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PBPK and Toxicokinetic Modeling

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- Generic vs. bespoke PBTK models
- High Throughput Toxicokinetics (HTTK)
- Model parameterization
 - Physiologic parameters
 - Chemical-specific parameters
- Model evaluation



Estimating Chemical Risk

- NRC (1983): Risk is a function of inherent chemical hazard, extent of exposure, and the dose-response relationship (including toxicokinetics)
- High throughput risk prioritization based upon *in vitro* screening requires comparison to exposure
- Data obtained *in vitro* must be placed in an *in vivo* context: *in vitro-in vivo* extrapolation (IVIVE)
- Information must be relevant to the scenario, for example, consumer, ambient, or occupational exposure.





Toxicokinetics

 Toxicokinetics describes the absorption, distribution, metabolism, and excretion of a chemical by the body:
Chemical-specific





In Vitro-In Vivo Extrapolation (IVIVE)

 Translation of *in vitro* high throughput screening requires chemical-specific toxicokinetic models for anywhere from dozens to thousands of chemicals



Breen *et al.* (2021)



Generic vs. bespoke PBTK models



Everyone Uses Models

- Toxicology has long relied upon model animal species
- People rely on mental models every day
 - For example, with repetitive activities like driving home from work
- Mathematical models offer some significant advantages:
 - Reproducible
 - Can (and should) be transparent
- ...with some disadvantages:
 - Sometimes reality is complex
 - Sometimes the model doesn't always work well
 - How do we know we can extrapolate?
- ...that can be turned into advantages:
 - If we have evaluated confidence/uncertainty and know the "domain of applicability" we can make better use of mathematical models





Fit for Purpose Models

• A "fit for purpose" model is an abstraction of a complicated problem that allows us to reach a decision.

"Now it would be very remarkable if any system existing in the real world could be *exactly* represented by any simple model. However, cunningly chosen parsimonious models often do provide remarkably useful approximations... **The only question of interest is 'Is the model illuminating and useful?'**" George Box

- A fit for purpose model is defined as much by what is omitted as what is included in the model.
- We must accept that there will always be areas in need of better data and models our knowledge will always be incomplete, and thus we wish to extrapolate.
 - How do I drive to a place I've never been before?



Fit for Purpose Toxicokinetics

- Chiu et al. (2007) "...[P]arsimony in selecting model structures is an important and guiding principle in developing models for use in risk assessments."
 - Complexity is constrained by limited data available to calibrate and test the model and the need to justify both the model assumptions and predictions
- Bessems et al. (2014): We need "a first, relatively quick ('Tier 1'), estimate" of concentration vs. time in blood, plasma, or cell
 - They suggested that we neglect active metabolism. But thanks to *in vitro* measurements we can now do better
 - We still neglect transport and other protein-specific phenomena





Bespoke vs. Generic

Bespoke, Tailored, Custom... *Requires specific measurements*



Generic, Off-the-Shelf/Rack, One-Size-Fits-Most *Approximately fits certain categories*





11 of 44

Bespoke Models

- Toxicokinetic (and pharmacokinetic) models are traditionally developed using *in vivo* data
 - These data could be from clinical trials (increasing the relevance but limiting the measurements) or possibly animal studies (allowing tissues to be sampled)
 - Potentially resource-intensive
- Physiologically-based toxicokinetic (PBTK) models allow extrapolation between species and routes of administration
 - Physiological information augments chemicalspecific data
- Can choose to make the complexity of the model and the number of physiological processes appropriate given the data and the decision context
 - This is how we "tailor" the model





Jones et al., 2012 PK of Statins In this case they had transporterspecific data



2. Schematic diagram of the in vivo PBPK model. EC, extracellular; IC, intracellular



Generic Models

- A standardized physiology is assumed, regardless of chemical:
 - The same parameters such as volumes, flows, and rates are used
 - The same processes are included (hepatic metabolism, glomerular filtration) or omitted
- A fixed set of descriptors (such as rate of metabolism and protein binding) are varied from chemical to chemical and potentially measured in vitro
- The generic model is implemented once, reducing the likelihood of coding errors and enhancing documentation
- We can estimate the accuracy of a generic model for a new chemical using performance across multiple chemicals where data happen to exist
- The generic model is a hypothesis
 - If we have evaluation data then we can check if we need to elaborate the model (for example, create a bespoke model)





Generic Models as a Hypothesis



- For pharmaceuticals, *in vitro* data plus a model including hepatic metabolism and passive glomerular filtration (kidney) are often enough to make predictions within a factor of 3 of *in vivo* data (Wang, 2010)
- For other chemicals there may be complications, for example, there is thought to be (Andersen et al. 2006) active transport of some per- and polyfluorinated alkyl substances (PFAS) in the kidney
- We could add a renal resorption process (that is, modify the generic model) only if there was some way to parameterize the process for most chemicals – otherwise we are back to tailoring the model to a chemical



Generic PBTK Models

The idea of generic PBTK has been out there for a while...





High Throughput Toxicokinetics



300

Most Chemicals Lack Toxicokinetic Data

- Most non-pharmaceutical chemicals for example, flame retardants, plasticizers, pesticides, solvents do not have human *in vivo* TK data.
 - Non-pesticidal chemicals are unlikely to have any *in vivo* TK data, even from animals



Figure from Bell et al. (2018)



HTTK: A NAM for Exposure

- In vitro high throughput toxicokinetic (HTTK) methods can provide toxicokinetic data for larger numbers of chemicals (for example, Rotroff et al., 2010, Wetmore et al., 2012)
- HTTK methods have been used by the pharmaceutical industry to determine range of efficacious doses and to prospectively evaluate success of planned clinical trials (Jamei, et al., 2009; Wang, 2010)
- The primary goal of HTTK is to provide a human dose context for bioactive concentrations from high throughput screening (that is, in vitro-in vivo extrapolation, or IVIVE) (for example, Wetmore et al., 2015)
- A secondary goal is to provide open-source data and models for evaluation and use by the broader scientific community (Pearce et al, 2017)



In Vitro-In Vivo Extrapolation (IVIVE)

Translation of *in vitro* high throughput screening requires chemical-specific toxicokinetic models for anywhere from dozens to thousands of chemicals





In vitro toxicokinetic data





In vitro toxicokinetic data

Typically, intrinsic hepatic clearance and fraction unbound in plasma



Rotroff et al. (2010) Wetmore et al. (2012) Wetmore et al. (2015) Wambaugh et al. (2019)



In vitro toxicokinetic data + generic toxicokinetic model



Rotroff et al. (2010) Wetmore et al. (2012) Wetmore et al. (2015) Wambaugh et al. (2019)



In vitro toxicokinetic data + generic toxicokinetic model



Rotroff et al. (2010) Wetmore et al. (2012) Wetmore et al. (2015) Wambaugh et al. (2019) Wambaugh et al. (2015) Pearce et al. (2017) Ring et al. (2017) Linakis et al. (2020)



In vitro toxicokinetic data + generic toxicokinetic model = high(er) throughput toxicokinetics





New HT-PBTK Models





Model parameterization



Model Parameters

Chemical-specific parameters	
Intrinsic hepatic clearance rate (CL _{int}) Fraction unbound to plasma protein (F _{up})	Measured in HT <i>in vitro</i> assays (Rotroff <i>et al.</i> 2010; Wetmore <i>et al.</i> 2012, 2014, 2015; Wambaugh <i>et al.</i> 2019) or predicted <i>in silico</i> (Sipes <i>et al.</i> 2017)
Tissue:blood partition coefficients (for compartmental models)	Predict from phys-chem properties and tissue properties (Pearce et al., 2017)
Physiological parameters	
Tissue masses (including body weight)	
Tissue blood flows	Gathered from data available in the
Glomerular filtration rate (passive renal clearance)	published literature [Wambaugh et al. 2015; Pearce et al. 2017a]
Hepatocellularity	

SEPA United States Environmental Protection Chemical-Specific In Vitro Measurements for TK



In drug development, HTTK methods allow IVIVE to estimate therapeutic doses for clinical studies – predicted concentrations are typically on the order of values measured in clinical trials (Wang, 2010)



PBTK Partition Coefficients

- Although in our model there are really three separate concentrations (C) that describe a tissue, we assume that they are related to each other by constants
- We assume that the ratio between the blood and plasma (*R_{blood:plasma}*) is a uniform constant throughout the body

 $C_{compartment, blood} = R_{blood: plasma} C_{compartment, plasma}$

We assume that all the tissues are "perfusion limited", which means that the tissue concentration instantly comes to equilibrium with the free fraction in plasma (concentration is limited by flow to the tissue)

 $C_{compartment,tissue} = K_{tissue:plasma} * f_{up} * C_{compartment,plasma}$

K_{tissue:plasma} is the tissue partition coefficient which we either measure experimentally or predict *in silico* (*for example* Schmitt's method)





Development of Analytical Chemistry Method is a Rate Limiting Step

- The HTTK in vitro assays need to measure differences in chemical concentration
- Historically, methods cannot be developed for all chemicals
- Different chemicals require different instruments and methods, for example liquid vs. gas chromatography mass spectrometry (LC vs. GC MS)





Development of Analytical Chemistry Method is a Rate Limiting Step

- The HTTK in vitro assays need to measure differences in chemical concentration
- Area of the internal standard (ITSD) at a known, fixed concentration fluctuates with time, depends on instrument and methodology
- Analytical chemist must find a peak that corresponds to chemical of interest, and then follow the ratio R of the chemical peak to the ITSD
- Ability to resolve peak depends on the matrix (for example, blood vs. DMSO)



Model parameters are either:

Physiological: determined by species and potentially varied via Monte Carlo (including HTTK-pop, Ring et al. 2017) Chemical-specific: physicochemical properties (Mansouri et al., 2018) and equilibrium partition coefficients plus plasma binding and metabolism rates that are determined from *in vitro* measurements or potentially predicted from structure

Parameter	Definition	Value (Mean)	Units	Reference
Q _{liverc}	Total blood flow to liver (arterial, gut)	3.6	1/h/kg BW	Davies and Morris (1993)
Q _{GFR}	Flow to glomerulus (glomerular filtration rate)	0.32	1/h/kg BW	Davies and Morris (1993)
n _{cell_density}	Hepatocellularity	110	Millions of cells / g Liver	Carlile et al. (1997)
V _{liverc}	Liver volume	0.0245	1/kg BW	Davies and Morris (1993)
d _{liver}	Liver density	1.05	g/ml	International Commission on Radiological Protection (1975)
Hematocrit	Fraction of blood that is red blood cells	0.43	Unitless	Davies and Morris (1993)
C _{protein}	Concentration of protein used in f _{up} assay	5	μΜ	Wambaugh et al. (2019)

 $Cl_{hepatic} = n_{cell \ density} \times V_{liverc} \times d_{liver} \times Cl_{int}$



Species-Specific Physiological Parameters for Physiologically-Based Toxicokinetics

Rates, volumes, and tissue-specific information (not shown) are needed for a species

Parameter	Units	Mouse	Rat	Dog	Human	Rabbit	Monkey
Total Body Water	ml/kg	725.000	668.000	603.600	600.000	40.812	693.000
Plasma Volume	ml/kg	50.000	31.200	51.500	42.857	110.000	44.800
Cardiac Output	ml/min/kg^(3/4)	150.424	209.304	213.394	231.401	266.576	324.790
Average BW	kg	0.020	0.250	10.000	70.000	2.500	5.000
Total Plasma Protein	g/ml	0.062	0.067	0.090	0.074	0.057	0.088
Plasma albumin	g/ml	0.033	0.032	0.026	0.042	0.039	0.049
Plasma a-1-AGP	g/ml	0.013	0.018	0.004	0.002	0.001	0.002
Hematocrit	fraction	0.450	0.460	0.420	0.440	0.360	0.410
Urine Flow	ml/min/kg^(3/4)	0.013	0.098	0.037	0.040	0.042	0.151
Bile Flow	ml/min/kg^(3/4)	0.026	0.044	0.015	0.010	0.083	0.004
GFR	ml/min/kg^(3/4)	5.265	3.705	10.901	5.165	3.120	2.080
Average Body Temperature	С	37.000	38.700	38.900	37.000	39.350	38.000
Plasma Effective Neutral Lipid Volume Fraction	unitless	0.004	0.002	0.001	0.007	0.002	0.007
Plasma Protein Volume Fraction	unitless	0.060	0.059	0.090	0.070	0.057	0.070
Pulmonary Ventilation Rate	l/h/kg^(3/4)	24.750	24.750	24.750	27.750	24.750	27.750
Alveolar Dead Space Fraction	unitless	0.330	0.330	0.330	0.330	0.330	0.330

• Davies, Brian, and Tim Morris. "Physiological parameters in laboratory animals and humans." Pharmaceutical research 10.7 (1993): 1093-1095.

• Brown, Ronald P., et al. "Physiological parameter values for physiologically based pharmacokinetic models." Toxicology and industrial health 13.4 (1997): 407-484.

• Birnbaum, L., et al. "Physiological parameter values for PBPK models." International Life Sciences Institute, Risk Science Institute, Washington, DC (1994).

• Robertshaw, D., Temperature Regulation and Thermal Environment, in Dukes' Physiology of Domestic Animals, 12th ed., Reece W.O., Ed. Copyright 2004 by Cornell University.

• Stammers, Arthur Dighton. "The blood count and body temperature in normal rats." The Journal of physiology 61.3 (1926): 329.

• Gordon, Christopher J. Temperature regulation in laboratory rodents. Cambridge University Press, 1993.

• Gauvin, David V. "Electrocardiogram, hemodynamics, and core body temperatures of the normal freely moving cynomolgus monkey by remote radiotelemetry", Journal of Pharmacological and Toxicological Methods



Tools for Chemical-Specific PBTK Parameters

Physiological parameters depend on species, but we must also make chemical-specific estimates of tissue partitioning...





Collaborative Evaluation of QSPRs (Quantitative Structure-Property Relationships) for HTTK

- Open-source QSPR predictions currently available for thousands of chemicals, including full Tox21 library
- EPA is leading an international collaborative evaluation of various QSPRs trained to both pharma and non-pharma chemicals for predicting HTTK







Model evaluation



Verifying PBTK Models

Process for the Evaluation of PBPK Models

- 1. Assessment of Model Purpose
- 2. Assessment of Model Structure and Biological Characterizations
- 3. Assessment of Mathematical Descriptions
- 4. Assessment of Computer Implementation
- 5. Parameter Analysis and Assessment of Model Fitness
- 6. Assessment of any Specialized Analyses

Clark et al. (2004)



FIG. 1. This figure shows examples of key considerations during model development, evaluation, and application that are necessary before a PBPK model may be adopted for use in a HHRA.

McLanahan et al. (2012)



- To evaluate a chemical-specific TK model for "chemical x" you can compare the predictions to *in vivo* measured data
 - Can estimate bias
 - Can estimate uncertainty
 - Can consider using model to extrapolate to other situations (dose, route, physiology) where you have no data





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- However, we do not typically have TK data
- We can parameterize a generic TK model, and evaluate that model for as many chemicals as we do have data
 - We do expect larger uncertainty, but also greater confidence in model implementation
 - Estimate bias and uncertainty, and try to correlate with chemical-specific properties



Cohen Hubal et al. (2019)



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Cohen Hubal et al. (2019)



41 of 44

Building Confidence in TK Models

- To evaluate a chemical-specific TK model for "chemical x" you can compare the predictions to *in vivo* measured data
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 - Can consider using model to extrapolate to other situations (chemicals without *in vivo* data)



Cohen Hubal et al. (2019)



Evaluation Example: Observed Total Clearance

- We estimate clearance from two processes – hepatic metabolism (liver) and passive glomerular filtration (kidney)
- This appears to work better for pharmaceuticals than other chemicals:
 - ToxCast chemicals are overestimated
- Non-pharmaceuticals may be subject to extrahepatic metabolism and/or active transport





CvTdb: An In Vivo TK Database

https://github.com/USEPA/CompTox-PK-CvTdb

- EPA has developed a public database of concentration vs. time data for building, calibrating, and evaluating TK models
- Curation and development is ongoing, but to date includes:
 - 198 analytes (EPA, National Toxicology Program, literature)
 - Routes: Intravenous, dermal, oral, sub-cutaneous, and inhalation exposure
- Standardized, open-source curve fitting software invivoPKfit used to calibrate models to all data:

https://github.com/USEPA/CompTox-ExpoCast-invivoPKfit







- Toxicokinetics links exposure with internal concentrations
- Physiologically-based toxicokinetic (PBTK) models allow extrapolation, including *in vitro-in vivo* extrapolation (IVIVE)
- PBTK models can be generic or bespoke
- Generic models allow for verification of model implementation
- High throughput toxicokinetics (HTTK) allow in vitro parameterization of generic PBTK models
- Comparing model predictions for chemicals with *in vivo* data allows estimation of confidence in predictions for chemicals without *in vivo* data

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References

- 1. Andersen, Melvin E., et al. "Pharmacokinetic modeling of saturable, renal resorption of perfluoroalkylacids in monkeys—probing the determinants of long plasma half-lives." *Toxicology* 227.1-2 (2006): 156-164.
- 2. Armitage, James M., et al. "Development and intercomparison of single and multicompartment physiologically-based toxicokinetic models: Implications for model selection and tiered modeling frameworks." *Environment International* 154 (2021): 106557.
- 3. Bell, Shannon, et al. "An integrated chemical environment with tools for chemical safety testing." *Toxicology in Vitro* 67 (2020): 104916.
- 4. Bessems, Jos G., et al. "PBTK modelling platforms and parameter estimation tools to enable animal-free risk assessment: recommendations from a joint EPAA–EURL ECVAM ADME workshop." *Regulatory Toxicology and Pharmacology* 68.1 (2014): 119-139.
- 5. Breen, Miyuki, et al. "High-throughput PBTK models for in vitro to in vivo extrapolation." *Expert Opinion on Drug Metabolism & Toxicology* just-accepted (2021).
- 6. Carlile, David J., Katayoun Zomorodi, and J. Brian Houston. "Scaling factors to relate drug metabolic clearance in hepatic microsomes, isolated hepatocytes, and the intact liver: studies with induced livers involving diazepam." *Drug metabolism and disposition* 25.8 (1997): 903-911.
- 7. Chiu WA, Barton HA, DeWoskin RS, Schlosser P, Thompson CM, Sonawane B, et al. Evaluation of physiologically based pharmacokinetic models for use in risk assessment. Journal of Applied Toxicology. 2007;27(3):218–37. pmid:17299829
- 8. Cho, A. K., et al. "Stereochemical differences in the metabolism of 3, 4-methylenedioxymethamphetamine in vivo and in vitro: a pharmacokinetic analysis." *Drug Metabolism and Disposition* 18.5 (1990): 686-691.
- 9. Clark, Leona H., R. Woodrow Setzer, and Hugh A. Barton. "Framework for evaluation of physiologically-based pharmacokinetic models for use in safety or risk assessment." *Risk Analysis: An International Journal* 24.6 (2004): 1697-1717.
- 10. Clewell III, Harvey J., et al. "Development of a physiologically based pharmacokinetic model of isopropanol and its metabolite acetone." *Toxicological Sciences* 63.2 (2001): 160-172.
- 11. Davies, Brian, and Tim Morris. "Physiological parameters in laboratory animals and humans." *Pharmaceutical research* 10.7 (1993): 1093-1095.
- 12. Eissing, Thomas, et al. "A computational systems biology software platform for multiscale modeling and simulation: integrating whole-body physiology, disease biology, and molecular reaction networks." *Frontiers in physiology* 2 (2011): 4.



References

- 13. Hubal, Elaine A. Cohen, et al. "Advancing internal exposure and physiologically-based toxicokinetic modeling for 21st-century risk assessments." *Journal of exposure science & environmental epidemiology* 29.1 (2019): 11-20.
- 14. International Commission on Radiological Protection. Report of the task group on reference man. Vol. 23. Pergamon, Oxford. 1975.121.
- 15. Jamei, Masoud, et al. "The Simcyp[®] population-based ADME simulator." *Expert opinion on drug metabolism & toxicology* 5.2 (2009): 211-223.
- 16. Jones, Hannah M., et al. "Mechanistic pharmacokinetic modeling for the prediction of transporter-mediated disposition in humans from sandwich culture human hepatocyte data." *Drug Metabolism and Disposition* 40.5 (2012): 1007-1017.
- 17. Jongeneelen, Frans J., and Wil F. Ten Berge. "A generic, cross-chemical predictive PBTK model with multiple entry routes running as application in MS Excel; design of the model and comparison of predictions with experimental results." *Annals of occupational hygiene* 55.8 (2011): 841-864.
- 18. Linakis, Matthew W., et al. "Development and evaluation of a high throughput inhalation model for organic chemicals." *Journal of exposure science & environmental epidemiology* 30.5 (2020): 866-877.
- 19. Lukacova, Viera, Walter S. Woltosz, and Michael B. Bolger. "Prediction of modified release pharmacokinetics and pharmacodynamics from in vitro, immediate release, and intravenous data." *The AAPS journal* 11.2 (2009): 323-334.
- 20. Mansouri, Kamel, et al. "OPERA models for predicting physicochemical properties and environmental fate endpoints." *Journal of cheminformatics* 10.1 (2018): 1-19.
- 21. McLanahan, Eva D., et al. "Physiologically based pharmacokinetic model use in risk assessment—why being published is not enough." *Toxicological Sciences* 126.1 (2012): 5-15.
- 22. Pearce, Robert G., et al. "Httk: R package for high-throughput toxicokinetics." *Journal of statistical software* 79.4 (2017a): 1.
- 23. Pearce RG, Setzer RW, Davis JL, Wambaugh JF. Evaluation and calibration of high-throughput predictions of chemical distribution to tissues. J Pharmacokinet Pharmacodyn. 2017b;44(6):549-65.
- 24. Pendse, Salil N., et al. "Population life-course exposure to health effects model (PLETHEM): An R package for PBPK modeling." *Computational Toxicology* 13 (2020): 100115. *armacology & Therapeutics* 18.4 (1975): 377-390.

25. Punt, Ans, et al. "Development of a Web-Based Toolbox to Support Quantitative In-Vitro-to-In-Vivo Extrapolations (QIVIVE) within Nonanimal Testing 46 of 44 Strategies." *Chemical research in toxicology* 34.2 (2020): 460-472.



References

- 26. Ring CL, Pearce RG, Setzer RW, et al. Identifying populations sensitive to environmental chemicals by simulating toxicokinetic variability. Environment International. 2017 2017/09/01/;106:105-118. Wetmore BA, Wambaugh JF, Ferguson SS, et al. Integration of dosimetry, exposure, and high-throughput screening data in chemical toxicity assessment. Toxicological Sciences. 2012 Jan;125(1):157-74.
- 27. Rotroff DM, Wetmore BA, Dix DJ, et al. Incorporating human dosimetry and exposure into high-throughput in vitro toxicity screening. Toxicological Sciences. 2010;117(2):348-358
- 28. Sayre, Risa R., John F. Wambaugh, and Christopher M. Grulke. "Database of pharmacokinetic time-series data and parameters for 144 environmental chemicals." *Scientific data* 7.1 (2020): 1-10.
- 29. Schmitt, Walter. "General approach for the calculation of tissue to plasma partition coefficients." *Toxicology in vitro* 22.2 (2008): 457-467.
- 30. Sipes NS, Wambaugh JF, Pearce R, et al. An Intuitive Approach for Predicting Potential Human Health Risk with the Tox21 10k Library. Environmental Science & Technology. 2017 2017/09/19;51(18):10786-10796.
- 31. Wambaugh, John F., et al. "Toxicokinetic triage for environmental chemicals." *Toxicological Sciences* 147.1 (2015): 55-67.
- 32. Wambaugh, John F., et al. "Evaluating in vitro-in vivo extrapolation of toxicokinetics." *Toxicological Sciences* 163.1 (2018): 152-169.
- 33. Wambaugh, John F., et al. "Assessing toxicokinetic uncertainty and variability in risk prioritization." *Toxicological Sciences* 172.2 (2019): 235-251.
- 34. Wang, Ying-Hong. "Confidence assessment of the Simcyp time-based approach and a static mathematical model in predicting clinical drug-drug interactions for mechanism-based CYP3A inhibitors." *Drug Metabolism and Disposition* 38.7 (2010): 1094-1104.
- 35. Wetmore BA, et al. Incorporating population variability and susceptible subpopulations into dosimetry for high-throughput toxicity testing. Toxicological Sciences. 2014 Nov;142(1):210-24
- 36. Wetmore BA, et al. Incorporating High-Throughput Exposure Predictions With Dosimetry-Adjusted In Vitro Bioactivity to Inform Chemical Toxicity Testing. Toxicological Sciences. 2015 Nov;148(1):121-36
- 37. Wilkinson, Grant R., and David G. Shand. "A physiological approach to hepatic drug clearance." *Clinical Pharmacology & Therapeutics* 18.4 (1975): 377-390.
- 38. World Health Organization. Characterization and application of physiologically based pharmacokinetic models in risk assessment. World Health Organization, International Programme on Chemical Safety, Geneva, Switzerland. 2010.



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