# Application of Systematic Review (SR) Principles to Ecotoxicity Studies: ECOTOXicology Knowledgebase

Jennifer H. Olker

US Environmental Protection Agency Center for Computational Toxicology and Exposure Duluth, MN Phone: 218.529.5119 Email: Olker.Jennifer@epa.gov

#### **Conflict of Interest Statement**

The author declares no actual or potential conflicts of interest.

The views expressed in this presentation are those of the author and do not necessarily reflect the views or policies of the US Environmental Protection Agency.

Mention of software packages is not an endorsement by the authors or the US Environmental Protection Agency.

#### **Abbreviations**

- AQUIRE: Aquatic toxicity Information Retrieval Database
- **CASRN**: Chemical Abstracts Service Registry Number
- **DTXSID**: US EPA's Distributed Structure-Searchable Toxicity database (DSSTox) substance identifier
- ECOTOX: ECOTOXicology Knowledgebase (www.epa.gov/ecotox)
- **GUI**: Graphical User Interface
- ITIS: Integrated Taxonomic Information System
- **LC50**: Lethal concentration to 50% of test organisms
- LOEC: Lowest Observed Effect Concentration
- LOEL: Lowest Observed Effect Level

- **NCBI**: National Center for Biotechnology Information
- NOEC: No Observed Effect Concentration
- **NOEL**: No Observed Effect Level
- NR: Not Reported
- **OECD**: Organisation for Economic Co-operation and Development
- **PECO**: Population, Exposure, Comparator/Control, Outcome
- SR: Systematic Review
- **STN:** Scientific and Technical information Network
- **TSN**: Taxonomic Serial Number
- **US EPA**: United State Environmental Protection Agency

#### **Objective and Outline**

- Systematic and transparent methods used for identification and review of ecotoxicity data
- Background and History
- Current Status and Applications
- ECOTOX Pipeline
- Ecotoxicology-specific considerations
- Examples with recently completed chemicals

### **Background and History**

- Ecological risk assessors needed cost-effective methods to locate high quality ecological toxicity data for use in:
  - Prioritizing chemical cleanup at hazardous waste sites
  - Assessment of potential hazards of pollutants through the Clean Air Act, the Clean Water Act, the Federal Insecticide, Fungicide and Rodenticide Act and the Toxic Substances Control Act.
- US EPA developed ecological toxicity databases:
  - 1) Authoritative source of toxicological data
  - 2) Efficient documentation of literature searches and data acquisition
  - 3) Data available for development and validation of in vitro and modeling methods
  - 4) Avoid duplicative efforts for data gathering across programs and agencies

### **Background and History**

Early 1980s:

- AQUatic toxicity Information Retrieval (AQUIRE) database for acute laboratory toxicity tests on <u>aquatic life</u>
  - Expanded to field and chronic exposures in early 1990s
- PHYTOTOX database for toxicity tests with terrestrial plants
- TERRETOX database for toxicity studies with terrestrial wildlife

Late 1990s:

• ECOTOXicology Knowledgebase (ECOTOX): AQUIRE, PHYTOTOX, TERRETOX combined into unified database and available on web

### What is the ECOTOX Knowledgebase?

- Systematic and transparent protocols developed over 30 years:
  - Conduct comprehensive literature searches for toxicity data in the peer-reviewed and grey literature
  - Review applicability of studies following established criteria
  - Extract relevant study and toxicity results into a structured database
- Accessible, structured empirical data from *in vivo* toxicity tests
- Toxicity data provided to US EPA Programs, Regions, and researchers
- Updated quarterly to public website (www.epa.gov/ecotox)

### What is the ECOTOX Knowledgebase?

ECOTOX Kno	wledgebase	Home	Search	Explore	Help	Contact Us
Data last updated Dec 15, 2022	Recent chemicals with full searches completed 6PPD		olyfluoroalkyl Sul	bstances (PFAS)	Total in database 12,714 Chemicals	13,803 Species
See update totals	Cyanotoxins	Thallium			53,763 References	1,134,537 Results

#### About ECOTOX

ECOTOX is a comprehensive Knowledgebase providing single chemical environmental toxicity data on aquatic and terrestrial species.

Read more in: Olker et al. 2022

Learn More



#### **Getting Started**

- Use <u>Search</u> if you know exact parameters or search terms (chemical, species, etc.)
- Use <u>Explore</u> to see what data may be available in ECOTOX (including data plots)
- ECOTOX Quick User Guide (2 pp, 141 K)
- ECOTOX User Guide (95 pp, 672 K)
- ECOTOX Terms Appendix

#### **Other Links**

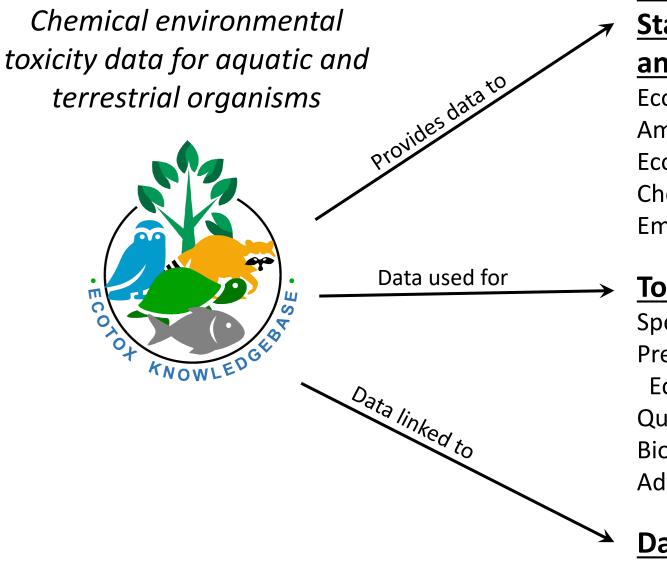
ECOTOX-related documentation and resources.

- Frequent Questions
- Limitations
- Other Tools/Databases
- <u>Recent Additions</u>
- Literature Search Dates

ECOTOX Overview: Olker et al. 2022 https://doi.org/10.1002/etc.5324

#### www.epa.gov/ecotox

### **Applications of ECOTOX**



#### **EPA Program Offices and Regions,**

#### States, Tribes, Other Federal Agencies

#### and International Entities

Ecological Risk Assessments Ambient Water Quality Criteria Ecological Screening Values Chemical Prioritization Emergency Response

#### Tools and Applications

Species Sensitivity Distributions Predicted No-Effect Concentrations and Eco-Thresholds for Toxicological Concern Quantitative Structure–Activity Relationships Bioaccumulation Factor Modeling and Validation Adverse Outcome Pathway Development

#### **Databases/Resources**

### **General SR Process Overview**

- 1. Planning
- 2. Framing the question
- 3. Developing and publishing of the protocol
- 4. Searching for evidence
- 5. Selecting the evidence
- 6. Extracting data
- 7. Assessing the evidence (evaluation)
- 8. Analyzing data
- 9. Interpreting the results
- 10. Report

### **General SR Process Overview**

- 1. Planning
- 2. Framing the question
- 3. Developing and publishing of the protocol
- 4. Searching for evidence
- 5. Selecting the evidence
- 6. Extracting data
- 7. Assessing the evidence (evaluation)
- 8. Analyzing data
- 9. Interpreting the results
- 10. Report

#### **ECOTOX Systematic Protocols**

Parallel general SR Steps

Inform study evaluation

Adapted from Hoffmann et al. 2017

### **General SR Process Overview**

- 1. Planning
- 2. Framing the question
- 3. Developing and publishing of the protocol
- 4. Searching for evidence
- 5. Selecting the evidence
- 6. Extracting data
- 7. Assessing the evidence (evaluation)
- 8. Analyzing data
- 9. Interpreting the results

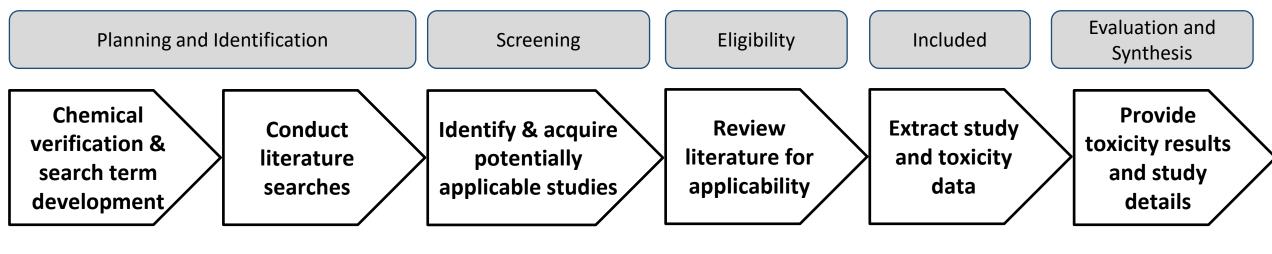
10. Report

**ECOTOX Systematic Protocols** 

Parallel general SR Steps

Inform study evaluation

Application Specific



#### **Chemical-based Search Terms**

- Chemical name and CASRN
- Synonyms, tradenames
- Other relevant forms

#### **Literature Search**

Use chemical-specific search terms to query multiple literature search engines. Title/Abstract Screening

#### Full Text Review

- Established applicability (inclusion) criteria
- Documentation of exclusion reason

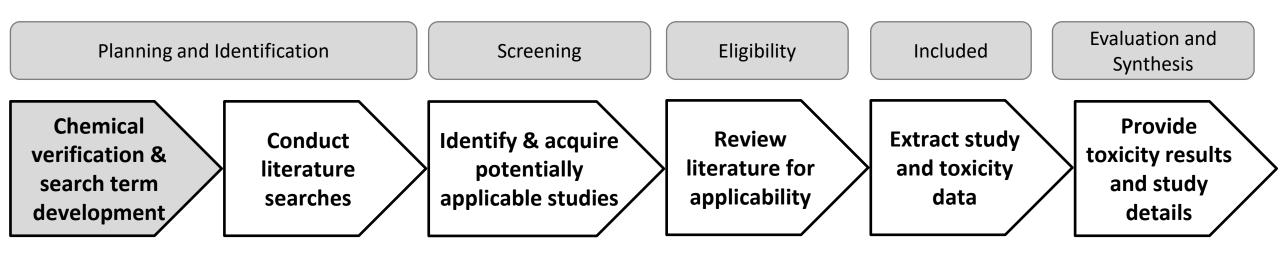
#### Data Extraction

- ECOTOX-specific Controlled Vocabularies
  - Test chemical
  - Test organism
  - Study methods and test conditions
  - Toxicity results
- Updated to public website, with downloadable outputs

#### **Considerations for Ecotoxicology**

- Sources used in literature searches
- Population of interest includes many species
- Diversity in types of effects measured
- Diversity of study design

• Data to support multiple applications

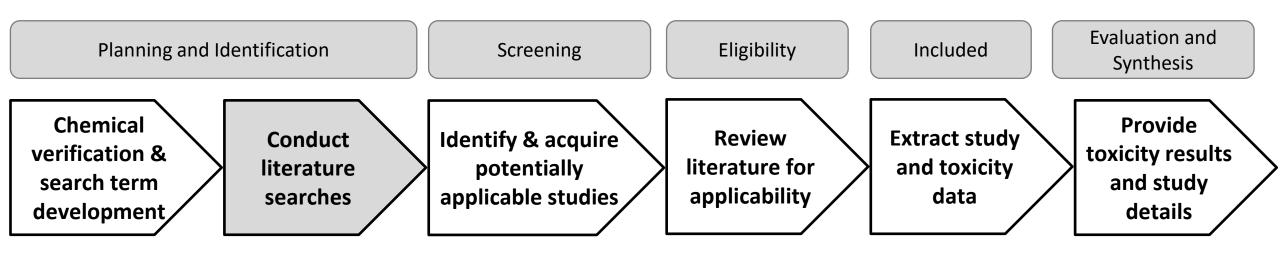


#### **Chemical-based Search Terms\***

- Verify CASRN
- Search various sources for chemical terms
  - STN
  - Pesticide Action Network
  - EPA's Pesticide Fate Database
  - EPA's Chemicals Dashboard
- Synonyms
- Eliminate poor search terms
- Develop search string

Tak(Acilid OR Albrass OR Bexton OR "CP 31393" OR "Kartex A" OR Muharicid OR Niticid OR Propachlor OR Propachlore OR Ramrod OR Satecid OR "US EPA PC Code 019101")

#### \*No ecotoxicology-specific considerations



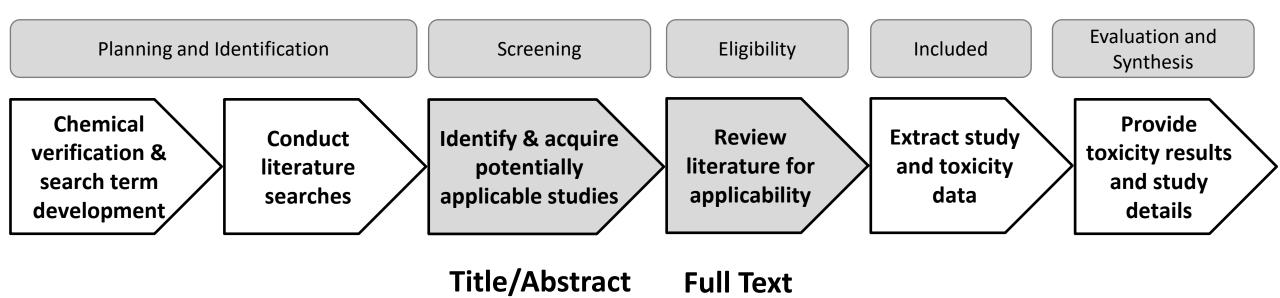
#### **Chemical-based Literature Searches**

#### Search Engines

- 1. Scopus/Science Direct
- 2. ProQuest
- 3. Web of Science
- 4. PubAg/AGRICOLA
- 5. PubMed Toxline/TOXNET
- 6. Dissertation Abstracts

#### \*Considerations for ecotoxicology

- Variety of sources needed to find published studies on aquatic and terrestrial organisms
- Journals with highest applicability rate are often not found through PubMed



Screening Review

- Established applicability (inclusion) criteria which can be expressed as PECO statement
- Documentation of exclusion reason

#### \*Considerations for ecotoxicology

- Population of interest includes many species
- Diversity in types of effects

In		poter applicab	& acquire ntially le studies	Review literature for applicability	
	Key Area	Data Requirement	L		
P (Population)	Species	<ul> <li>Taxonomically verifiable, ecologically-relevant orgation (including cells, organs, gametes, embryos, plant content [NOT bacteria, humans, monkeys, viruses, or yeast]</li> </ul>	uttings)		
E (Exposure)	Chemical	• Single, verifiable chemical toxicants, administered acceptable route.	Single, verifiable chemical toxicants, administered through an acceptable route.		
	Exposure Amount (Concentration)	<ul> <li>Exposure amount is quantified, either as a concent environment when administered via soil or water, or when introduced directly into or on the organism, orally, or topically.</li> </ul>	or as a dosage		
	Exposure Duration	• Known duration from the time of initial exposure to measurement.	o the time of		
С	Control	Must have a control treatment			
(Comparator/ Control)					
0	Effect	<ul> <li>Biological effect measured</li> </ul>			
(Outcome)		Effect concurrent with associated chemical exposu	re		
	Publication Type	<ul> <li>Primary source of the data [NOT a Review]</li> <li>Study must be a full article in English</li> </ul>		Adapted from Olker et al. 2022	

Species Group	Habitat(s)	Num. Species in ECOTOX	Standard Test Species
Fish	Aquatic	1,006	42 species Examples: Rainbow Trout, Zebrafish, Fathead Minnow, Common Carp
Amphibians	Aquatic, Terrestrial	229	7 species Examples: African clawed frog (Xenopus laevis), Northern leopard frog, Bullfrog, Wood frog
Reptiles	Aquatic, Terrestrial	75	none
Birds	Terrestrial	283	11 species Examples: Mallard duck, Japanese quail, Zebra finch
Mammals	Terrestrial, Aquatic*	205	9 species Examples: Norway rat, House mouse, European rabbit

\*currently only *in vitro* studies for aquatic mammals

Species Group	Habitat(s)	Num. Species in ECOTOX	Standard Test Species	Effects and End	points (examples)
Fish	Aquatic	1,006	42 species Examples: Rainbow Trout, Zebrafish, Fathead Minnow, Common Carp	Mortality (i.e., LC50) Time to death Hatch Fecundity Progeny counts Spawning frequency Weight, Length	Limb/body part regeneration Sexual development Vitellogenin
Amphibians	Aquatic, Terrestrial	229	7 species Examples: African clawed frog (Xenopus laevis), Northern leopard frog, Bullfrog, Wood frog	Mortality Time to death Hatch Viability Ovulation rate Weight, Length Condition index	Snout-vent length Metamorphosis Limb/body part regeneration Acetylcholinesterase Testosterone Thyroxine
Reptiles	Aquatic, Terrestrial	75	none	Mortality Time to death Hatch Courtship behavior	Weight, Length Snout-vent length Testosterone 17-beta Estradiol

Species Group	Habitat(s)	Num. Species in ECOTOX	Standard Test Species	Effects and End	points (examples)
Birds	Terrestrial	283	11 species Examples: Mallard duck, Japanese quail, Zebra finch	Mortality Time to death Hatch Viability Progeny counts Eggs per nest Pipped Weight, Length	Condition index Growth rate Fledged/female Food conversion efficiency Cholinesterase Cholesterol
Mammals	Terrestrial, Aquatic	205	9 species Examples: Norway rat, House mouse, European rabbit	Mortality Time to death Viability Progeny counts Resorbed embryos Gestation time	Weight, Length Weight gain Acetylcholinesterase Hemoglobin Testosterone

Species Group	Habitat(s)	Num. Species in ECOTOX	Standard Test Species
Worms	Aquatic, Terrestrial	75	14 species Examples: Earthworm (Eisenia fetida), Nematode (Caenorhabditis elegans), Oligochaete (Lumbriculus variegatus)
Crustaceans	Aquatic, Terrestrial	936	46 species Examples: Daphnia magna, Daphnia pulex, Ceriodaphnia dubia, Hyalella azteca, Opossum shrimp
Molluscs	Aquatic, Terrestrial	678	12 species Examples: Common bay mussel, Virginia oyster, Great pond snail
Insects/ Spiders	Aquatic, Terrestrial	3,590	18 species Examples: Honeybee, Yellow fever mosquito, Midge (Chironomus riparius, Chironomus tentans, Chironomus dilutus)
Other Invertebrates	Aquatic, Terrestrial	648	15 species Examples: Purple-spined sea Urchin, Brown shrimp, Rotifer (Brachionus calyciflorus), Springtail (Folsomia candida)

Species Group	Habitat(s)	Num. Species in ECOTOX	Standard Test Species	Effects and Endpoints (examples)
Worms	Aquatic, Terrestrial	75	14 species Examples: Earthworm (Eisenia fetida), Nematode (Caenorhabditis elegans), Oligochaete (Lumbriculus variegatus)	Mortality Time to death Lifespan Hatch
Crustaceans	Aquatic, Terrestrial	936	46 species Examples: Daphnia magna, Daphnia pulex, Ceriodaphnia dubia, Hyalella azteca, Opossum shrimp	Fecundity Progeny counts Fertile cocoons Gravid Time to first progeny
Molluscs	Aquatic, Terrestrial	678	12 species Examples: Common bay mussel, Virginia oyster, Great pond snail	Weight, Length Weight gain Condition Index Biomass
Insects/ Spiders	Aquatic, Terrestrial	3,590	18 species Examples: Honeybee, Yellow fever mosquito, Midge (Chironomus riparius, Chironomus tentans, Chironomus dilutus)	Growth rate Abnormal growth Oxygen consumption Catalase Respiration
Other Invertebrates	Aquatic, Terrestrial	648	15 species Examples: Purple-spined sea Urchin, Brown shrimp, Rotifer (Brachionus calyciflorus), Springtail (Folsomia candida)	Reactive oxygen species Acetylcholinesterase Lipid Glycogen Calcium content

Species Group	Habitat(s)	Num. Species in ECOTOX	Standard Test Species (examples)
Algae	Aquatic, Terrestrial	1,265	23 species Examples: Green algae (Chlorella vulgaris, Chlamydomonas reinhardtii), Diatoms (Skeletonema costatus, Phaeodactylum tricornutum), Blue-green algae (Microcystis aeruginosa, Anabaena flosaquae)
Vascular Plants	Aquatic, Terrestrial	3,232	30 species Examples: Corn, Soybean, Duckweed, Common onion, Mouse-ear cress
Fungi	Aquatic, Terrestrial	862	none
Moss, Hornworts	Aquatic, Terrestrial	65	none

Species Group	Habitat(s)	Num. Species in ECOTOX	Standard Test Species (examples)	Effects and End	lpoints (examples)
Algae	Aquatic, Terrestrial	1,265	23 species Examples: Green algae (Chlorella vulgaris, Chlamydomonas reinhardtii), Diatoms (Skeletonema costatus, Phaeodactylum tricornutum), Blue-green algae (Microcystis aeruginosa)	Mortality Population abundance and growth rate Biomass Growth rate Photosynthesis	Respiration Germination Chlorophyll Catalase Fluorescence
Vascular Plants	Aquatic, Terrestrial	3,232	30 species Examples: Corn, Soybean, Duckweed, Common onion, Mouse-ear cress	Mortality Germination Seed number Fruit, fruiting Viability Fertility Weight, Height	Length Biomass # of leaves, # of roots Chlorophyll Photosynthesis Respiration Nitrogen content
Fungi	Aquatic, Terrestrial	862	none	Mortality Germination Seed or spore production Viability Weight, Length	Biomass Time to harvest Respiration Aflatoxin B1 Hydrogen peroxide
Moss, Hornworts	Aquatic, Terrestrial	65	none	Mortality Germination Length, Diameter	Chlorophyll Photosynthesis

Inclusion	Criteria	(PECO)
-----------	----------	--------

Identify & acquire potentially applicable studies Review literature for applicability

Image: Problem state in the state in th		Key Area	Data Requirement		
(Exposure)       acceptable route.         Exposure       Exposure acceptable route.         Amount       Exposure amount is quantified, either as a concentration in the environment when administered via soil or water, or as a dosage when introduced directly into or on the organism, via injection, orally, or topically.         Exposure       Known duration from the time of initial exposure to the time of measurement.         C       Control       Must have a control treatment         O       Effect       Biological effect measured         (Outcome)       Effect       Biological effect measured         Publication       Primary source of the data [NOT a Review]         Additionary       Study must he a full actiple in English	P (Population)	Species	(including cells, organs, gametes, embry	os, plant cuttings)	
Amount (Concentration)       environment when administered via soil or water, or as a dosage when introduced directly into or on the organism, via injection, orally, or topically.         Exposure Duration       •       Known duration from the time of initial exposure to the time of measurement.         C (Comparator/ Control)       Control       •       Must have a control treatment         O (Outcome)       Effect       •       Biological effect measured •       Effect concurrent with associated chemical exposure         Publication       •       Primary source of the data [NOT a Review]       Ada		Chemical		ninistered through an	
Duration       measurement.         C       Control       Must have a control treatment         (Comparator/ Control)       Effect       Biological effect measured         O       Effect       Biological effect measured         (Outcome)       Effect concurrent with associated chemical exposure         Publication       Primary source of the data [NOT a Review]         Ada		Amount	environment when administered via soil when introduced directly into or on the o	or water, or as a dosage	
(Comparator/ Control)       •       Biological effect measured         O       Effect       •       Biological effect measured         (Outcome)       •       Effect concurrent with associated chemical exposure         Publication       •       Primary source of the data [NOT a Review]         Twpo       •       Study must be a full article in English		•		exposure to the time of	
(Outcome)       • Effect concurrent with associated chemical exposure         Publication       • Primary source of the data [NOT a Review]         Type       • Study must be a full article in English	(Comparator/	Control	Must have a control treatment		
Publication • Primary source of the data [NOT a Review]	0	Effect	Biological effect measured		
Ada	(Outcome)		• Effect concurrent with associated chemic	cal exposure	
		Publication Type	<ul> <li>Primary source of the data [NOT a Review</li> <li>Study must be a full article in English</li> </ul>	w]	Adapted from Olker et al. 2022

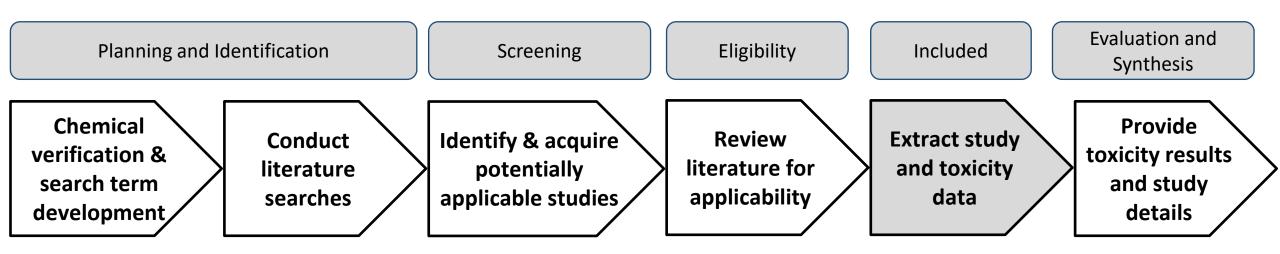
### **Documentation of Exclusion Reasons**

#### All <u>Excluded</u> and <u>Non-Applicable</u> studies are Tagged with the reason for rejection

- Abstract published as an abstract
- Bacteria only test organism is a bacteria
- CAS # Unavailable could not verify/locate chemical CAS Registry number
- Chemical method description of chemical analysis procedures
- Fate only report chemical distribution in media
- Human Health data on human subjects of surrogate animal subjects for human health risk assessment
- Incident reports death of animal by poison, but does not provide concentration/duration of exposure
- Method paper only reports methods for conducting a toxicity test or other aspect of an experiment
- Mixture paper reports results from mixture of chemicals; no single chemical exposure results
- Modeling results of the development of a model; no primary data available

- No Conc the authors report a response in an organism but do not provide conc/dose/app rate
- No Duration duration of exposure is not presented
- No Effect paper does not report observed responses adverse of otherwise
- No Toxicant (ozone, CO2)
- Non-English
- Nutrient in situ chemical tested as nutrient
- PUBL AS duplicate data published elsewhere
- Retracted paper retracted by Journal
- Review primary data published elsewhere
- Sediment only sediment concentration presented
- Survey chemical measured in organism, but lack quantification of exposure (dose/duration)
- Virus virus is only test organism
- Yeast yeast is only test organism

#### \*Allows for re-use of literature search and screening results



#### \*Considerations for ecotoxicology

- Diversity of study methods and types
- Pertinent study details and test conditions vary greatly, requiring many data fields

#### **Data Extraction**

- ECOTOX Data Fields consist of ~90 entities
- ECOTOX-specific Controlled Vocabularies
- Developed from 30+ years reviewing the ecotoxicological literature
- Custom GUI designed for ECOTOX data extraction
  - Computationally-assisted forms constrained to controlled vocabularies

### **Fields Included in ECOTOX Data Extraction**

Category	ECOTOX data fields (examples)	and
Chemical	Chemical identifier (CASRN, DTXSID)	
	Chemical Analysis	
	Chemical Formulation & Grade	
	<ul> <li>Concentration(s)/Dose(s) tested</li> </ul>	
Species	Species identifiers (ITIS TSN, NCBI TaxID, Taxonomy)	
	Life stage, Age, Sex	
	Organism Source	
Study	Experimental design	
Methods &	Control(s)	
Test Conditions	Test location and method	* ECOTO
Conditions	Exposure type, route, and media	ox/help.ct
	Study and exposure duration	definition
	<ul> <li>Physical and Chemical Soil and Water Parameters (e.g., pH, Temperature, Dissolved Oxygen)</li> </ul>	* ECOTO
Test Results	Specific Effect Measured (with higher-level groups)	https://cf
	Calculated Endpoint	ox/help.cl
	<ul> <li>Concentration associated with effect and endpoint</li> </ul>	
	<ul> <li>Response site (e.g., whole organism, specific organ or body part)</li> </ul>	Adapted from
	<ul> <li>Statistical significance and level of response</li> </ul>	Olker et al. 2022

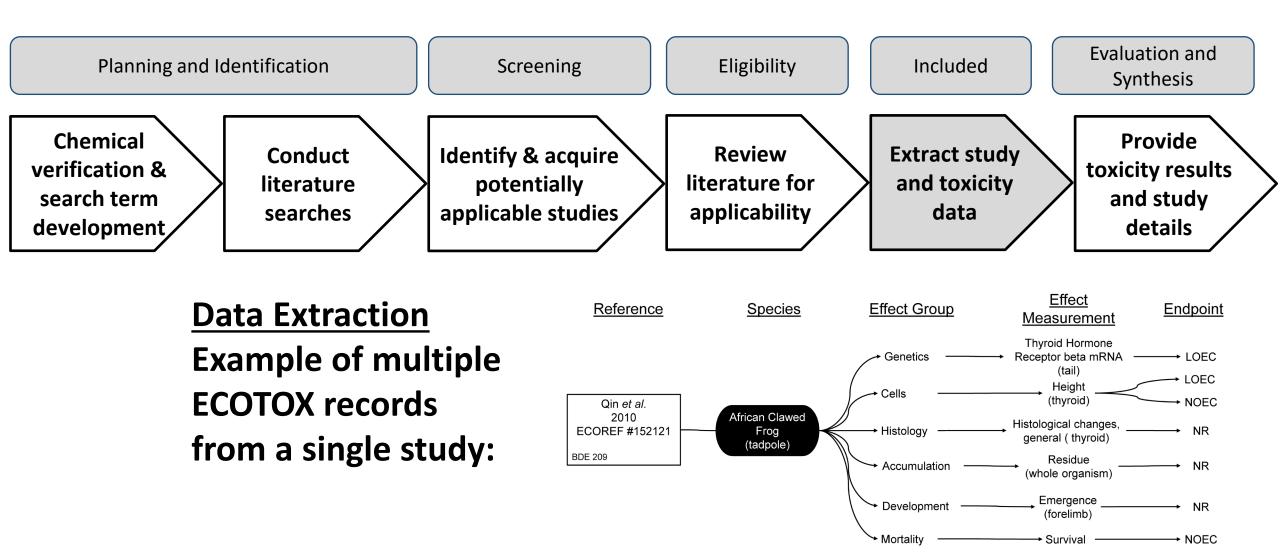
Extract study and toxicity data

#### \* ECOTOX Data Fields

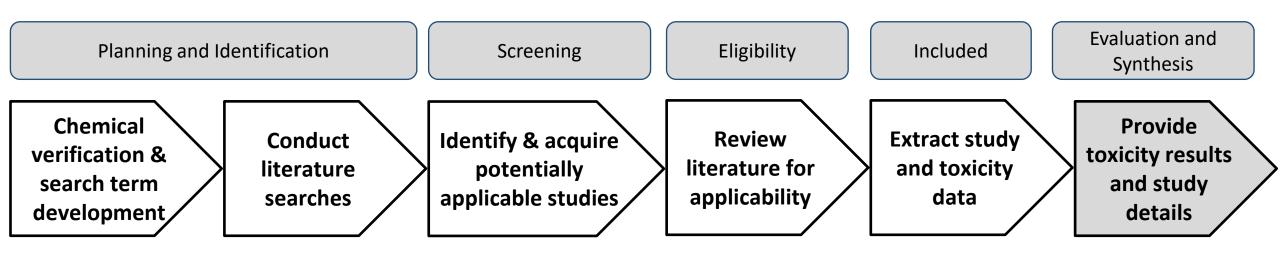
https://cfpub.epa.gov/ecot ox/help.cfm?sub=widefinitions

#### \* ECOTOX Vocabularies:

https://cfpub.epa.gov/ecot ox/help.cfm?sub=termappendix



NOEC = No Observed Effect Level LOEC = Lowest Observed Effect Level NR = Not Reported



#### \*Considerations for ecotoxicology

- Toxicity data from ECOTOX are used for multiple applications with potentially differing criteria for relevance and reliability
- Methodology varies across diversity of taxa

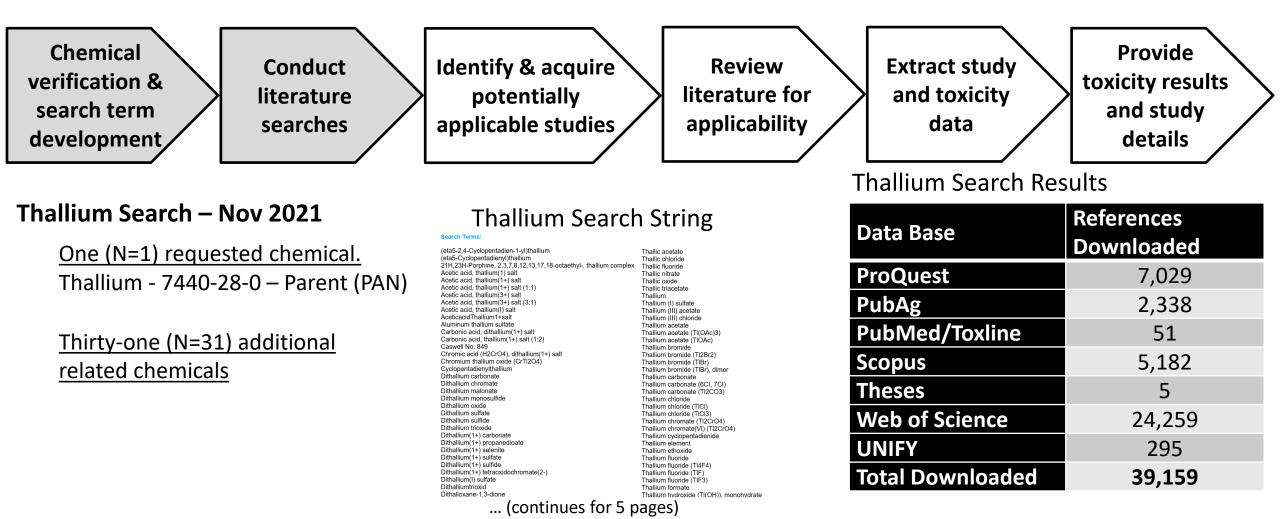
# Supporting Study Evaluation and Data Synthesis

- ECOTOX inclusion criteria overlaps with standard study evaluation questions
- ECOTOX data fields for study design, test conditions, and results inform detailed study evaluation
- Multiple output options for further analysis and synthesis

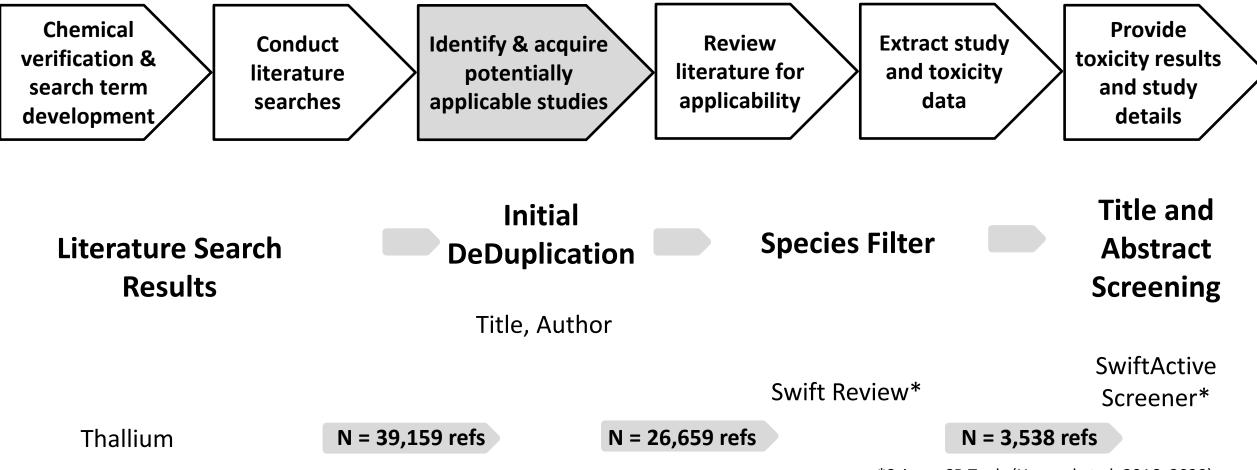
# **Informing Study Evaluation**

Catagory	Solact study ovaluation quastions* with relevant ECOTOX field(s)	deta
Category Chemical	<ul> <li>Select study evaluation questions* with relevant ECOTOX field(s)</li> <li>Is the test substance identified? Required for inclusion in ECOTOX</li> <li>Is the purity of the test substance reported? <u>Chemical Purity</u></li> <li>Were chemical concentrations verified? <u>Chemical Analysis</u> (e.g., nominal versus measu concentrations)</li> </ul>	red
Species	<ul> <li>Is the species given? Verifiable species (<u>Scientific Name</u>, etc.) required for inclusion in ECOTOX</li> <li>Are the organisms well described? <u>Organism Source</u>, <u>Lifestage</u>, <u>Age</u>, <u>Gender</u>, <u>Initial</u> an <u>Final Weight</u></li> </ul>	
Study Methods & Test Conditions	<ul> <li>Are appropriate controls performed? Control required for inclusion in ECOTOX, type described in <u>Control</u></li> <li>Is a guideline method (e.g., OECD) used? <u>Test Method</u></li> <li>Are the experimental conditions appropriate and acceptable for the test substance and organism? <u>Test Method</u>, <u>Media Type</u>, <u>Test Location</u>, <u>Experimental Design</u>, Physical and Chemical Soil and Water Parameters (e.g., <u>pH</u>, <u>Temperature</u>, <u>Dissolved Oxygen</u>)</li> </ul>	
Test Results	<ul> <li>Are the reported effects and endpoints appropriate for the purpose, test substance and organism? <u>Effect Measurement</u>, <u>Endpoint</u></li> <li>Is the response/effect statistically significant? <u>Statistical Significance</u>, <u>Significance Leve</u></li> </ul>	
Adapted from Olker et al. 2022	*based on published evaluation guidelines (Kaltenhaus et al. 2017; Moermond et al. 2016; Schneider et al. 200	

**Problem Statement**: Updated toxicity data are needed to develop an ecological screening value for Thallium, a naturally occurring elemental metal which also has industrial sources.

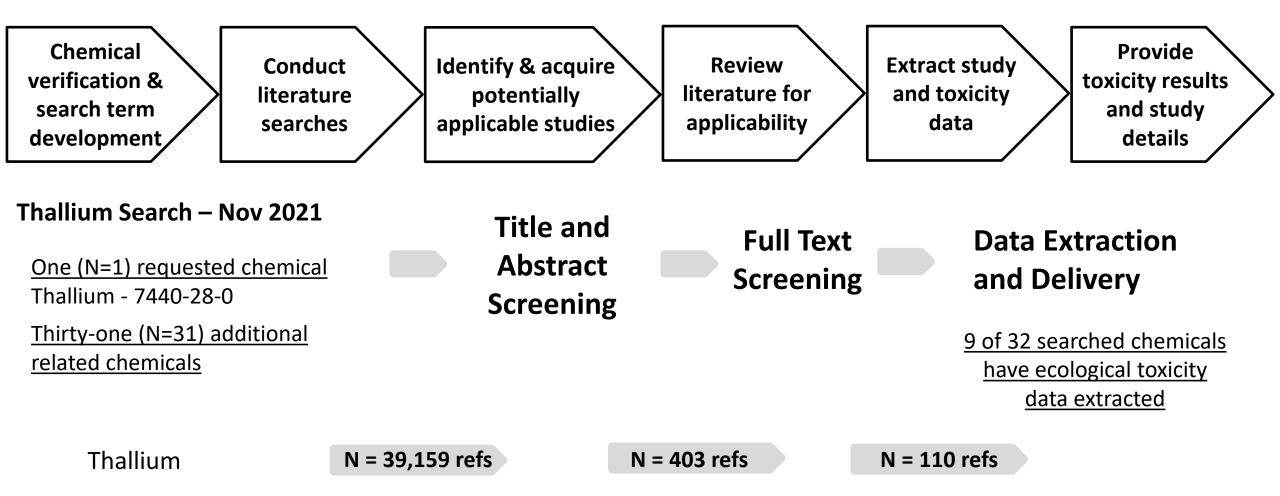


**Problem Statement**: Updated toxicity data are needed to develop an ecological screening value for Thallium, a naturally occurring elemental metal which also has industrial sources.

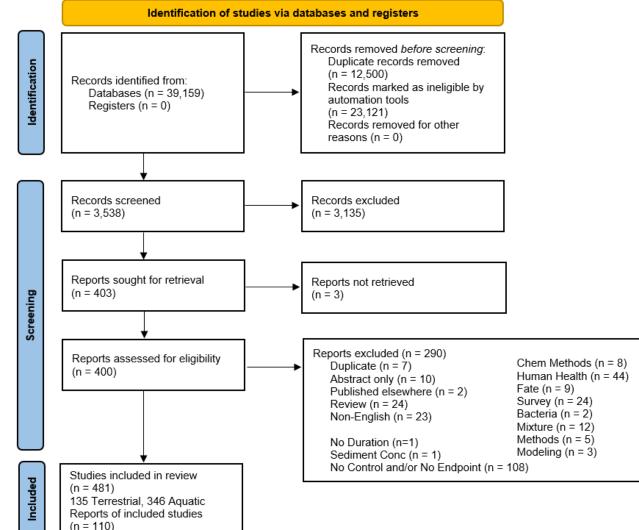


\*Sciome SR Tools (Howard et al. 2016, 2020)

**Problem Statement**: Updated toxicity data are needed to develop an ecological screening value for Thallium, a naturally occurring elemental metal which also has industrial sources.



#### **Study Flow Diagram**

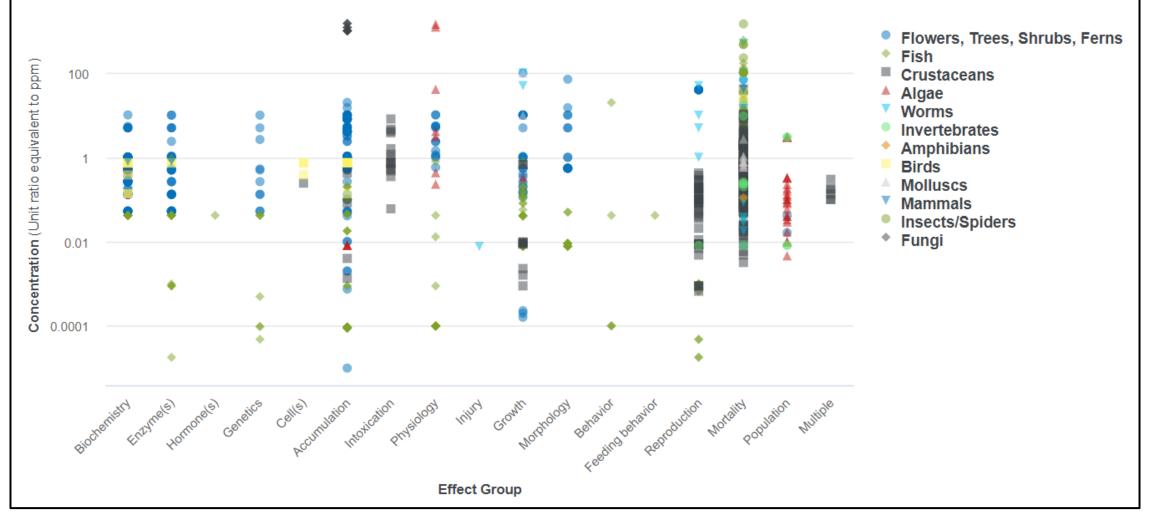


#### Following template from Page et al. 2021

<b>ECOTOX</b> Knowledge	Home Search	Explore He	lp	Cont	act Us		
८ Explore 👌 Chemicals Tha							
Aquatic Terrestrial Group Summary Records Plot View					🛛 🗹 Ser	nd Query Filters to Search (	Ð
Query Filters       9 Chemicals         Select one or more or more or decent filter to reduce the records.       of each filter to Chemicals are ordered by CAS Number.					🗹 Export C	SV	
Chemicals (9) Showing all 9 chemicals from 563688 to 13746980		n <b>563688</b> to <b>13746980</b>					
		CHEMICAL NAME	RECORDS	PUBLICATIONS	YEAR MIN	YEAR MAX	
Species Group (12)	type to filter						
All	563688	Thallium (I) acetate	198	12	1975	2021	>
Class (25)	2570630	Thallium (III) acetate	2	1	2005	2005	>
Order (57) All V	6533739	Thallium carbonate (Tl2CO3) Chemicals Dashboard	15	1	2002	2002	>

https://cfpub.epa.gov/ecotox/explore.cfm?cgid=41

#### **ECOTOX Explore Plot view**



https://cfpub.epa.gov/ecotox/explore.cfm?cgid=41

### Example: Thallium Export of records – Mortality data for Terrestrial Invertebrates

										Chemical	
		Chemical			Species Common	Organism	Media	Test		Analysis	Number of
CAS Number 💌		Grade 💌		• • • • • • • • • • • • • • • • • • •			Туре 💌			Method	Doses 💌
7440280	Thallium		NR	Caenorhabditis elegans	Nematode	Adult	Culture	Lab	Culture medium	Unmeasured	
7440280	Thallium		NR	Caenorhabditis elegans	Nematode	Adult	Culture	Lab	Culture medium	Unmeasured	
7440280	Thallium		NR	Caenorhabditis elegans	Nematode	Adult	Culture	Lab	Culture medium	Unmeasured	
7440280	Thallium		NR	Caenorhabditis elegans	Nematode	Adult	Culture	Lab	Culture medium	Unmeasured	
6533739	Carbonic acid, Thallium(1+) salt (1:	2)	NR	Arianta arbustorum	Copse Snail	Egg	Artificial s	Lab	Culture medium	Unmeasured	6
6533739	Carbonic acid, Thallium(1+) salt (1:	2)	NR	Eisenia fetida	Earthworm	Mature	Artificial s	Lab	Culture medium	Unmeasured	7
6533739	Carbonic acid, Thallium(1+) salt (1:	2)	NR	Eisenia fetida	Earthworm	Mature	Artificial s	Lab	Culture medium	Unmeasured	7
563688	Acetic acid, Thallium(1+) salt (1:1)		>99.995	Caenorhabditis elegans	Nematode	Larva	Aqueous	Lab	Aqueous	Unmeasured	8
563688	Acetic acid, Thallium(1+) salt (1:1)		>99.995	Caenorhabditis elegans	Nematode	Larva	Aqueous	Lab	Aqueous	Unmeasured	8
563688	Acetic acid, Thallium(1+) salt (1:1)		>99.995	Caenorhabditis elegans	Nematode	Larva	Aqueous		Aqueous	Unmeasured	8
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva		Lab	Culture medium	Unmeasured	3
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode		Culture	Lab	Culture medium	Unmeasured	2
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva	Culture	Lab	Culture medium	Unmeasured	8
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva	Culture	Lab	Culture medium	Unmeasured	8
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva	Culture	Lab	Culture medium	Unmeasured	3
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva	Culture	Lab	Culture medium	Unmeasured	3
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva	Culture	Lab	Culture medium	Unmeasured	3
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva	Culture	Lab	Culture medium	Unmeasured	2
	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva	Culture	Lab	Culture medium	Unmeasured	
				-							3
563688	Acetic acid, Thallium(1+) salt (1:1)		NR	Caenorhabditis elegans	Nematode	Larva	Culture	Lab	Culture medium	Unmeasured	3

Reference	Chemical	Species	Effect and Endpoint
Williams and Dusenbery 1990	Thallium (CASRN 7440-28-0)	Nematode ( <i>Caenorhabditis elegans</i> )	1-day LC50 >200,000 ug/L 2-day LC50 = 194,000 ug/L 3-day LC50 = 172,000 ug/L 4-day LC50 = 123000 ug/L
Heim et al. 2002	Thallium carbonate (Tl2CO3) (CASRN 6533-73-9)	Copse Snail ( <i>Arianta arbustorum</i> )	Hatch LOEL = 1 mg/kg soil
Heim et al. 2002	Thallium carbonate (Tl2CO3) (CASRN 6533-73-9)	Earthworm ( <i>Eisenia fetida</i> )	28-day Mortality NOEL = 500 mg/kg soil LOEL = 500 mg/kg soil
Varao et al. 2021	<b>Thallium (I) acetate</b> (CASRN 563-68-8)	Nematode ( <i>Caenorhabditis elegans</i> )	25-hr Mortality LC50 = 59.5 mg/L NOEL = 50 μM LOEL = 100 μM
Hurtado- Diaz et al. 2020	Thallium (I) acetate (CASRN 563-68-8)	Nematode ( <i>Caenorhabditis elegans</i> )	Mortality 1-hr LOEL = $15 \mu$ M 24-hr LOEL = $2.5 \mu$ M 16-day 100% lethality = $5 \mu$ M

Reference	Chemical	Species	Effect and Endpoint	Control
Williams and Dusenbery 1990	Thallium (CASRN 7440-28-0) Nominal concentrations	Nematode ( <i>Caenorhabditis elegans</i> )	1-day LC50 >200,000 ug/L 2-day LC50 = 194,000 ug/L 3-day LC50 = 172,000 ug/L 4-day LC50 = 123000 ug/L	~
Heim et al. 2002	Thallium carbonate (Tl2CO3) (CASRN 6533-73-9) Nominal concentrations	Copse Snail ( <i>Arianta arbustorum</i> )	Hatch LOEL = 1 mg/kg soil	Concurrent
Heim et al. 2002	Thallium carbonate (Tl2CO3) (CASRN 6533-73-9) Nominal concentrations	Earthworm ( <i>Eisenia fetida</i> )	28-day Mortality NOEL = 500 mg/kg soil LOEL = 500 mg/kg soil	Concurrent
Varao et al. 2021	Thallium (I) acetate (CASRN 563-68-8) >99.995% Purity Nominal concentrations	Nematode ( <i>Caenorhabditis elegans</i> )	25-hr Mortality LC50 = 59.5 mg/L NOEL = 50 μM LOEL = 100 μM	Concurrent
Hurtado- Diaz et al. 2020	Thallium (I) acetate (CASRN 563-68-8) Nominal concentrations	Nematode ( <i>Caenorhabditis elegans</i> )	Mortality 1-hr LOEL = $15 \mu$ M 24-hr LOEL = $2.5 \mu$ M 16-day 100% lethality = $5 \mu$ M	Concurrent

### **Challenges and Limitations**

- Immensity of ecotoxicological literature
- Missing information due to insufficient reporting in publications
- Mixture data not included
- Evolving toxicology testing requires evolution of ECOTOX while continuing to support traditional risk assessment
- Collaboration and alignment with human health hazard assessment methodologies

#### **Summary and Conclusion**

- ECOTOXicology Knowledgebase was developed for and continues to support chemical risk assessment and research
- Protocols and vocabulary evolved over 30+ year history

- Methods consistent with much of current systematic review guidelines
- Must consider ecotoxicological-specific aspects for species, effects, and sources of information

### **ECOTOX Protocols Parallel SR**

	Planning Framing the question	ECOTOX Standard Operating Procedures - Updated and reviewed annually				
2.	Framing the question	Established ECOTOX Inclusion Criteria				
3.	3. Developing and publishing of the protocol					
4.	Searching for evidence	Chemical-based literature searches				
5.	Selecting the evidence	Study identification and screening - Documented and reproducible				
6.	Extracting data	ECOTOX-specific data extraction application				
7.	Assessing the evidence (evaluation	-Structured data to support data evaluation and synthesis				
8.	Analyzing data	Application Specific				
9.	Interpreting the results					
10.	Report					

#### References

- Heim, M., O. Wappelhorst, and B. Markert. (2002). Thallium in Terrestrial Environments Occurrence and Effects. Ecotoxicology11(5): 369-377.
- Hoffmann, S., de Vries, R., Stephens, M. L., Beck, N. B., Dirven, H., Fowle, J. R., 3rd, Goodman, J. E., Hartung, T., Kimber, I., Lalu, M. M., Thayer, K., Whaley, P., Wikoff, D., & Tsaioun, K. (2017). A primer on systematic reviews in toxicology. Archives of Toxicology, 91(7), 2551–2575. <u>https://doi.org/10.1007/s00204-017-1980-3</u>
- Howard BE, Phillips J, Miller K, Tandon A, Mav D, Shah MR, Holmgren S, Pelch KE, Walker V, Rooney AA, Macleod M, Shah R, Thayer, K (2016). SWIFT-Review: A Text-mining Workbench for Systematic Review. Systematic Reviews, 5:87.
- Howard BE, Phillips J, Tandon A, Maharana A, Elmore R, Mav D, Sedykh A, Thayer K, Merrick A, Walker V, Rooney A, Shah RR (2020). "SWIFT-Active Screener: accelerated document screening through Active Learning and integrated recall estimation." Environment International 138 May 2020, 105623. doi: 10.1016/j.envint.2020.105623.
- Hurtado-Diaz, M.E., R. Estrada-Valencia, E. Rangel-Lopez, M. Maya-Lopez, A. Colonnello, S. Galvan-Arzate, S.V. Verstraet. (2020). Thallium Toxicity in Caenorhabditis elegans: Involvement of the SKN-1 Pathway and Protection by S-Allylcysteine. Neurotox. Res. 38(2): 287-298.
- Kaltenhäuser, J., Kneuer, C., Marx-Stoelting, P., Niemann, L., Schubert, J., Stein, B., & Solecki, R. (2017). Relevance and reliability of experimental data in human health risk assessment of pesticides. *Regulatory Toxicology and Pharmacology*, *88*, 227–237. <u>https://doi.org/10.1016/j.yrtph.2017.06.010</u>
- Moermond, C. T., Kase, R., Korkaric, M., & Ågerstrand, M. (2016). CRED: Criteria for reporting and evaluating ecotoxicity data. *Environmental Toxicology & Chemistry*, 35(5), 1297–1309. <u>https://doi.org/10.1002/etc.3259</u>
- Olker, J. H., Elonen, C. M., Pilli, A., Anderson, A., Kinziger, B., Erickson, S., Skopinski, M., Pomplun, A., LaLone, C. A., Russom, C. L., & Hoff, D. (2022). The ECOTOXicology Knowledgebase: A Curated Database of Ecologically Relevant Toxicity Tests to Support Environmental Research and Risk Assessment. *Environmental toxicology and chemistry*, 41(6), 1520–1539. <u>https://doi.org/10.1002/etc.5324</u>
- Page M J, McKenzie J E, Bossuyt P M, Boutron I, Hoffmann T C, Mulrow C D et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews BMJ 2021; 372:n71 doi:10.1136/bmj.n71 <a href="https://www.bmj.com/content/372/bmj.n71">https://www.bmj.com/content/372/bmj.n71</a>
- Qin,X., X. Xia, Z. Yang, S. Yan, Y. Zhao, R. Wei, Y. Li, M. Tian, X. Zhao, Z. Qin, and X. Xu. (2010). *Thyroid Disruption by Technical Decabromodiphenyl Ether (DE-83R) at Low Concentrations in Xenopus laevis*. J. Environ. Sci. (China)22(5): 744-751.
- Schneider, K., Schwarz, M., Burkholder, I., Kopp-Schneider, A., Edler, L., Kinsner-Ovaskainen, A., Hartung, T., & Hoffmann, S. (2009). "ToxRTool", a new tool to assess the reliability of toxicological data. *Toxicology Letters*, *189*(2), 138–144. <u>https://doi.org/10.1016/j.toxlet.2009.05.013</u>
- Varao, A.M., J.D.S. Silva, L.O. Amaral, L.L.P. Aleixo, A. Onduras, C.S. Santos, L.P.D. Silva, D.E. Ribeiro, J.L.L. Filho. (2021). Toxic Effects of Thallium Acetate by Acute Exposure to the Nematode C. elegans. J. Trace Elem. Med. Biol.68:10 p.
- Williams, P.L., and D.B. Dusenbery. (1990). Aquatic Toxicity Testing Using the Nematode, Caenorhabditis elegans. Environ. Toxicol. Chem.9(10): 1285-1290.