

NAMs for Estrogenicity

Richard Judson, PhD

National Academies Committee on Variability and Relevance of Current Laboratory Mammalian Toxicity Tests and Expectations for New Approach Methods (NAMs) for use in Human Health Risk Assessment

May 12, 2022



Phone: 919-449-7514 Judson.richard@epa.gov

The views expressed in this presentation are those of the author and do not necessarily reflect the views or policies of the U.S. EPA



- Community Needs
 - People are exposed to thousands of chemicals
 - In vitro NAMs can be used to rapidly test many chemicals
- Variability and Concordance
 - There is variability across in vitro methods, and within in vivo methods (lab-to-lab)
 - In vitro to in vivo concordance is approximately the same as lab-to-lab in vivo
- In Vivo vs. In Vitro Adversity
 - In vitro assays are good predictors of the uterotrophic response
 - Metabolic activation / deactivation is challenging, but NAM methods to address this are being developed
- Data for Benchmarking New Approaches
 - A large collection of reference chemicals are available
 - 1800+ chemicals have been evaluated *in vitro*, although few have corresponding *in vivo* data
 - Very little direct human data

Community Needs

- Over a lifetime, people can be exposed to many thousands of chemicals
 - Few of these have been thoroughly tested for toxicity, including estrogenicity
- EPA is required to test for potential endocrine disrupting chemicals through the Food Quality Protection Act (FQPA) and other statutes
 - Origin of the Endocrine Disruptor Screening Program (EDSP)
 - There are ~10,000 chemicals on the EDSP queue (pesticides, chemicals potentially found in drinking water)
- In vitro NAMs have been used to test 1,800 chemicals and more are in queue
- In silico NAMs (QSAR models) have tested all defined chemicals in the EDSP Universe

SEPA



Variability and Concordance

- We have tested 1800 chemicals for estrogenicity in 18 separate *in vitro* assays
 - Different points on the biological pathway
 - Different cell types
 - Different readout technologies
- Key learnings:
 - All *in vitro* (and probably all *in vivo*) assays can show false positive results through "assay interference" – the assay is positive for reasons unrelated to the target the assay is supposed to assess
 - Best to use "orthogonal" assays if multiple different technologies give the same results, confidence in result (positive or negative) is increased.
 - Quantitative results (potency) may still be variable

Judson et al., Tox.Sci. doi: 10.1093/toxsci/kfw092 (2016) Judson, et al., ToxSci 148 (1) pp 137-154 (2015) Browne et al., Environ. Sci. Technol. 2015, 49, 8804–8814, DOI: 10.1021/acs.est.5b02641

Example of Quantitative Uncertainty

80–05–7 : Bisphenol A



SEPA

Judson, et al., ToxSci 148 (1) pp 137-154 (2015)

SEPA *In Vitro* to *In Vivo* Concordance





42 Chemicals with *in vitro* and *in vivo* data
1 false negative – chemical was volatile *in vitro*1 false positive – chemical is metabolically deactivated *in vivo*

Variability in published *in vivo* uterotrophic data for Bisphenol A Potency spans ~3 orders of magnitude

Browne et al. Environ. Sci. Technol. 2015, 49, 8804–8814, DOI: 10.1021/acs.est.5b02641

Set EPA

In Vivo and In Vitro Adversity

- NAMs can model some of the *in vivo* effects of estrogenic chemicals
- Estrogen receptor (ER) in vitro assays answer the following questions
 - Does the chemical interact with the ER?
 - Is the interaction in an agonist or antagonist mode?
 - Does the chemical cause ER-dependent cell proliferation?
 - What blood concentration is required to have an ER-related effect?
 - Assays test the human ER
- In vitro toxicokinetics assays and models answer the following question
 - What <u>human</u> oral dose is required to reach the blood level that is required to have an ERrelated effect?
- Exposure NAM models answer the following question
 - Are <u>humans</u> likely to be exposed at levels that cause ER-related effects?
- Cannot directly predict what adverse phenotype will be seen

Data for Benchmarking New Methods

• Does a chemical interact with ER?

SEPA

- Good reference chemicals and published assays to compare against (see Browne et al. (2016))
- Is the chemical estrogenic through a non-genomic mechanism (GPR30)?
 - Reference chemicals and methods are sparse
 - Not clear how important this is for environmental chemicals
- Does the chemical affect estrogen signaling through the steroidogenesis pathway?
 - Chemicals can block the production of estrogen
 - Reference chemicals and methods exist (see Haggard et al. (2017))

SEPA Bibliography

- 1. P Browne, RS Judson, W Casey, N Kleinstreuer, RS Thomas, "Screening chemicals for estrogen receptor bioactivity using a computational model", Environ Sci. & Technology, 49 (14) pp 8804-8814 (2015).
- RS Judson, FM Magpantay, V Chickarmané, C Haskell, N Tania, J Taylor, M Xia, R Huang, DM Rotroff, DL Filer, KA Houck, MT Martin, N Sipes, AM Richard, K Mansouri, RW Setzer, TB Knudsen, KM Crofton, RS Thomas, "Integrated Model of Chemical Perturbations of a Biological Pathway Using 18 In Vitro High Throughput Screening Assays for the Estrogen Receptor", ToxSci 148 (1) pp 137-154 (2015)
- 3. Kleinstreuer N, P Ceger, M Martin, K Houck, P Browne, R Thomas, E Watt, W Casey, D Dix, D Allen, M Xia, R Huang, R Judsón, "Development and Validation of a Computational Model for Androgen Receptor Activity" Chem.Res.Tox. vol 30 946-964(2016)
- 4. Lynch et al. "Identifying Environmental Chemicals as Agonists of the Androgen Receptor by Using a Quantitative High-throughput Screening Platform" Toxicology Vol. 385 48-58 (2017)
- 5. Judson et al. "On selecting a minimal set of in vitro assays to determine the estrogenicity of a chemical" RegToxPharm Vol 91, p 39 (2017)
- 6. Watt E., R Judson, "Uncertainty analysis of in vitro Estrogen Receptor Model" PLoS One Vol 13 e019693 (2018)
- 7. Kleinstreuer NC, Browne P, Chang X, Judson R, Casey W, Ceger P, Deisenroth C, Baker N, Markey K Thomas RS, "Evaluation of Androgen Assay Results using a Curated Hershberger Database", Reproductive Toxicology, Vol 81, pp 272-280 (2018)
- 8. Browne P, N. Kleinstreuer, P Ceger, C Deisenroth, N Baker, K Markey, R Thomas, R Judson, W Casey, "Development of a Curated Hershberger Database" Reproductive Toxicology, Vol 81, pp 259-271 (2018)
- 9. Judson et al. "New Approach Methods for Testing Chemicals for Endocrine Disruption Potential" Current Opinions in Toxicology Vol 9, pp 40-47 (2019)
- 10. K Mansouri [39 other authors], R.S. Judson, "CERAPP: Collaborative Estrogen Receptor Activity Prediction Project", Environ. Health Perspec. 124:7 pp 1023-1033 (2016)
- 11. Mansouri et al. "CoMPARA: Collaborative Modeling Project for Androgen Receptor Activity", Env Health Pers, vol 128 no. 2 (2020)
- 12. R Judson, K Houck, K Paul Friedman, J Brown, P Browne, PA Johnston, DA Close, N Kleinstreuer, "Selecting a Minimal set of Androgen Receptor Assays for Screening Chemicals", RegToxPharm, vol 117, https://doi.org/10.1016/j.yrtph.2020.104764 (2020)
- 13. R Judson, K Houck, M Martin, AM Richard, TB Knudsen, I Shah, S Little, J Wambaugh, RW Setzer, P Kothya, J Phuong, D Filer, D Smith, D Reif, D Rotroff, N Kleinstreuer, N Sipes, M Xia, R Huang, K Crofton, RS Thomas, "Analysis of the Effects of Cell Stress and Cytotoxicity on In Vitro Assay Activity in the ToxCast Dataset", Tox.Sci. doi: 10.1093/toxsci/kfw092 (2016)
- 14. Haggard, D., AL Karmaus, MT Martin, RS Judson, RW Setzer, K Paul-Friedman, "High-throughput H295R steroidogenesis assay: utility as an alternative and a statistical approach to characterize effects on steroidogenesis", Tox Sci Vol 162 p. 509 (2017)