# Microphysiological Models: Tools and Applications for Developmental Toxicology

## Sid Hunter

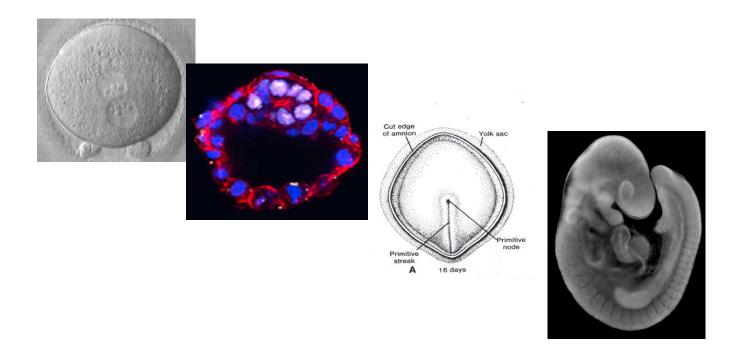
## Biomolecular and Computational Toxicology Division Center for Computational Toxicology and Exposure ORD/ US EPA



Disclaimer: The views and opinions in this presentation are those of the presenter and does not represent US EPA policy. Mention of trade names or commercial products does not constitute endorsement or recommendation for use.

Sid Hunter has no financial conflict of interest

### From the time of fertilization through organogenesis the conceptus undergoes profound structural and functional changes



Ebner et al., Figure 1B

Bedzhov and Zernicka-Goetz, Figure 3A

Sadler, Medical Embryology, Figure 5-1

Hunter. Original

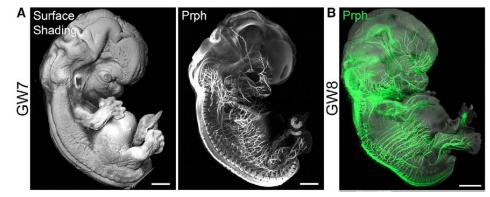


Hypothesis: Understanding the causes and prevention of birth defects requires advancing models for human biology-based assessments and decisions

Understanding the complex genesis of human birth defects will require models that recapitulate the physiological environment of pregnancy, mimic chemical exposure and the consequences of those exposures on developing cells, tissues and organs.



http://www.cdc.gov/ncbddd/birthdefects/index.html



Belle et al., 2017, Cell 169, 161-173

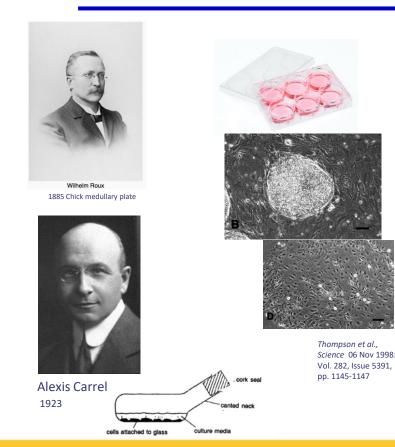


Microphysiological System (MPS): cell culture systems replicating (patho)physiology through engineered organ architecture and functionality. This includes especially 3D-(co-)cultures such as organoids, organ-on-chip models, and multi-organ models, as well as the technologies to engineer and analyze these systems.

> MICROPHYSIOLOGICAL SYSTEMS WORLD SUMMIT May 30<sup>th</sup> – June 3<sup>rd</sup>, 2022

https://mpsworldsummit.com

### Incredible Advancements in the Tools and Approaches to Cell Culture

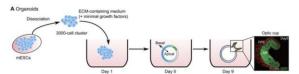


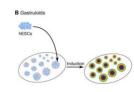


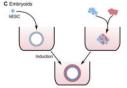
https://www.thescientist.com/image-of-theday/image-of-the-daymonkeying-around-34841

UNIVERSITY OF WISCONSIN-MADISON, <u>S.C. VERMILYEA, S. GUTHRIE, T.G.</u> <u>GOLOS, M.E. EMBORG</u>

Self-organization into organoids, gastruloids and embryoids.





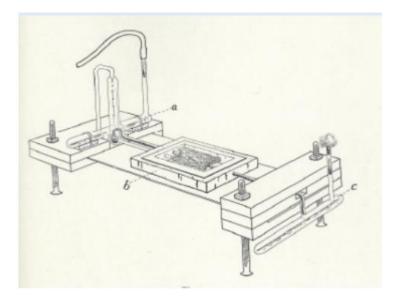


Mijo Simunovic, and Ali H. Brivanlou Development 2017:144:976-985

### Development of continuous flow hanging drop culture device - 1912



Montrose T Burrows



A method of furnishing a continuous supply of new medium to a tissue culture  $\ensuremath{\mathsf{In}}\xspace{\mathsf{Vitro}}^\dagger$ 

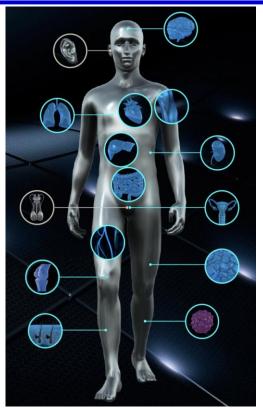
#### Montrose T. Burrows

First published:March 1912 | https://doi.org/10.1002/ar.1090060307 | Citations: 21

<sup>†</sup> Read before the American Association of Anatomists, December 27, 1911, at Princeton, N. J.

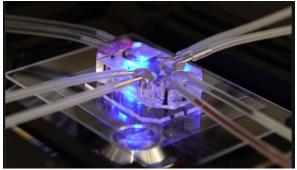


### Organ-on-Chip, Microphysiological Models are being developed for many organs



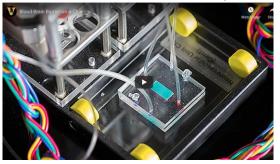
https://ncats.nih.gov/tissuechip/chip

Lung on a Chip



wyss.harvard.edu

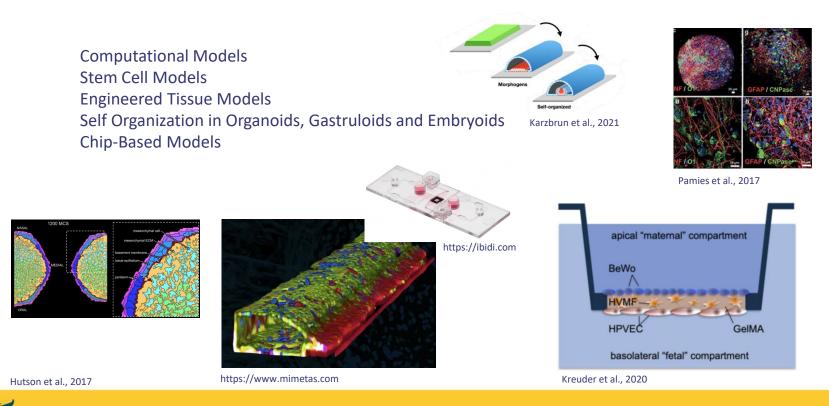
NeuroVascular Unit Chip



https://news.vanderbilt.edu/2016/12/06/ blood-brain-barrier-on-a-chip-sheds-new-light-on-silent-killer/

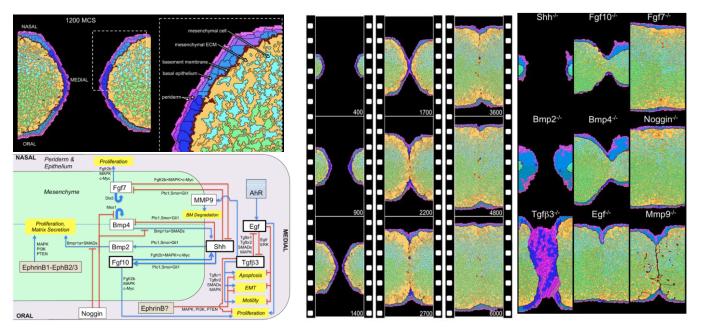


### Many MPS models can be used to assess developmental toxicity



### **Computational Models**

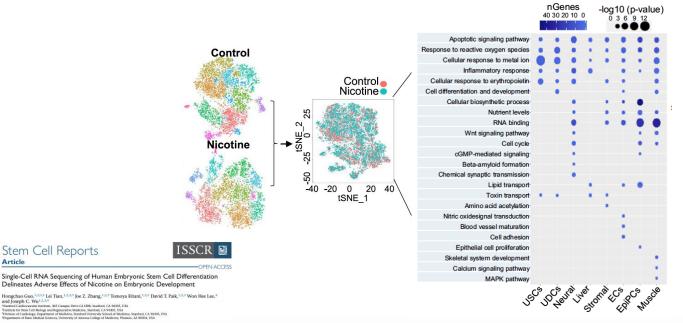
Models based upon cellular and molecular relationships driving cellular responses Tissue models – Limb, palate, neurovascular unit, vasculogenesis, somite formation Enables hypothesis building and testing. Cybermorphs



https://pubs.acs.org/doi/pdf/10.1021/acs.chemrestox.6b00350

## Pluripotent Stem Cell Models

Focus on differentiation, cytotoxicity, proliferation Directly plated, embryoid body, spheroid culture models Directed or not-directed differentiation Endpoints: Morphological, metabolomic, differentiation-marker expression Endpoints: Single feature, many features, single cell approach



Society for Birth Defects Research and Prevention 62<sup>nd</sup> Annual Meeting

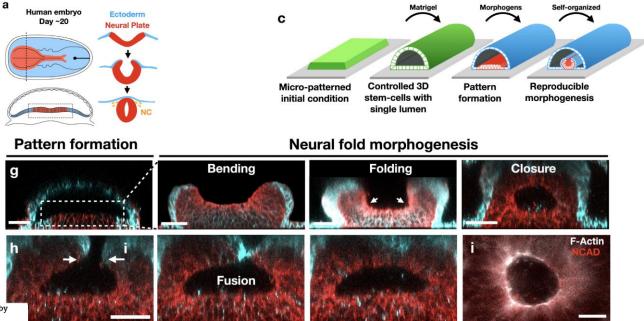
Stem Cell Reports

Article

and Joseph C. Wu12.3.\*

### **Engineered Models**

### Morphological Models of Tissues or Processes Embryoid Bodies, Bioprinted, Geometrically Constrained

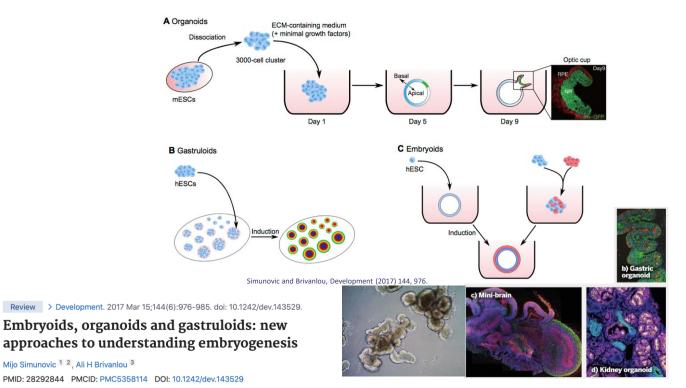


> Nature. 2021 Nov;599(7884):268-272. Human neural tube morphogenesis in vitro by geometric constraints

Eyal Karzbrun <sup>#</sup> 1 <sup>2</sup>, Almal H Khankhel <sup>#</sup> <sup>3</sup>, Heitor C Megale <sup>4</sup>, Stella M K Glasauer <sup>5</sup> <sup>6</sup>, Yofel Wyle <sup>5</sup>, George Britton <sup>7</sup>, Aryeh Warmflash <sup>®</sup> <sup>9</sup>, Kenneth S Kosik <sup>®</sup> 6, Eric D Siggla <sup>10</sup>, Boris I Shraiman <sup>4</sup> <sup>11</sup>, Sebastian J Streichan <sup>12</sup> <sup>13</sup> PMID: 34707280 PMCID: PMC6828633 DOI: 101038/s41586-021-04026-9

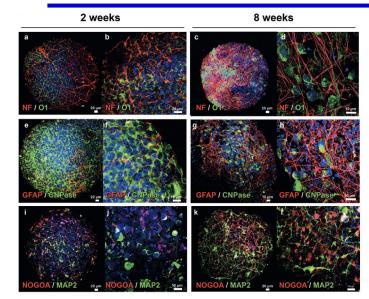


### Self Organization in Organoids, Gastruloids and Embryoids Models



https://www.rsb.org.uk/biologist-features/fromorganoids-to-gastruloids

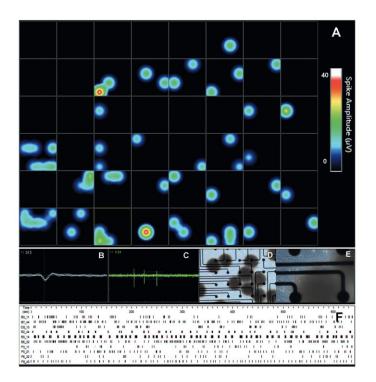
### Human Neural Organoid Models



> ALTEX. 2017;34(3):362-376. doi: 10.14573/altex.1609122. Epub 2016 Nov 24.

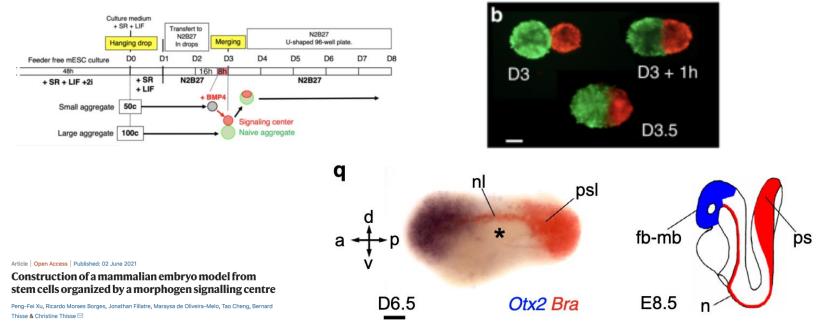
A human brain microphysiological system derived from induced pluripotent stem cells to study neurological diseases and toxicity

David Pamies <sup>1</sup>, Paula Barreras <sup>2</sup> <sup>3</sup>, Katharina Block <sup>1</sup>, Georgia Makri <sup>2</sup> <sup>4</sup>, Anupama Kumar <sup>2</sup> <sup>3</sup>, Daphne Wiersma <sup>1</sup>, Lenna Smirnova <sup>1</sup>, Ce Zang <sup>2</sup> <sup>4</sup>, Joseph Bressler <sup>5</sup>, Kimberly M Christian <sup>2</sup> <sup>4</sup>, Georgina Harris <sup>1</sup>, Guo-Li Ming <sup>2</sup> <sup>4</sup> <sup>6</sup>, Cindy J Berlinicke <sup>7</sup>, Kelly Kyno<sup>8</sup>, Hongjun Song <sup>2</sup> <sup>4</sup> <sup>6</sup>, Carlos A Pardo <sup>2</sup> <sup>3</sup>, Thomas Hartung <sup>1</sup> <sup>9</sup>, Helena T Hogberg <sup>1</sup> PMID: 27883356 PMCID: PMC6047513 DOI: 10.14573/altex.1609122



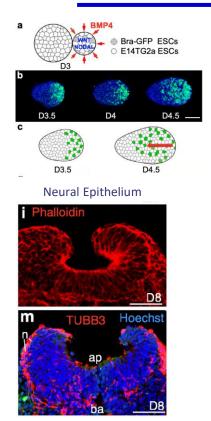
### Embryoid Model

Morphological models of tissues or processes Embryoid body, bioprinted organization, geometrically constrained

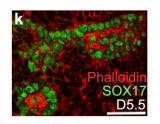


Nature Communications 12, Article number: 3277 (2021) | Cite this article

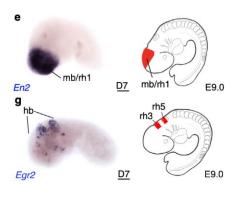
### Embryoid Model

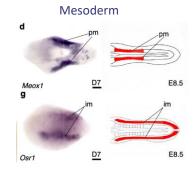


### Endoderm

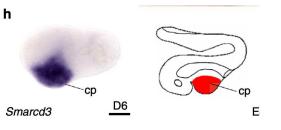


### Neuroectoderm





Cardiac



#### Article | Open Access | Published: 02 June 2021

Construction of a mammalian embryo model from stem cells organized by a morphogen signalling centre

Peng-Fei Xu, Ricardo Moraes Borges, Jonathan Fillatre, Maraysa de Oliveira-Melo, Tao Cheng, Bernard Thisse & Christine Thisse ⊠

Nature Communications 12, Article number: 3277 (2021) | Cite this article

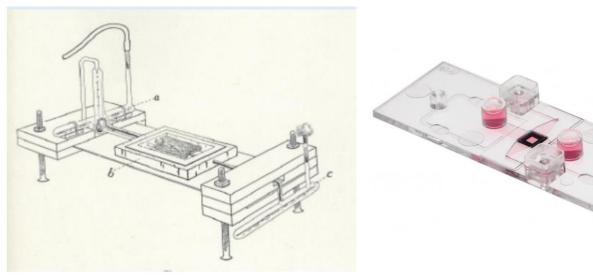


### Tissue Chip-based Model

Constructed models that mimic tissue-level organization and physiology Recapitulate a tissue Establish progenitor model for morphogenesis

1912

2022



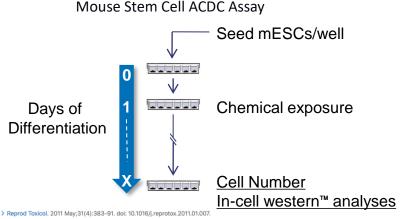
Burrows, 1912

https://ibidi.com



Application of Complex Models to Identify Putative Developmental Toxicants

70-90% of chemicals that affect stem cell endpoints are developmental toxicants in vivo
HOWEVER 50 – 60% of developmental toxicants do not affect the stem cell endpoints evaluated



Mouse embryonic stem cell adherent cell differentiation and cytotoxicity (ACDC) assay

Marianne Barrier<sup>11</sup>, Susan Jeffay, Harriette P Nichols, Kelly J Chandler, Maria R Hoopes Kimberly Slentz-Kesler, E Sidney Hunter 3rd PMID: 21296659 DOI: 10.1016/j.reprotox.2011.01.007 Why did so may chemicals not affect stem cells endpoints?

- Differentiation Hypothesis
- Developmental Process Hypothesis
- Pregnancy Triad System Hypothesis Developmental toxicity can be produced by adverse effects on the embryo, mother, or placenta. A pregnancy systems model will be necessary to accurately identify all potential developmental toxicants.

### There is increasing evidence of developmental consequences of placentation defects

### Placentation defects are highly prevalent in embryonic lethal mouse mutants

Vicente Perez-Carcial-<sup>2</sup>a, Elena Finebergh<sup>2,2</sup>a, Robert Wilson<sup>3</sup>, Alexander Murray<sup>1,2</sup>, Coclina Icoresi Mazzeo<sup>4</sup>, Catherine Tudor<sup>4</sup>, Arnold Sienerth<sup>1,2</sup>, Jacqueline K. White<sup>4</sup>, Elizabeth Tucle<sup>4</sup>, Edward J. Ryder<sup>4</sup>, Diane Gleeson<sup>4</sup>, Emma Siragher<sup>4</sup>, Hannah Wardle-Jones<sup>4</sup>, Nicole Staudt<sup>4</sup>, Neha Walt<sup>4</sup>, John Collins<sup>4</sup>, Stefan Geyer<sup>3</sup>, Elisabeth M. Busch-Nentwich<sup>4,6</sup>, Antonella Gall<sup>4</sup>, James C. Smith<sup>3</sup>, Elizabeth Robertson<sup>7</sup>, David J. Adams<sup>4</sup>, Wolfgang J. Weninger<sup>3</sup>, Timothy Mohun<sup>3</sup> & Myriam Hemberger<sup>1,2</sup>

Abnormal placenta assoicated dysmorphologies	% difference	р value
Growth/size phenotype	31.28	1.93E-06
Nervous system development	22.03	1.26E-06
Brain morphology	29.17	2.49E-04
Heart morphology	31.22	5.18E-05
Blood vessel morphology	23.62	3.17E-03
Subcutaneous edema	22.25	8.64E-03



doi:10.1038/nature26002

RESEARCH ARTICLE CELL BIOLOGY

OPEN ACCESS

## Inefficient development of syncytiotrophoblasts in the *Atp11a*-deficient mouse placenta

Yuki Ochiai<sup>a</sup>, Chigure Suzuki<sup>b,c</sup>, Katsumori Segawa<sup>a,1</sup>, Yasuo Uchiyama<sup>b,c</sup>, and Shigekazu Nagata<sup>a,d,2</sup> 💿

Contributed by Shigekazu Nagata; received January 12, 2022; accepted March 22, 2022; reviewed by Raymond Birge and Yasushi Hirota

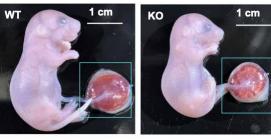
### SCIENCE ADVANCES | RESEARCH ARTICLE

### CELL BIOLOGY

Modified from Zhang

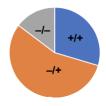
## TMEM16F phospholipid scramblase mediates trophoblast fusion and placental development

Yang Zhang<sup>1,2</sup>, Trieu Le<sup>1</sup>, Ryan Grabau<sup>3</sup>, Zahra Mohseni<sup>4</sup>, Hoejeong Kim<sup>5</sup>, David R. Natale<sup>6</sup>, Liping Feng<sup>4,7</sup>, Hua Pan<sup>3</sup>, Huanghe Yang<sup>1,2</sup>\*



Postnatal Genotype

Atn1

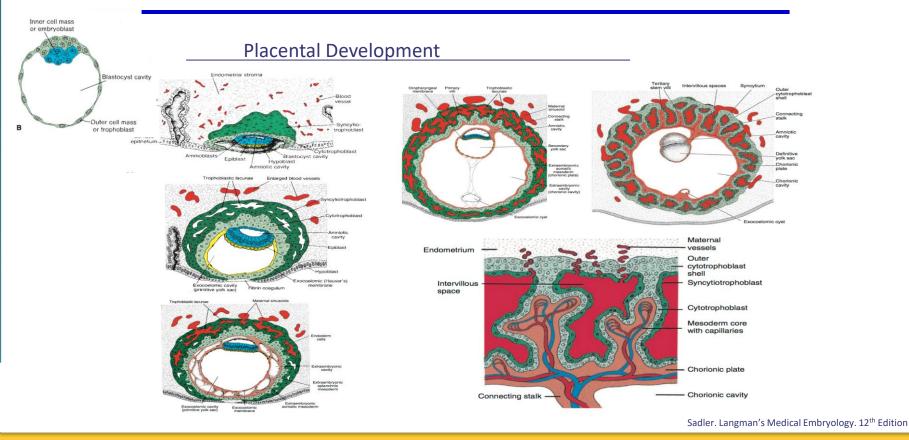


Modified from Zhang

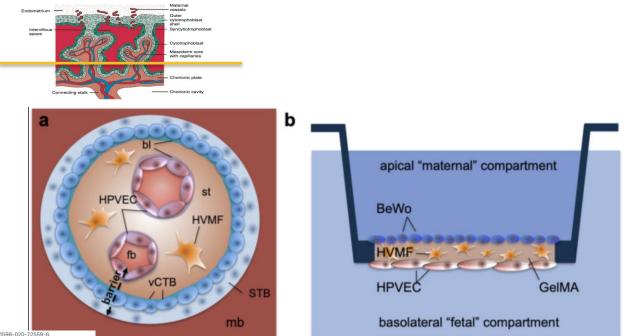


Modified from Perez-Garcia

### At the time of Implantation, the conceptus consists of the inner cell mass and the surrounding trophoblast layer



### Placental Models for Developmental Toxicity studies



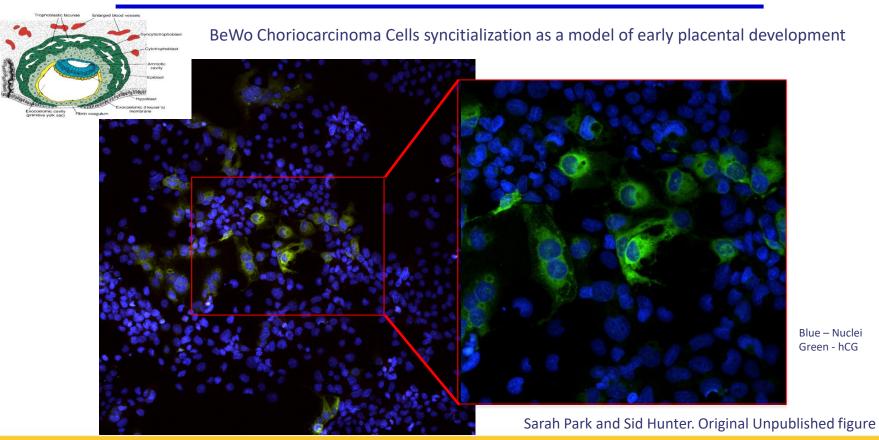
> Sci Rep. 2020 Sep 24;10(1):15606. doi: 10.1038/s41598-020-72559-6.

Inspired by the human placenta: a novel 3D bioprinted membrane system to create barrier models

Anna-Elisabeth Kreuer <sup>1</sup> <sup>2</sup>, Aramis Bolaños-Rosales <sup>3</sup> <sup>4</sup>, Christopher Palmer <sup>4</sup>, Alexander Thomas <sup>3</sup> <sup>4</sup>, Michel-Andreas Geiger <sup>4</sup>, Tobias Lam <sup>4</sup>, Anna-Klara Amler <sup>3</sup> <sup>4</sup>, Udo R Marker <sup>6</sup>, Roland Lawster <sup>3</sup>, Lutz Kloke <sup>6</sup>

https://www.ted.com/talks/anna\_elisabeth\_kreuder\_bioprinting\_the\_human\_placenta

### Placental Models for Developmental Toxicity studies



Ø

### Chemicals affect Cytotrophoblasts and mouse Stem Cells

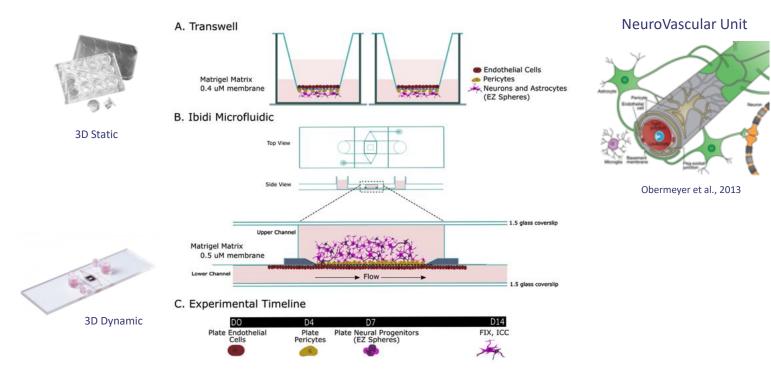
Chemical	Cytotrophoblast Cell Death (LDH)	Cytotrophoblast Live Cell (MTS)	Cytotrophoblast (hCG)	mESC ACDC Assessment GSC Differentiation	mESC ACDC Assessment Cytotoxicity
5-Fluorouracil	10	10	10	0.7	0.5
6-aminonicotinamide	1	1	1	8	8
Tributyltin	0.5	0.5	0.5	0.02	0.2
Diphenhydramine HCl	100	-	-	-	
Diphenylhydantoin	-	-	-	-	-
Sulfasalazine	-	-	-	-	-
Carbamazepine	-	-	-	-	-
Busulfan	-	-	-	20	-
Tebuconazole	100	-	30	-	-
Butylparaben	100	100	100	-	-
Zearalenone	100	100	30	No Data	No Data
TBBPA	-	-	100	No Data	No Data
Braga (TMEM16F)					
Triclosan	30	100	30	20.4	20.4
Simvastatin	30	30	2	-	-
Promethazine	30	30	30	No Data	No Data
Paroxetine	10	30	30	No Data	No Data

- No Effect, Chemical Concentrations (µM)

Sarah Park and Sid Hunter. Original Unpublished Data

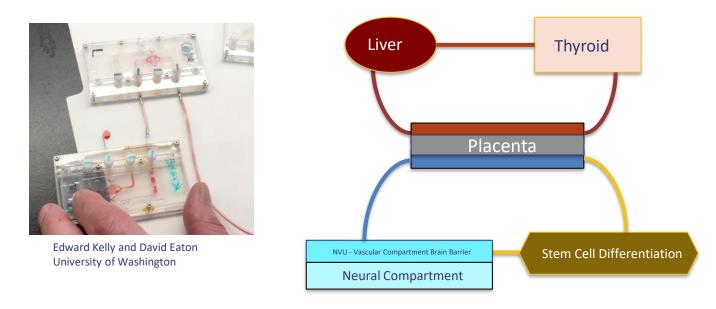


## Organotypic Culture model of Early Human Brain development



Andrew Schwab et al., Submitted for publication

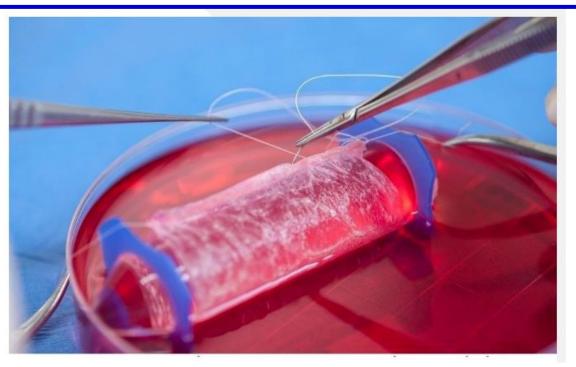
### Next Generation: Integrated Systems Model



### Multi-chip dual flow system design

### V. Richardson - Original Image

### Tissue-Engineered Uterus Supports Live Birth in Rabbits

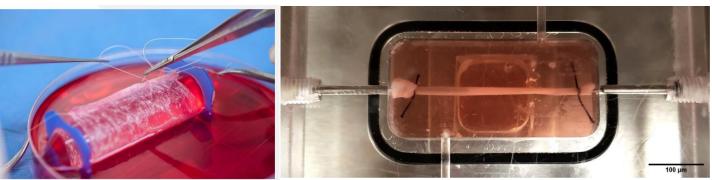


https://health-innovations.org/2020/07/03/a-tissue-engineered-uterus-supports-live-births-in-rabbits/

Magalhaes, R.S., Williams, J.K., Yoo, K.W. et al. A tissue-engineered uterus supports live births in rabbits. Nat Biotechnol (2020). https://doi.org/10.1038/s41587-020-0547-7

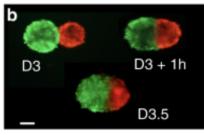


Is it possible that combining the tools used to create blood vessels with tissue engineering approach for a uterus could establish an in vitro model for pregnancy model and embryonic development



https://health-innovations.org/2020/07/03/a-tissue-engineered-uterus-supports-live-births-in-rabbits/

https://bme.duke.edu/about/news/engineered-living-cell-blood-vessel-provides-new-insights-progeria



https://www.nature.com/articles/s41467-021-23653-4

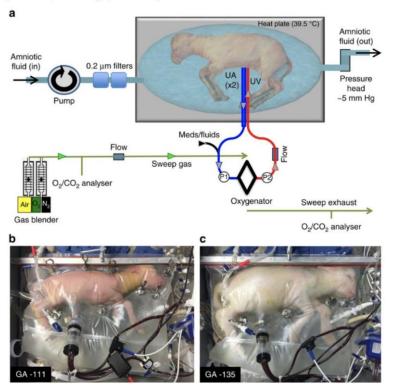


Hunter. Original



### Aldous Huxley's Brave New World???

#### Figure 1: UA/UV Biobag system design.



#### Open Access | Published: 25 April 2017

#### An extra-uterine system to physiologically support the extreme premature lamb

Emily A. Partridge, Marcus G. Davey, Matthew A. Hornick, Patrick E. McGovern, All Y. Mejaddam, Jasse D. Vrecenak, Carmen Mesas-Burgos, Aliza Olive, Robert C. Caskey, Theodore R. Weiland, Jancheng Han, Alexander J. Schupper, James T. Connelly, Kevin C. Dysart, Jack Rychik, Holly L. Hedrick, William H. Peranteau & Alan W. Flake 🖂

Nature Communications 8, Article number: 15112 (2017) Cite this article



## What opportunities will come today!

## Hunter. Original

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Pamies, D., Barreras, P., Block, K., Makri, G., Kumar, A., Wiersma, D., Smirnova, L., Zhang, C., Bressler, J., Christian, K. M., Harris, G., Ming, G.- Ii, Berlinicke, C. J., Kyro, K., Song, H., Pardo, C. A., Hartung, T. and Hogberg, H. T. (2017) *ALTEX - Alternatives to animal experimentation*, 34(3), 362–376. doi: 10.14573/altex.1609122.

Karzbrun, E., Khankhel, A.H., Megale, H.C. Glasauer, S.M.K. Wyle, Y. Britton, G., Warmflash, A. Kosik, K.S., Siggia, E.D., Shraiman, B.I., Streichan, S.J. (2021). Nature 599(7884):268-272. doi: 10.1038/s41586-021-04026-9.

Schwab, A.J., Jeffay, S.C., Nichols, H.P., Hunter, 3rd, E.S. (2022) Submitted for publication

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Obermeier, B., Daneman, R. & Ransohoff, R. Development, maintenance and disruption of the blood-brain barrier. Nat Med 19, 1584–1596 (2013). https://doi.org/10.1038/nm.3407

Magalhaes, R.S., Williams, J.K., Yoo, K.W. et al. A tissue-engineered uterus supports live births in rabbits. Nat Biotechnol (2020). https://doi.org/10.1038/s41587-020-0547-7

Bedzhov, I., Zernicka-Goetz, M. (2014). Self-Organizing Properties of Mouse Pluripotent Cells Initiate Morphogenesis upon Implantation. Cell. 156: 1032–1044

Deglinverti, A, Croft, GF., Pietila, LN, Zernicka-Goetz, M., Siggia, ED, Brivanlou, AH. (2016). Self-Organization of the In Vitro Attached Human Embryo. Nature. 533: 251–254.





**Reviews:** 

Sonia Youhanna, Aurino M. Kemas, Lena Preiss, Yitian Zhou, Joanne X. Shen, Selgin D. Cakal, Francesco S. Paqualini, Sravan K. Goparaju, Reza Zandi Shafagh, Johan Ulril Lind, Carl M. Sellgren and Volker M. Lauschke. Organotypic and Microphysiological Human Tissue Models for Drug Discovery and Development—Current State-of-the-Art and Future Perspectives Pharmacological Reviews January 1, 2022, 74 (1) 141-206; DOI: <a href="https://doi.org/10.1124/pharmrev.120.000238">https://doi.org/10.1124/pharmrev.120.000238</a>

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Truskey George A. (2018). Human Microphysiological Systems and Organoids as in Vitro Models for Toxicological Studies. Frontiers in Public Health, DOI=10.3389/fpubh.2018.00185

Luz AL, Tokar EJ. Pluripotent Stem Cells in Developmental Toxicity Testing: A Review of Methodological Advances. Toxicol Sci. 2018 Sep 1;165(1):31-39. doi: 10.1093/toxsci/kfy174. PMID: 30169765; PMCID: PMC6111785.

