

Molecular Point of Departure (mPOD) Determination From *In Vitro* High-Throughput Transcriptomics Data.

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NAMs-Based Tiered Hazard Evaluation Approach

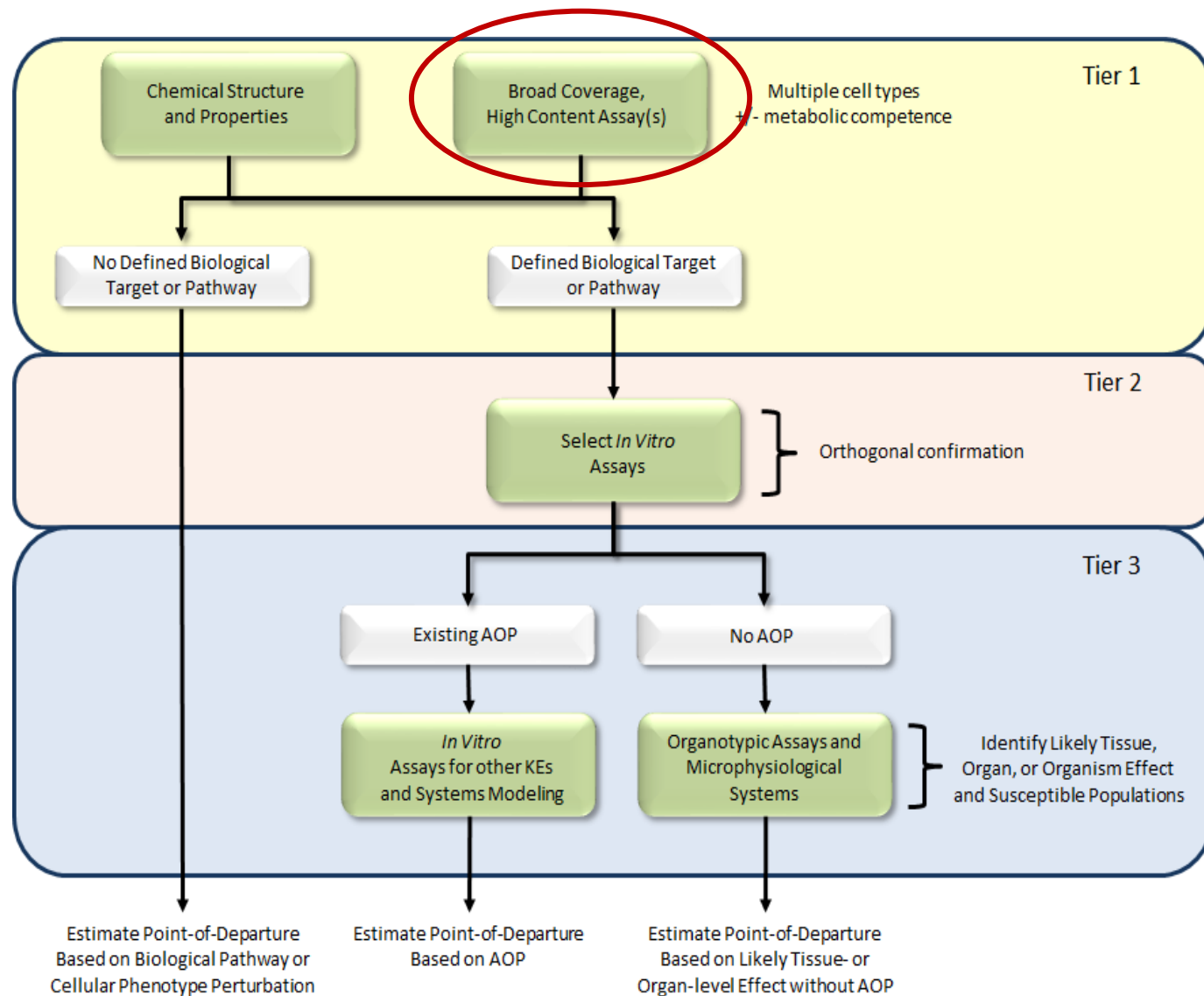
High throughput profiling (HTP) assays are proposed as the first tier in a NAMs-based hazard evaluation approach.

HTP Assay Criteria:

1. Yield bioactivity profiles that can be used for **potency estimation, mechanistic prediction** and evaluation of **chemical similarity**.
2. Compatible with multiple human-derived culture models.
3. Concentration-response screening mode.
4. Cost-effective.

To date, EPA has identified and implemented two HTP assays that meet this criteria.

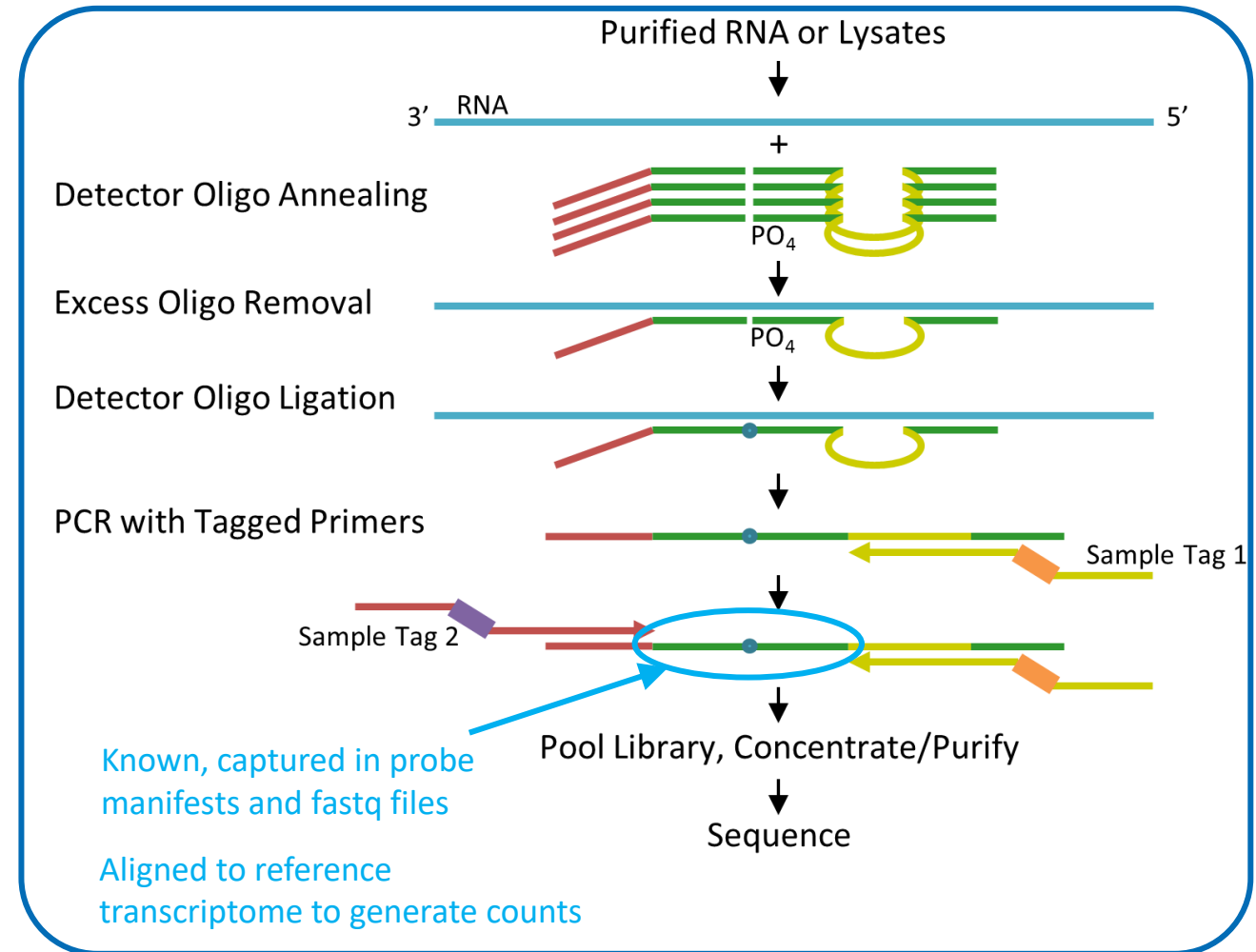
- **High-Throughput Transcriptomics [HTTr]**
- **High-Throughput Phenotypic Profiling [HTPP]**



Templated Oligo with Sequencing Readout (TempO-Seq)

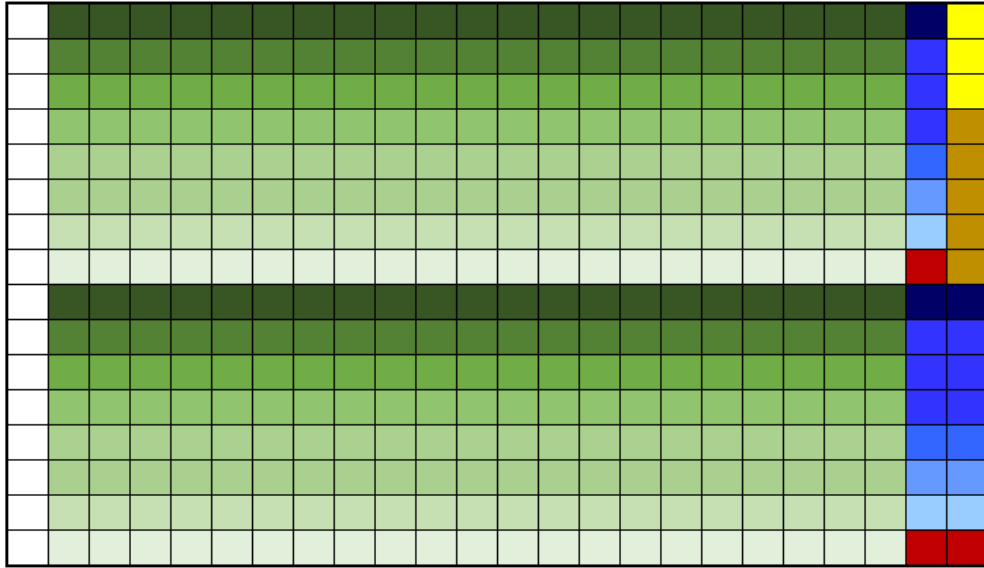
- The **TempO-Seq** human whole transcriptome assay measures the expression of greater than 20,000 transcripts.
- Requires only picogram amounts of total RNA per sample.
- Compatible with purified RNA samples or **cell lysates**.
- Lysates are barcoded according to sample identity and combined in a single library for sequencing using industry standard instruments.
- Scalable, targeted assay:
 - 1) specifically measures transcripts of interest
 - 2) ~50-bp reads for all targeted genes
 - 3) requires less flow cell capacity than RNA-Seq

TempO-Seq Assay Illustration



Generic Experimental Design for HTTr

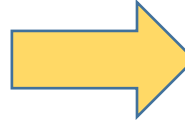
Dose Plate



- = Test chemicals in 8-point dilution series
- = Vehicle controls (DMSO)

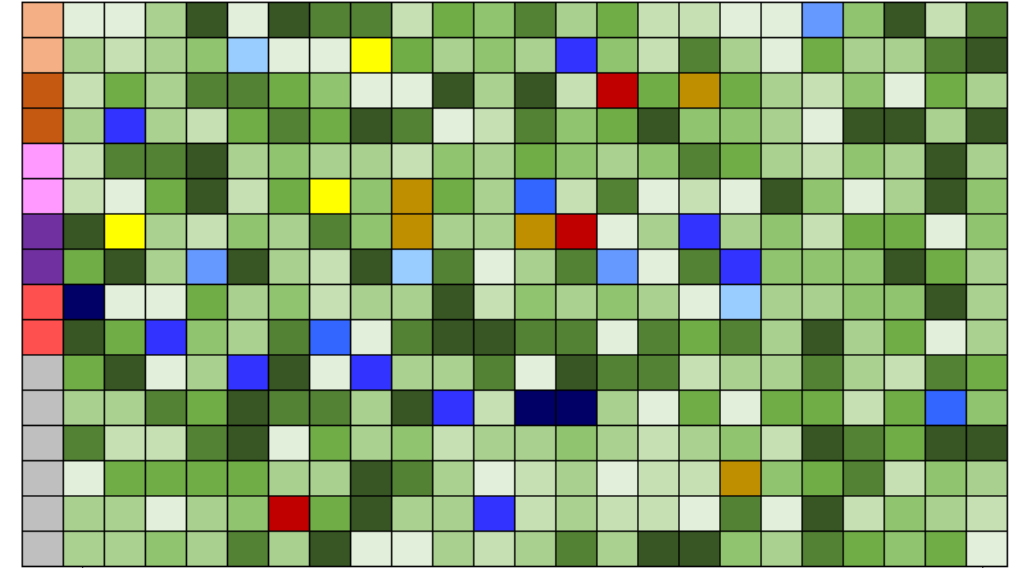
- = Reference chemicals in 7-point dilution series
- = Trichostatin A (cell type agonist reference chem)
- = Staurosporine (cell type agnostic cell viability control)

Used to track assay performance.



**LabCyte® Echo 550
Acoustic Dispenser**

Assay Plate



No cells

Cells






- = Reference RNAs
- = Reference Lysates
- = Bulk Lysates

*Used to track assay performance independent of
Chemical treatments and responsivity of culture.*

- = Reserved for sequencing vendor

MCF7 Pilot Experimental Design

High-Throughput Transcriptomics Platform for Screening Environmental Chemicals

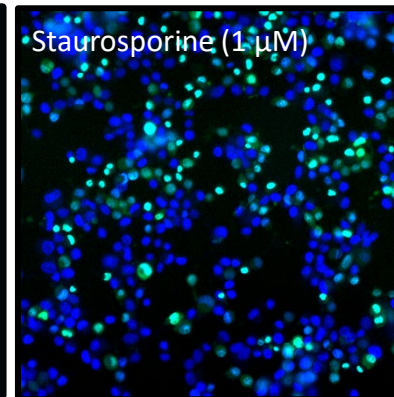
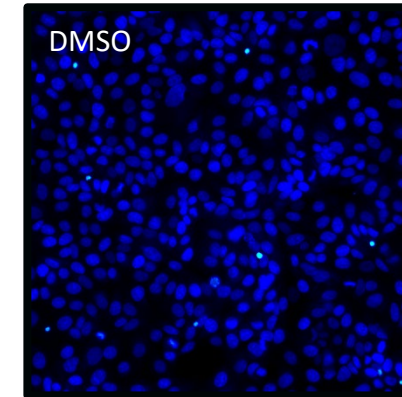
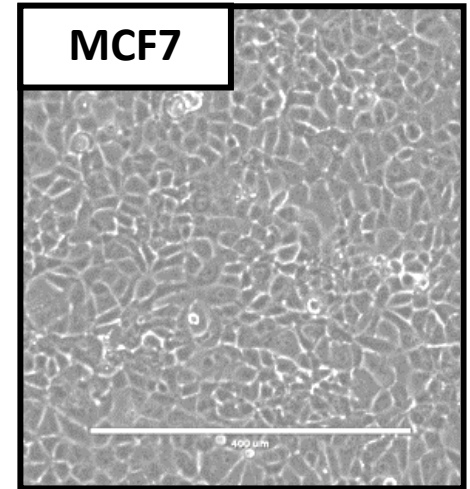
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TOXICOLOGICAL SCIENCES, 2021, 1–22

doi: [10.1093/toxsci/kfab009](https://doi.org/10.1093/toxsci/kfab009)

Advance Access Publication Date: 4 February 2021
Research Article

Parameter	Multiplier	Notes
Cell Type(s)	1	MCF7
Assay Formats:	2	High-Throughput Transcriptomics Cell Viability
Culture Condition	1	DMEM + 10% HI-FBS
Chemicals	44	ToxCast chemicals
Time Points:	1	6 hours
Concentrations:	8	3.5 log ₁₀ units; semi log ₁₀ spacing
Biological Replicates:	3	Independent cultures



CellEvent Caspase 3/7

MCF7 Pilot Chemical List

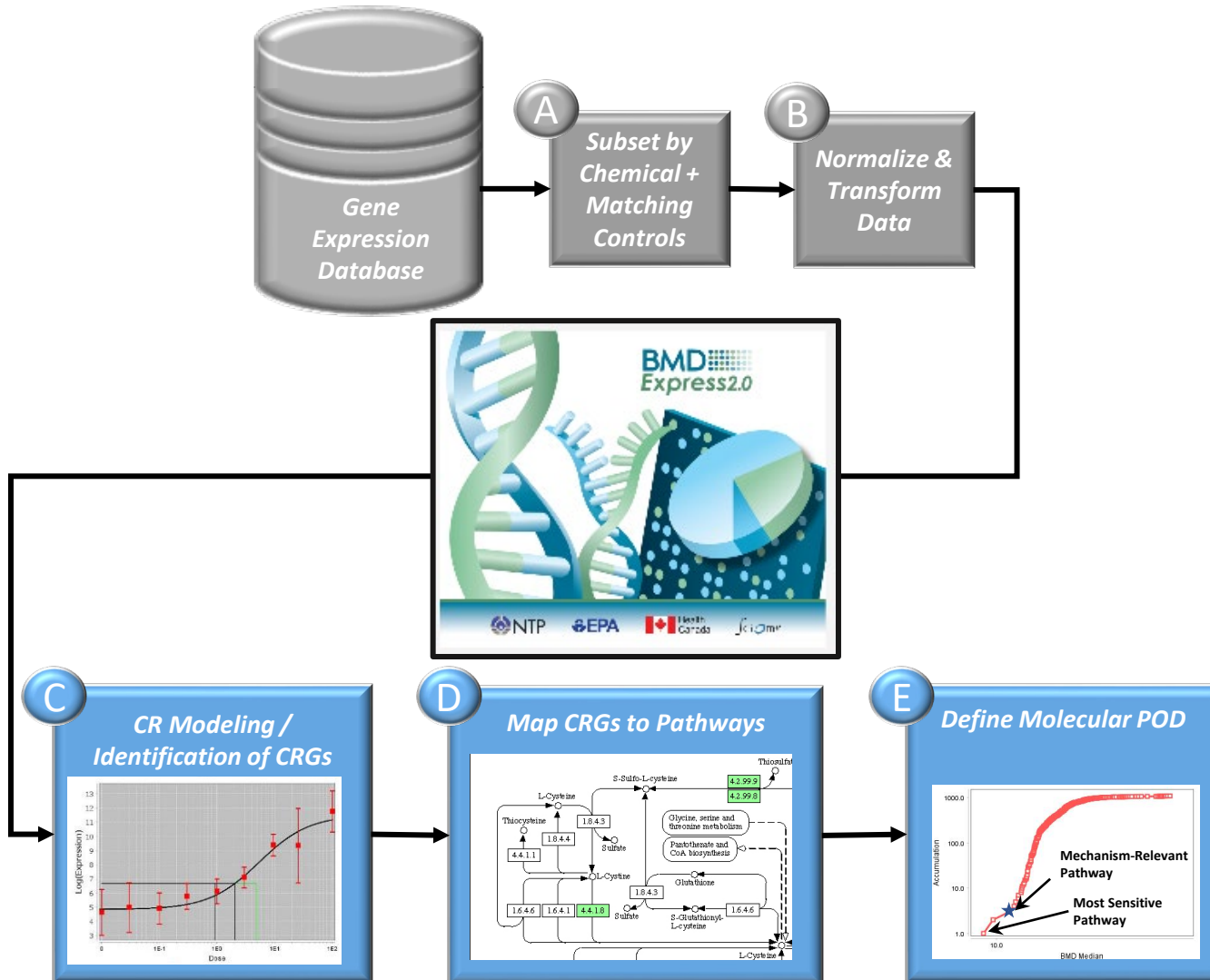
Table 1. Chemicals Used in the Study

Name	Target Annotation	Name	Target Annotation
Cyproterone acetate	AR antagonist	Lovastatin	HMGCR inhibitor
Flutamide	AR antagonist	Simvastatin	HMGCR inhibitor
Nilutamide	AR antagonist	Maneb	Inhibition of metal-dependent and sulfhydryl enzyme systems
Vinclozolin	AR antagonist	Thiram	Inhibition of metal-dependent and sulfhydryl enzyme systems
Amiodarone hydrochloride	Blocks myocardial calcium, potassium and sodium channels	Ziram	Inhibition of metal-dependent and sulfhydryl enzyme systems
Cladribine	DNA synthesis inhibitor	Reserpine	Inhibition of the ATP/Mg ²⁺ pump
4-Cumylphenol	ER agonist	Rotenone	Mitochondria (complex I inhibitor)
4-Nonylphenol, branched	ER agonist	Pyraclostrobin	Mitochondria (complex III inhibitor)
Bisphenol A	ER agonist	Trifloxystrobin	Mitochondria (complex III inhibitor)
Bisphenol B	ER agonist	Fenpyroximate (Z, E)	Mitochondrial electron transport inhibitor
4-Hydroxytamoxifen	ER antagonist	Clofibrate	PPAR α agonist, upregulates extrahepatic lipoprotein lipase
Clomiphene citrate (1:1)	ER antagonist	Fenofibrate	PPAR α agonist, upregulates extrahepatic lipoprotein lipase
Fulvestrant	ER antagonist	Farglitazar	PPAR γ agonist
Cyproconazole	Ergosterol-biosynthesis inhibitor. Pan-cyp inhibitor	Perfluorooctanoic acid (PFOA)	PPAR γ , PPAR α agonist
Imazalil	Ergosterol-biosynthesis inhibitor. Pan-cyp inhibitor	Perfluorooctanesulfonic acid (PFOS)	PPAR γ , PPAR α agonist
Prochloraz	Ergosterol-biosynthesis inhibitor. Pan-cyp inhibitor	Troglitazone	PPAR γ , PPAR α agonist
Propiconazole	Ergosterol-biosynthesis inhibitor. Pan-cyp inhibitor	Cycloheximide	Protein synthesis inhibitor
Atrazine	Herbicide, photosystem II inhibitor	Bifenthrin	Sodium channel modulator
Cyanazine	Herbicide, photosystem II inhibitor	Cypermethrin	Sodium channel modulator
Simazine	Herbicide, photosystem II inhibitor	Tetrac	T4 synthesis inhibitor
Butafenacil	Herbicide, PPO inhibition	3,5,3'-triiodothyronine	THR agonist
Fomesafen	Herbicide, PPO inhibition		
Lactofen	Herbicide, PPO inhibition		

- Chemicals were selected that cover a broad range of molecular targets with some redundancy within target class.
- Intentionally selected some chemicals whose molecular targets are not expressed in MCF7 cells (or in mammalian tissues).

BMDExpress for mPOD Determination

Based on National Toxicology Program Approach to Genomic Dose-Response Modeling (NTP RR 5)



BMDExpress Parameter	Criteria
Pre-filter:	$ FC > 2$ at any test concentration
Models	Hill, Power, Linear, Poly2, Exponential 2 3 4 5
BMR Factor:	$1.349 \times \text{SD of controls (10\%)}$
Best Model Selection:	Lowest AIC
Hill Model Flagging:	'k' < 1/3 Lowest Positive Dose Exclude Flagged Hill from Best Model Selection
Conc-Response Hit Criteria	$(0.1 \times \text{lowest conc.} < \text{BMC} < \text{highest conc.})$ BMC fit p-value > 0.1 $\text{BMCL} / \text{BMCU} < 40$
Gene Set Analysis:	≥ 3 Concentration-responsive genes $\geq 5\%$ Gene Set Coverage
Gene Set Collections:	MSigDB (Liberzon et al. 2015) BioPlanet (Huang et al. 2019) CMAP (Subramanian et al. 2005)
Molecular Point of Departure	Most Sensitive Gene Set

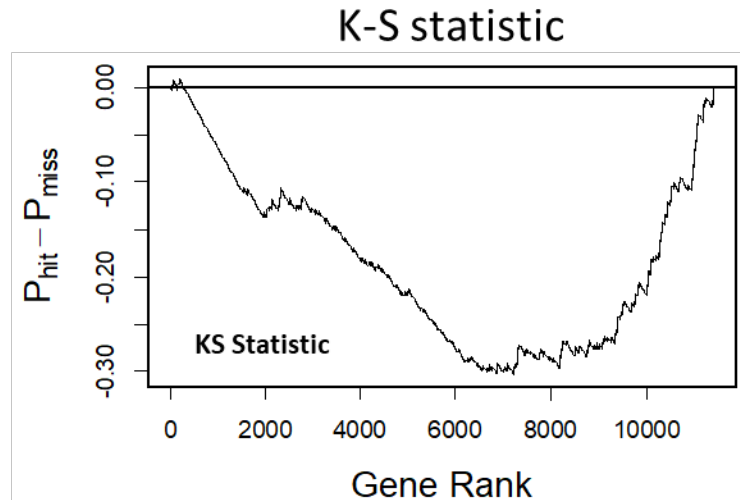
Modeling of Signature Scores for mPOD Determination (1)

Step 1: Inputs

Experimental Data: Chemical_Conc × Gene matrix of \log_2 (fold-change) (l2fc) values.
Signature Collections: MSigDB (*Liberzon et al. 2015*), BioPlanet (*Huang et al. 2019*), CMAP (*Subramanian et al. 2005*)

Step 2: Pathway Scoring

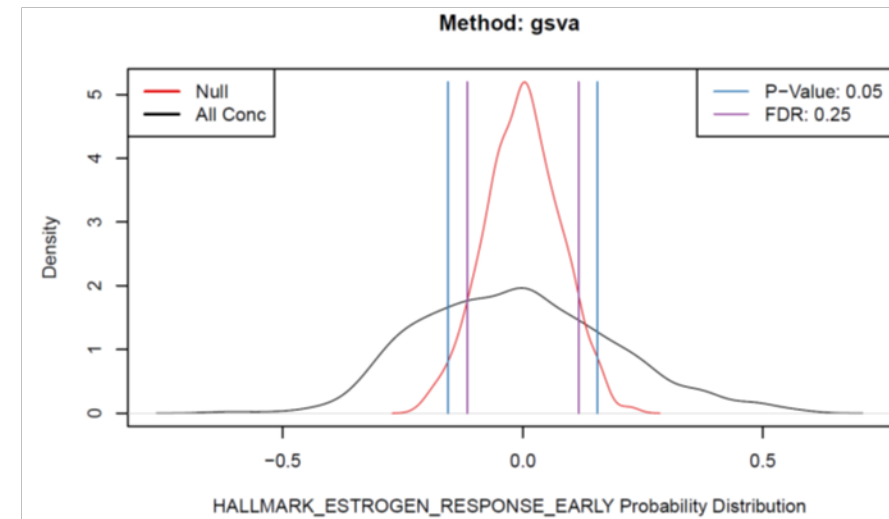
Scores based on single sample GSEA method (Barbie et al. 2009)



Chemical_Conc × Pathway matrix of scores.

Step 3: Cut-off Estimation via NULL Modeling

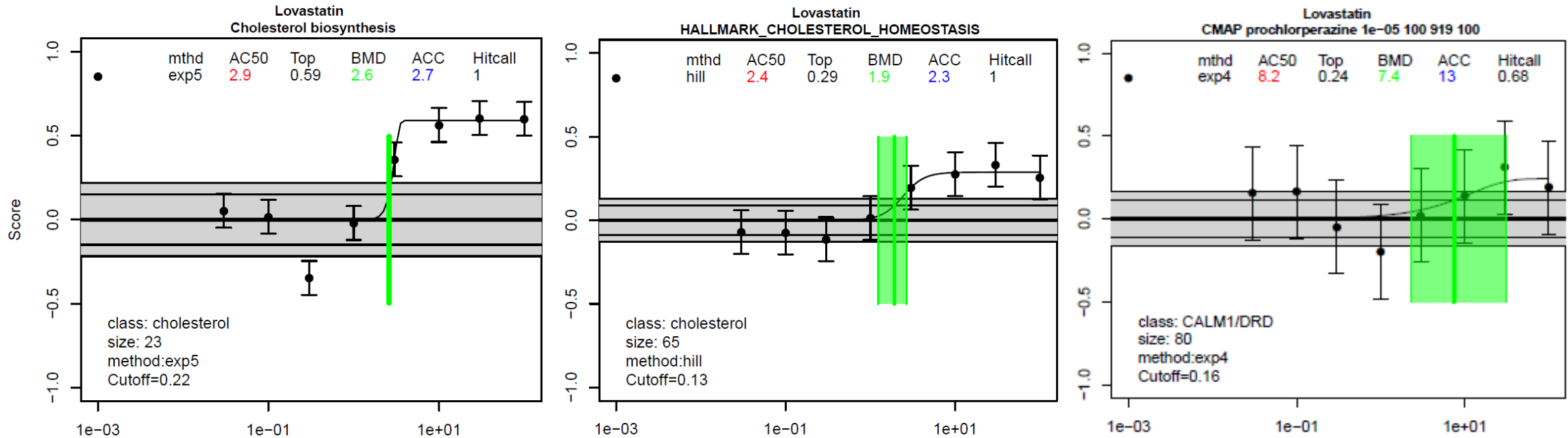
- For each gene, **resample** l2fc based on the cross-sample gene distribution
- Calculate **pathway scores for “null” data**
 - One null distribution (n = 1000 scores) / pathway



Modeling of Signature Scores for mPOD Determination (2)

Step 4: CR Modeling

Concentration response modeling of signature scores using *tcplfit2* (Sheffield et al. (2021) [10.1093/bioinformatics/btab779](https://doi.org/10.1093/bioinformatics/btab779))

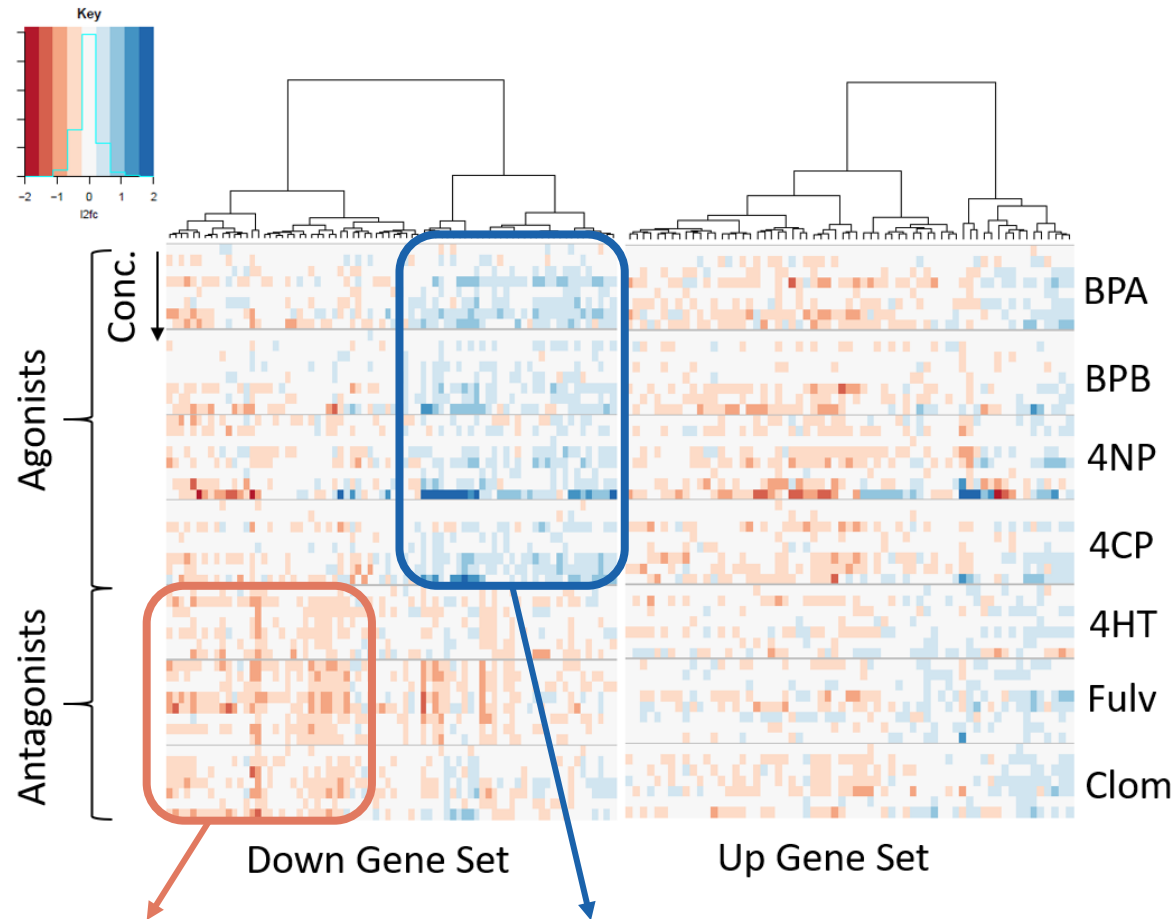


- Takes into account coordinated changes in gene expression that may not be identified using gene level fitting approaches.
- All curve forms from BMDExpress, plus constant model.
- Provides continuous hit calls for identifying high confidence and low confidence hits.

MCF7 Pilot Results: Directionality of Signature Scores

A

Fulvestrant Signature (Top 100 Up & Down Genes)

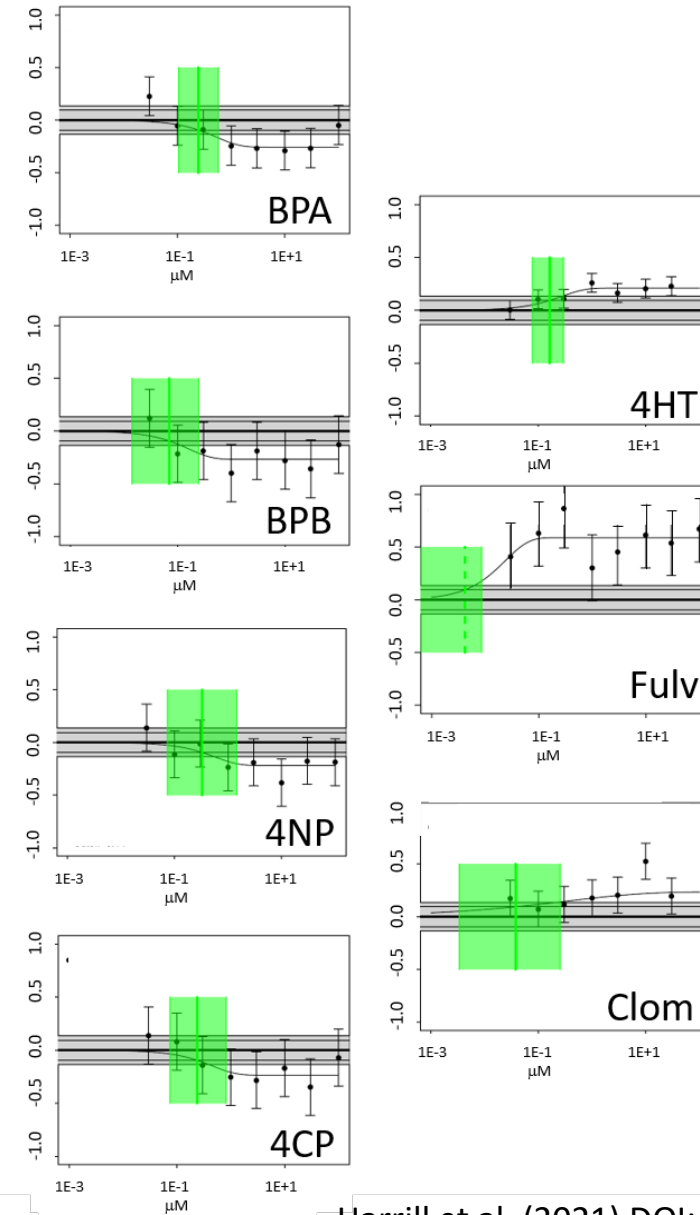


These
gene level
data are
noisy!

The expression of fulvestrant
signature "down" genes goes down
following ER antagonist treatment

The expression of fulvestrant
signature "down" genes goes up
following ER agonist treatment

B



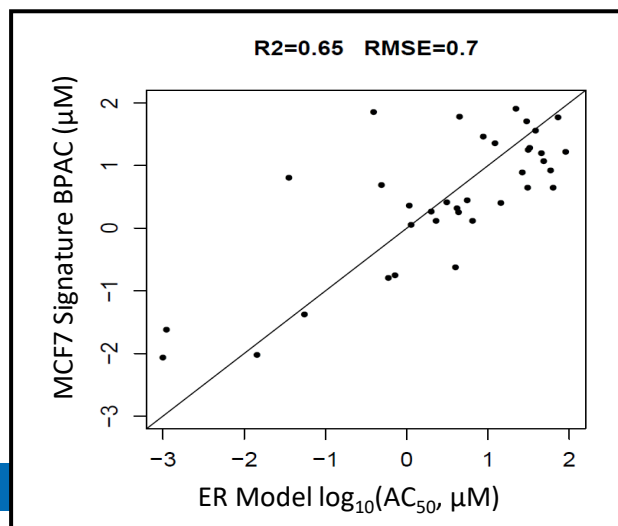
Signature
level results
display
correct
directionality!

MCF7 Pilot Results: Comparison of mPOD Approaches

- BPAC_{Sig}** → 5th lowest BPAC of active signatures
- BPAC_{BMDX}** → Most sensitive signature / pathway
- BPAC_{HTS}** → Lower 5th percentile of active AC50 values for ToxCast assays that pass a series of quality filters.

BPAC_{HTS} and BPAC_{Sig} are in better agreement than BPAC_{HTS} and BPAC_{BMDX}

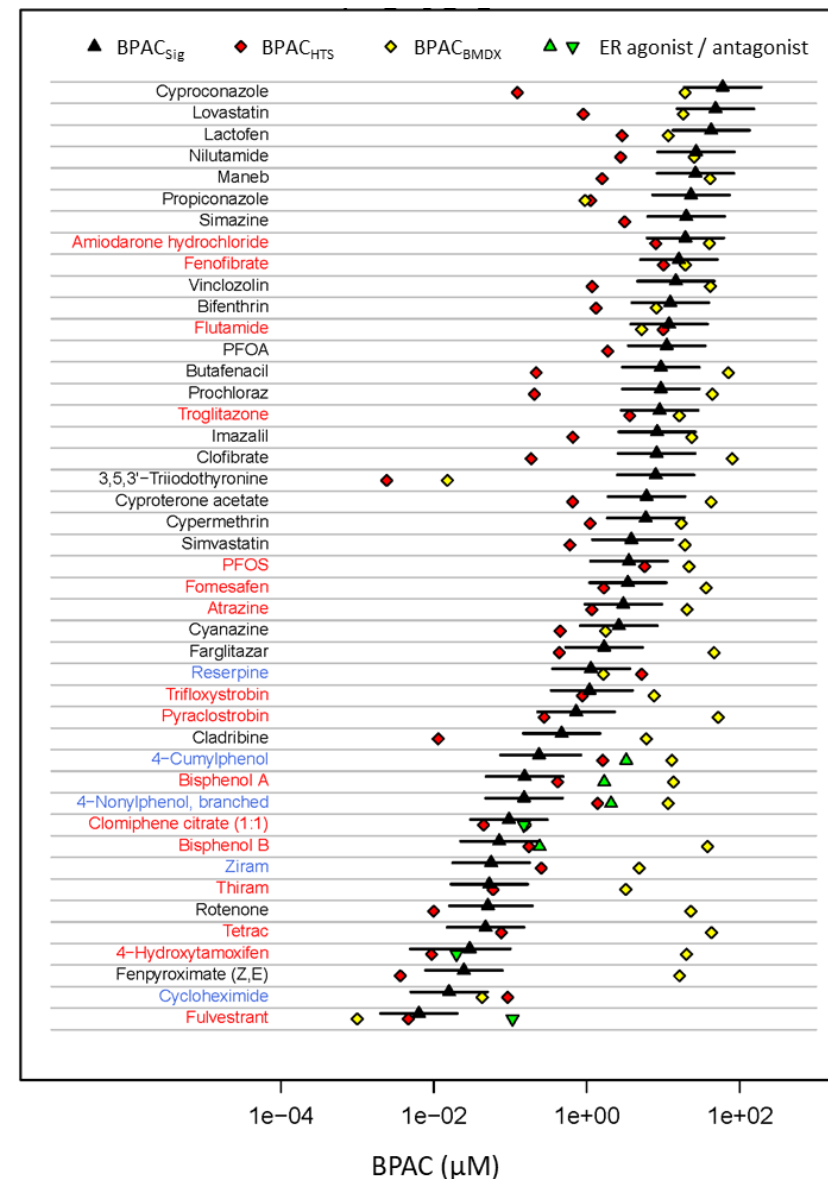
In most of these cases, BPAC_{HTS} is also more potent than BPAC_{BMDX}.



Signature-based BPACs in MCF7 are concordant with ToxCast estrogen receptor (ER) model predictions.

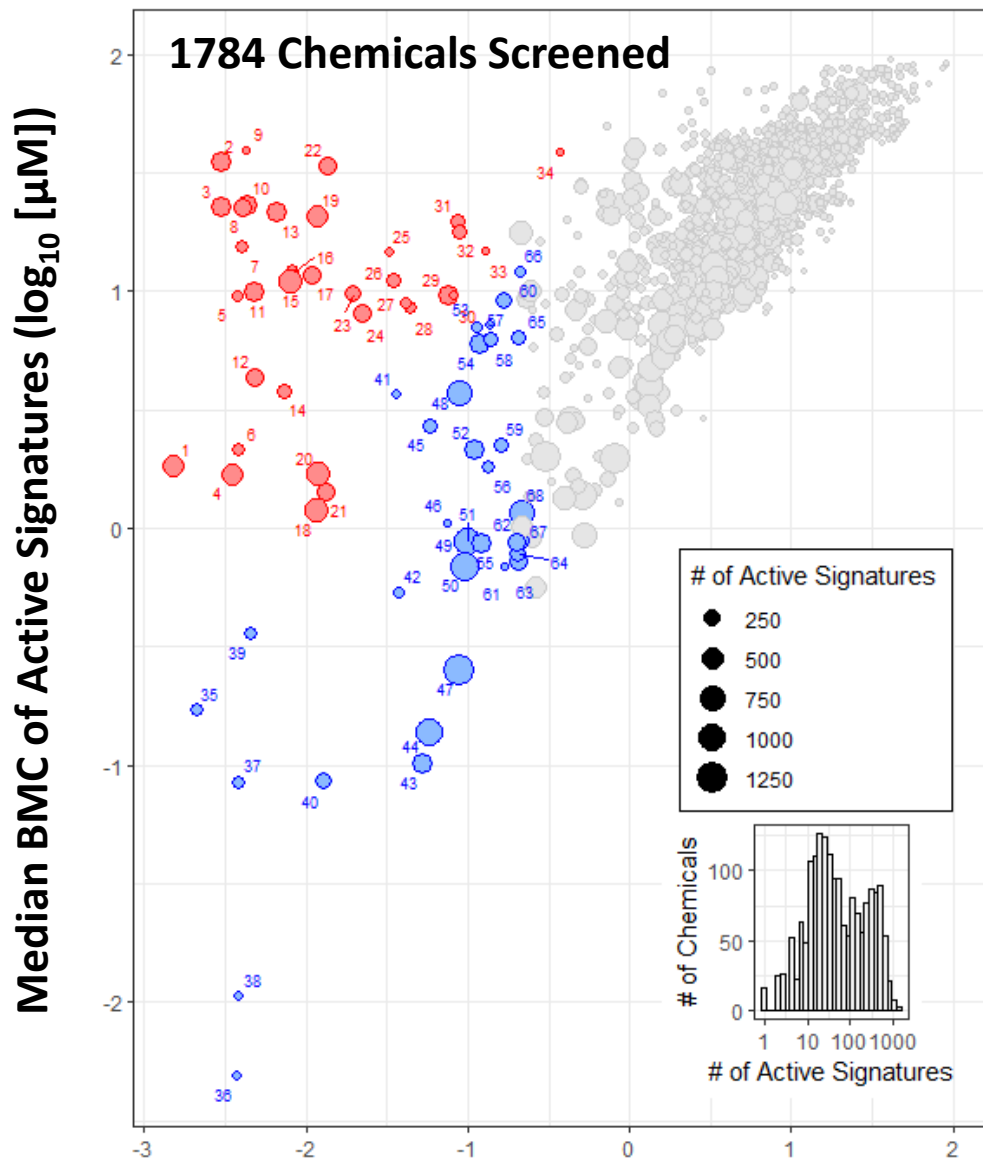
- Brown et al. (2015) DOI: [10.1021/acs.est.5b02641](https://doi.org/10.1021/acs.est.5b02641)

BPAC = Biological Pathway Altering Concentration



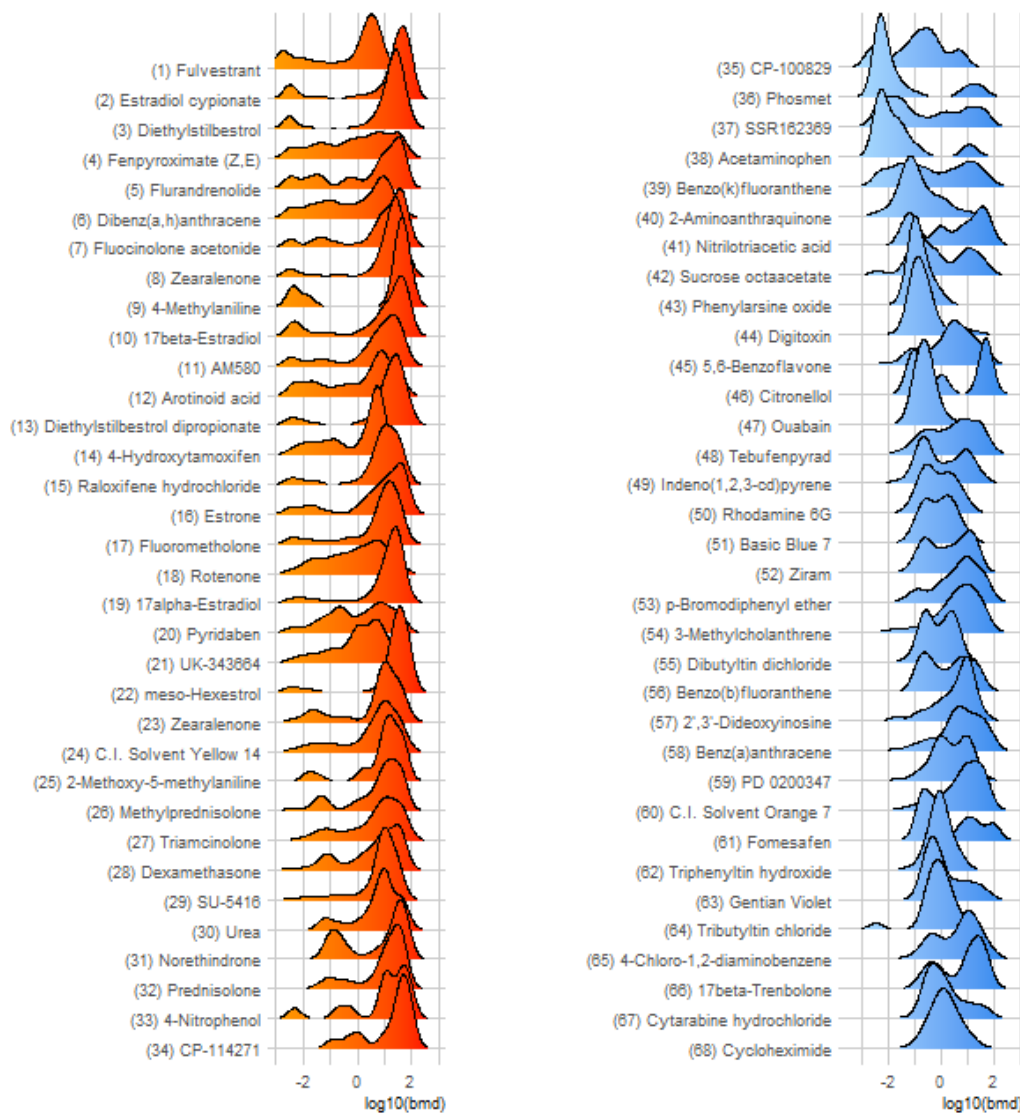
Harrill et al. (2021) DOI: [10.1093/toxsci/kfab009](https://doi.org/10.1093/toxsci/kfab009)

MCF7 HTTr Screening Results (1)



5th % BMC of Active Signatures ($\log_{10} [\mu\text{M}]$)

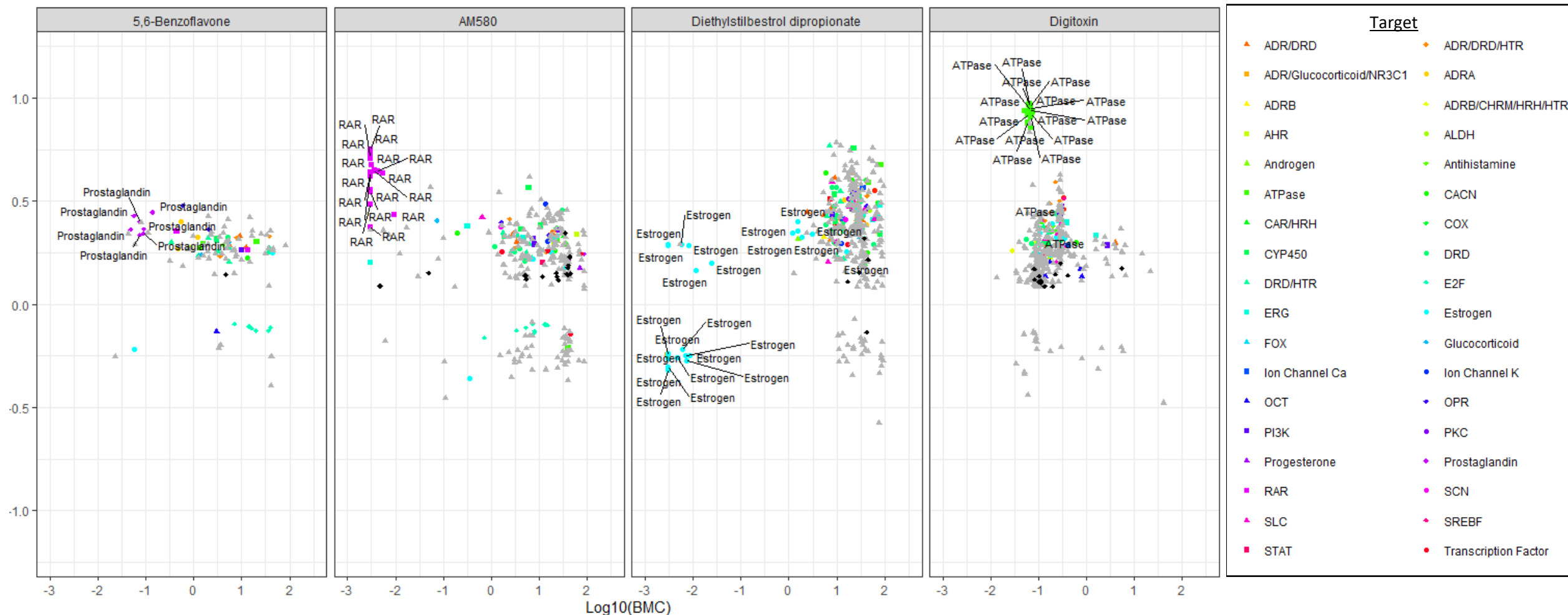
Distribution of BMCs of Active Signatures



Other potent toxicants (organometallics, dyes, etc) cause many signatures to be affected near the onset of biological activity.

Chemicals with known pharmacological targets show an “early wave” of biological activity.

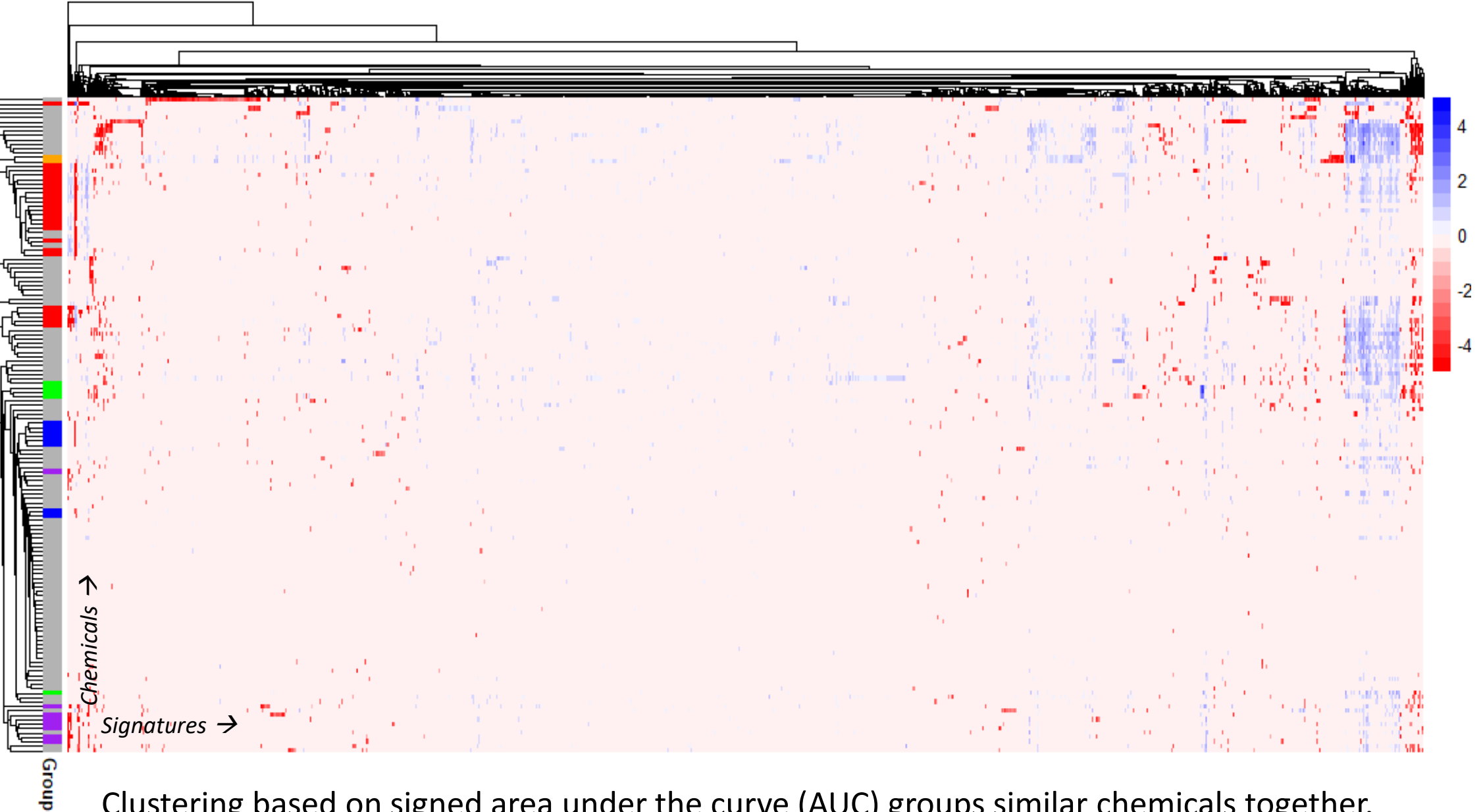
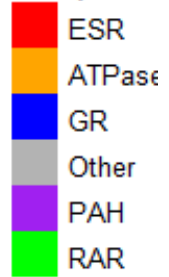
MCF7 HTTr Screening Results (2)



The most potent and efficacious signature hits correspond to known mechanisms for these chemicals.

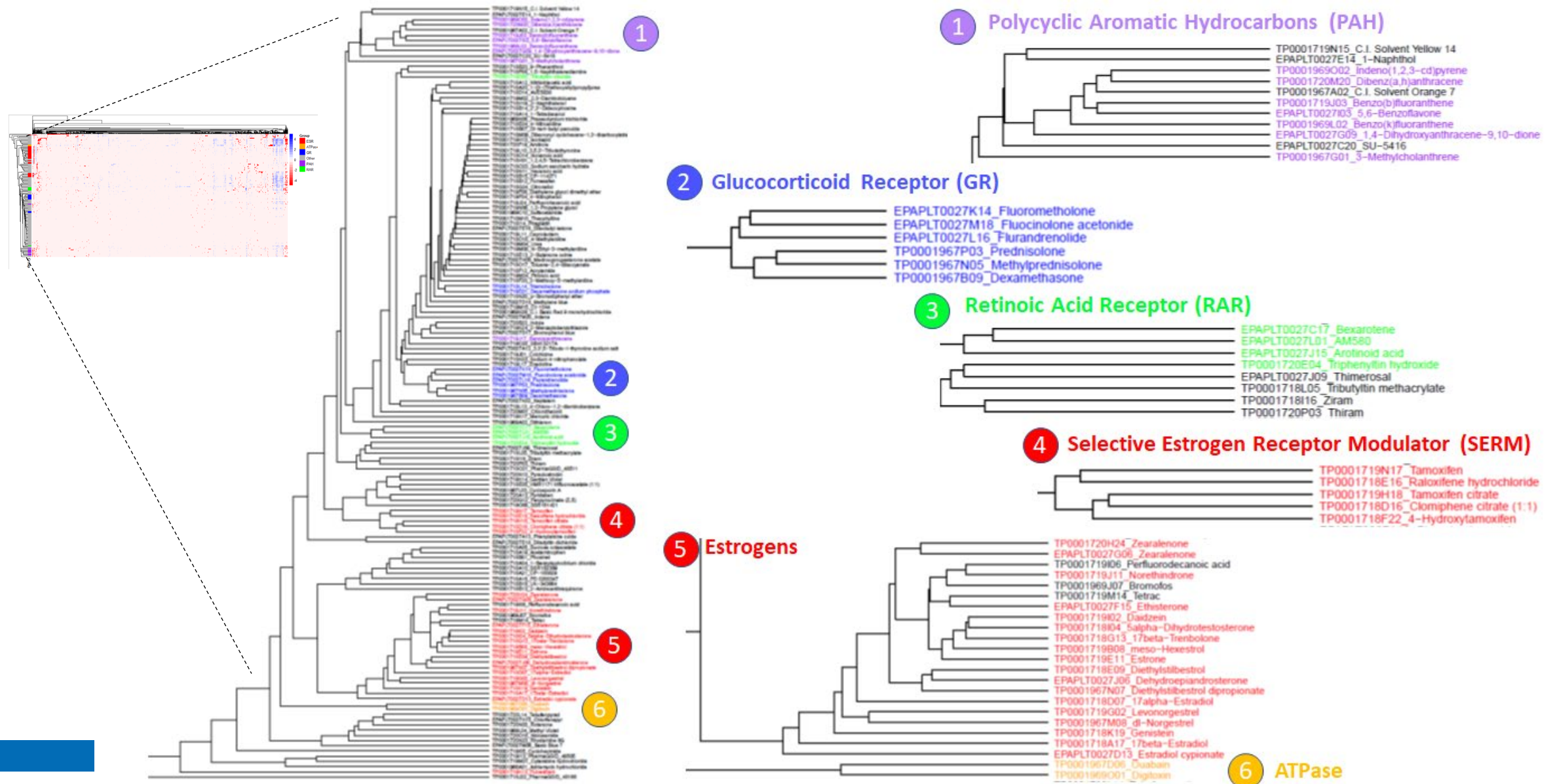
MCF7 HTTr Screening Results (3)

Chemical Target Group

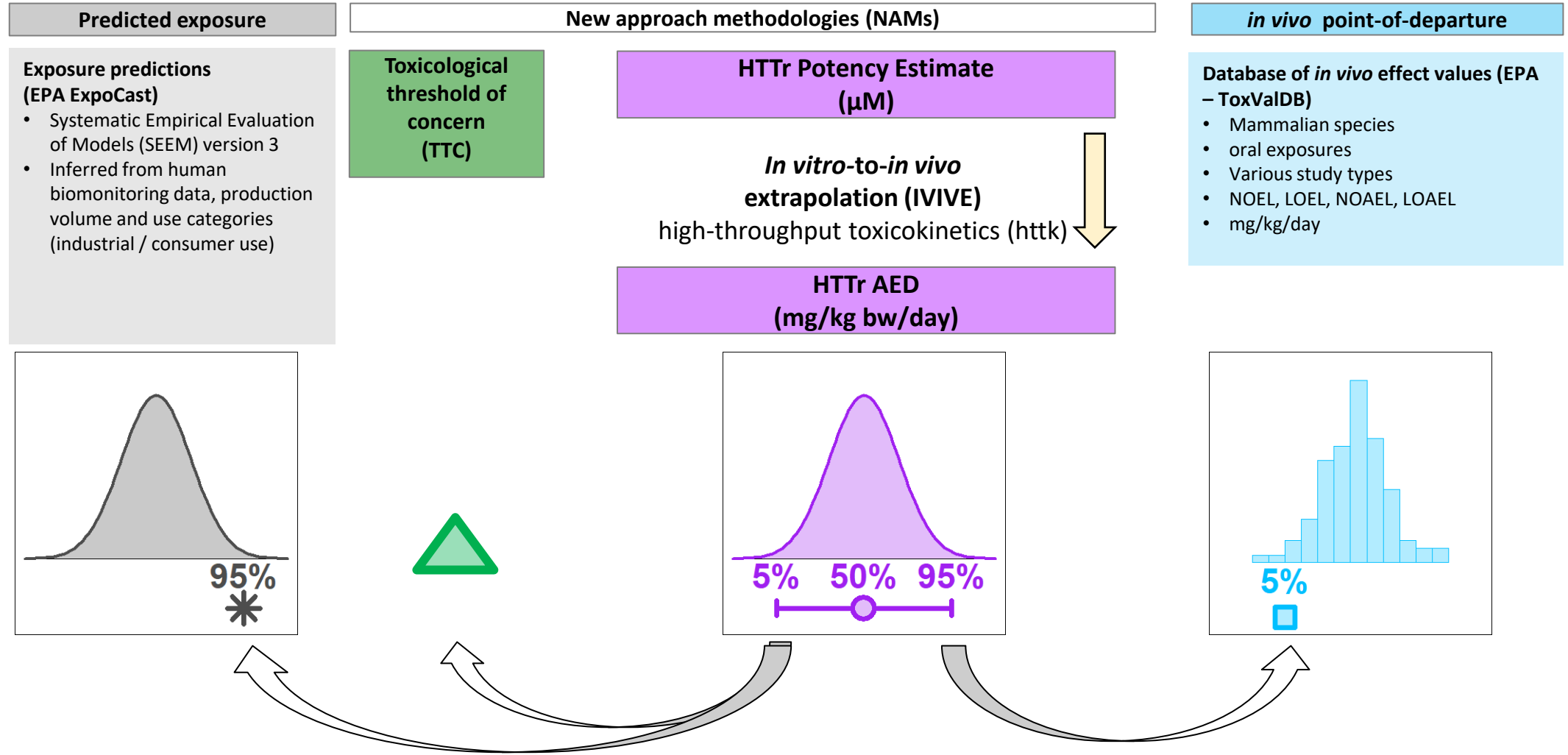


Clustering based on signed area under the curve (AUC) groups similar chemicals together.

MCF7 HTTr Screening Results (4)

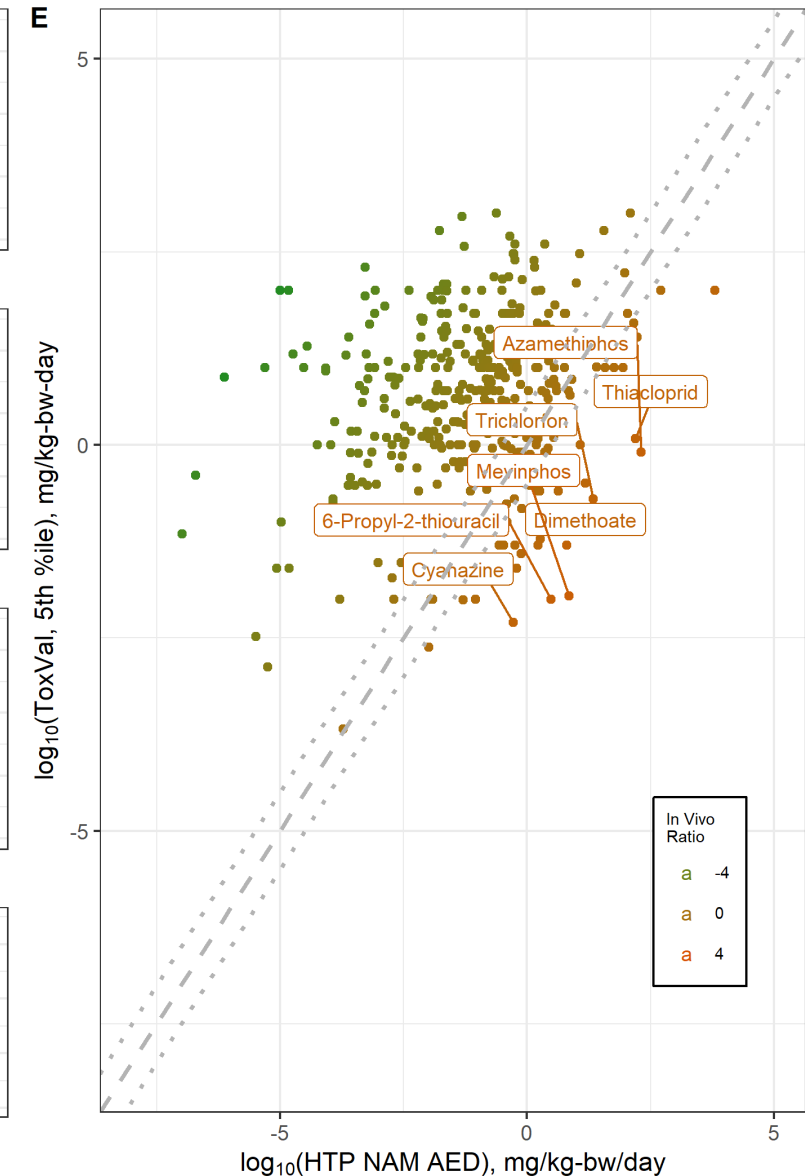
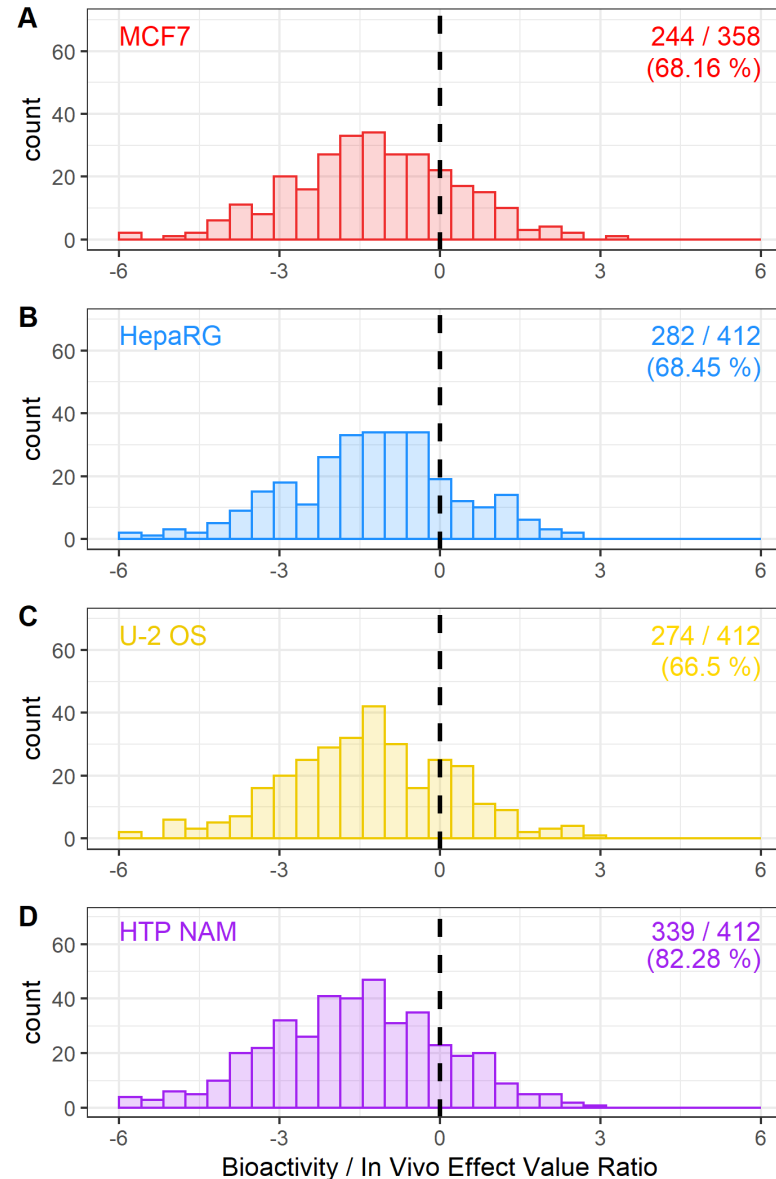


In Vitro to *In Vivo* Extrapolation (IVIVE) Using High-Throughput Toxicokinetic (httk) Modeling



Bioactivity / *In Vivo* Effect Value Ratio Analysis

- Negative ratios** indicate that AEDs derived from HTP NAMs molecular PODs are **conservative** surrogates for traditional *in vivo* PODs.
- When cell lines are considered individually, **~66-68%** of chemicals had negative ratios.
- When considered in combination, the number and percentage of chemicals with negative ratios **increased (82.3 %)**.
- Paul Friedman et al. (2020) (PMID: [31532525](#))
 - Using ToxCast, **89 %** of APCRA chemicals had negative ratios.
- When multiple cell types are considered, mPODs from HTTr screening appear to be conservative surrogates for *in vivo* PODs.
- Correlation of *in vitro* and *in vivo* is low.



Acknowledgements



Office of Research and Development (ORD) Center for Computational Toxicology and Exposure (CCTE)

- Johanna Nyffeler
- Clinton Willis
- Rick Brockway
- Megan Culbreth
- Dan Hallinger
- Terri Fairley
- Ann Richard
- Kathy Coutros
- Maureen Gwinn
- Sandy Roberts
- [Russell Thomas](#)
- [Logan Everett](#)
- [Imran Shah](#)
- [Richard Judson](#)
- Derik Haggard
- Thomas Sheffield
- Joseph Bundy
- Woody Setzer
- Katie Paul Friedman
- John Wambaugh



National Toxicology Program
U.S. Department of Health and Human Services

- Scott Auerbach

Bio:Spyder™

- Jo Yeakley
- Bruce Seligmann
- Joel McComb
- Pete Shepherd
- Milos Babic
- Dalia Gonzalez
- Kyle LeBlanc
- Garrett McComb