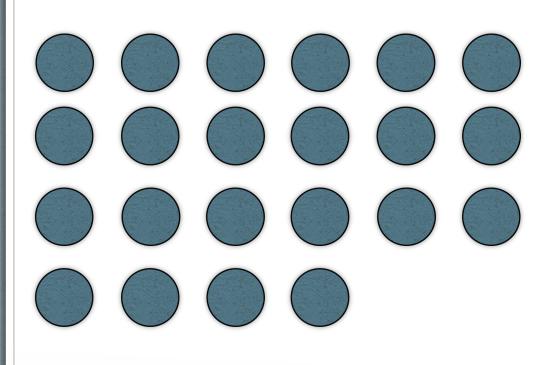
Study organism: *Bythotrephes cederströmii/longimanus*Spiny Water Flea



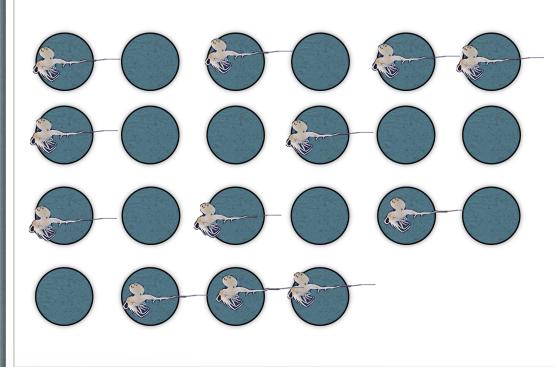




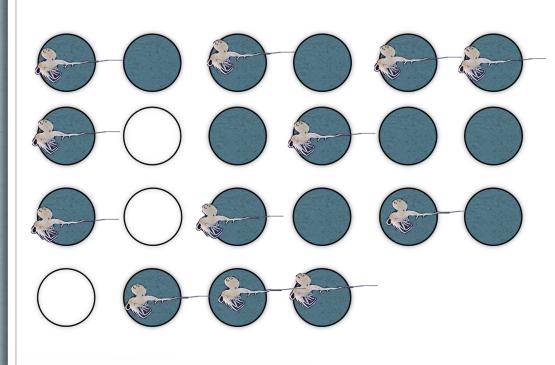
• 22, ~1 m³ mesocosm filled with harbor water



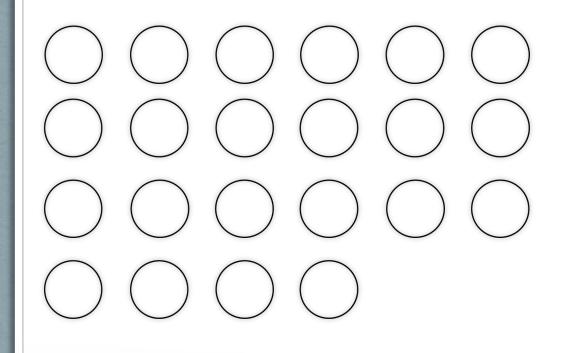
• 22, ~1 m³ mesocosm filled with harbor water



- 22, 1 m³ mesocosm filled with harbor water
- Introduce spiny water flea (SWF)



- 22, 1 m³ mesocosm filled with harbor water
- Introduce spiny water flea (SWF)
- Drain subsets over time



- 22, 1 m³ mesocosm filled with harbor water
- Introduce spiny water flea (SWF)
- Drain subsets over time
- Examine population at draining

Phase 1

If we introduce spiny water flea around the International Maritime Organization standard of 10 organisms per m³, what happens? Biol Invasions (2019) 21:3655–3670 https://doi.org/10.1007/s10530-019-02077-8

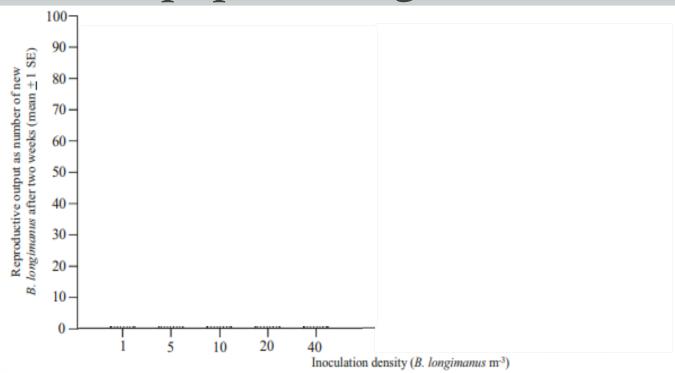
ORIGINAL PAPER

Evaluation of a method that uses one cubic meter mesocosms to elucidate a relationship between inoculation density and establishment probability for the nonindigenous, invasive zooplankter, *Bythotrephes longimanus*

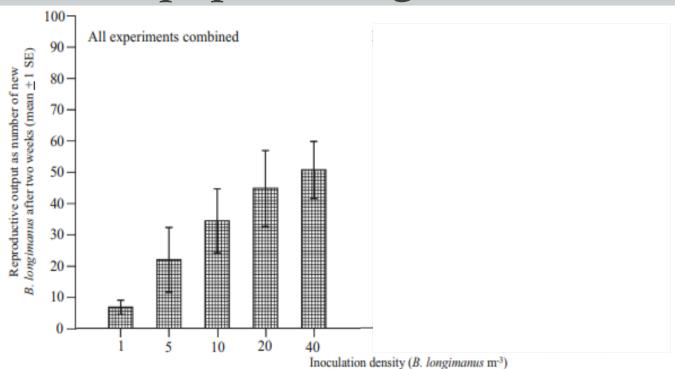
Donn K. Branstrator • · Matthew C. TenEyck · Matthew A. Etterson · Euan D. Reavie · Allegra A. Cangelosi

(

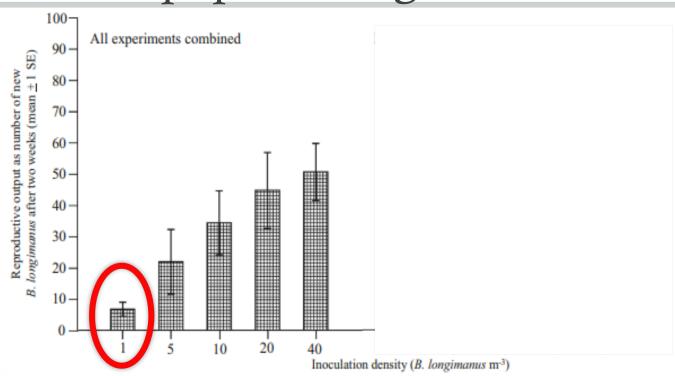
How does inoculation density affect population growth?



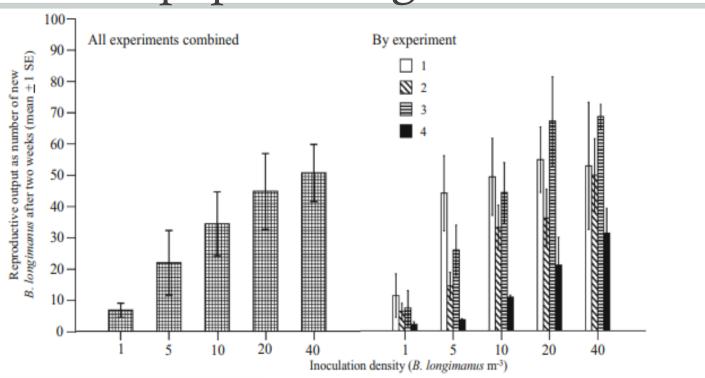
How does inoculation density affect population growth?



How does inoculation density affect population growth?



How does background conditions affect population growth?



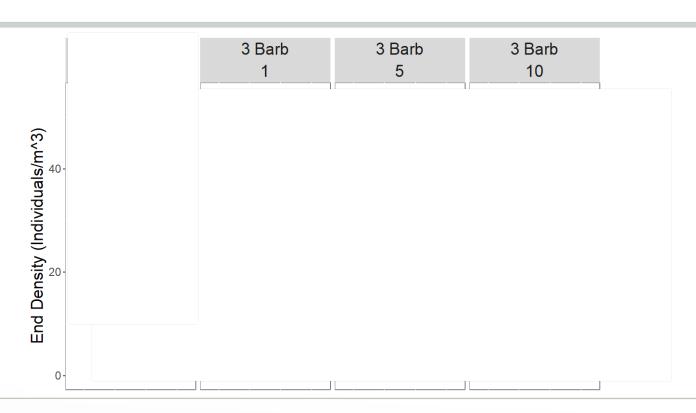
Phase 1 Conclusions

- Population growth occurs with even 1 inoculate SWF at 3-barb life stage
- Background conditions affect overall growth, but same trends for each experiment

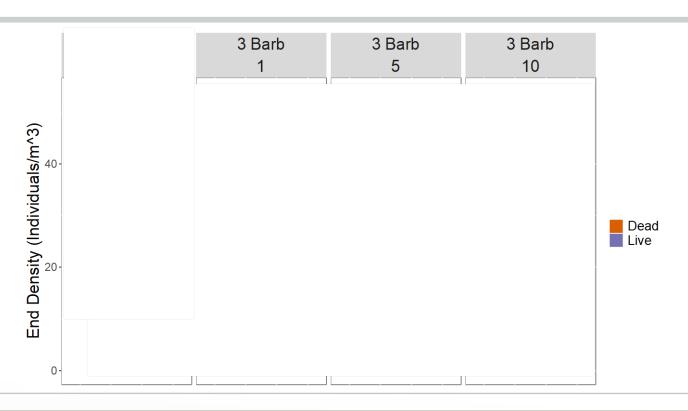
Next questions – Phase 2 Summer 2021

- What about different life stages?
 - Compare 2 and 3 barb
 - Examine live vs. dead

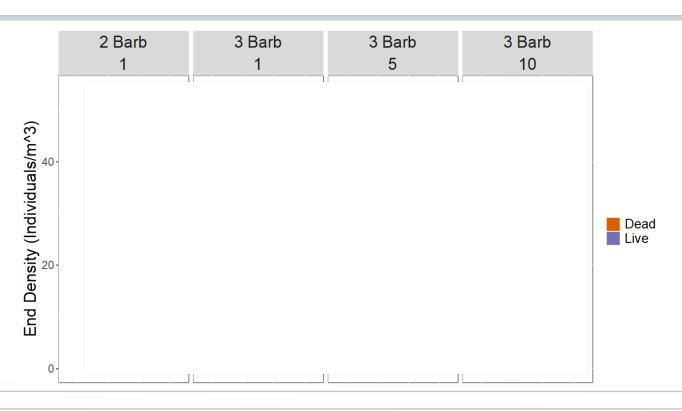
Same treatments as Phase 1



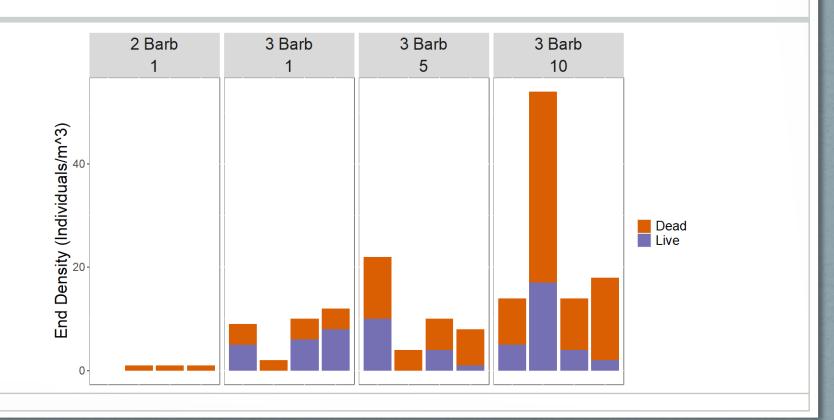
Distinguish live and dead



Add 2 barb life stage



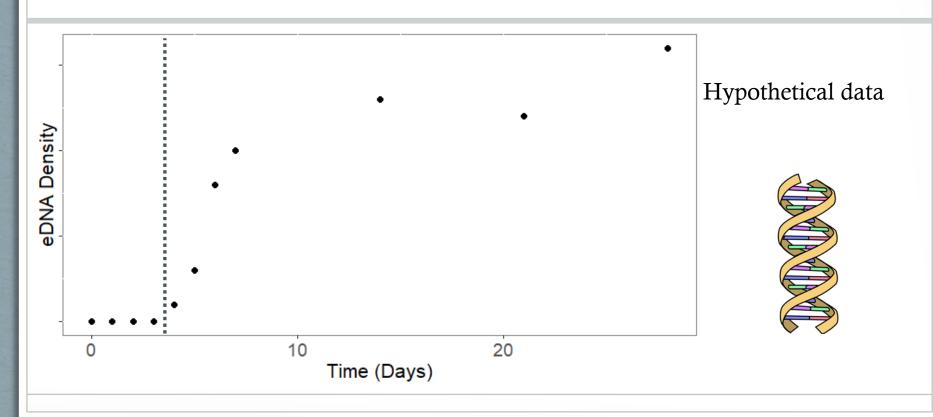
Life stage affects population growth



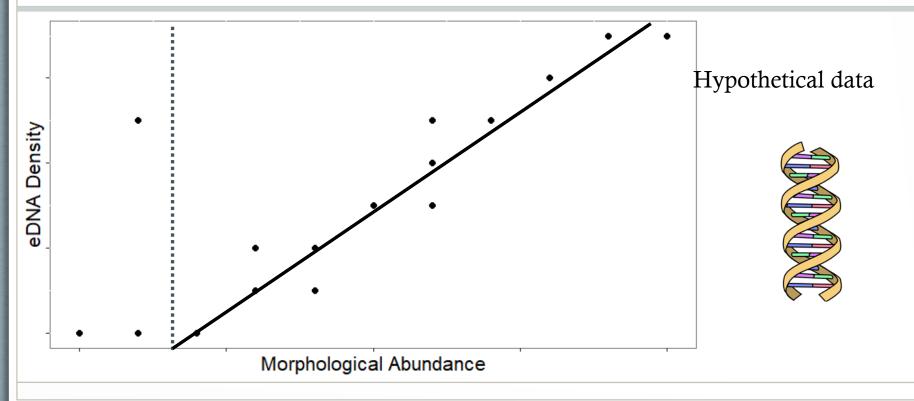
Future questions: eDNA



Predictions: eDNA lag time to detection



Predictions: eDNA density can be calibrated to number of individuals



Phase 3: Summer 2022

- How many individuals at an earlier life stage need to be introduced to get population growth?
- What is the eDNA signature for different spiny water flea life stages?

Life stage	1	5	10
1-barb		X	X
2-barb		X	X
3-barb	X	X	
		†	†
		Average	IMO Standard

clutch size

Conclusions

- Mesocosm experiments can be used to determine reasonable thresholds of introduction in evaluating ballast treatment
- Develop eDNA tool for evaluating ballast treatment

Acknowledgements

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Great Lakes
RESTORATION

Great Waters

Research Collaborative









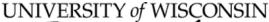
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Time

